

# Interleukins in Atherosclerosis: Molecular Pathways and Therapeutic Potential

JAN H. VON DER THÜSEN, JOHAN KUIPER, THEO J. C. VAN BERKEL, AND ERIK A. L. BIESSEN

*Division of Biopharmaceutics, Leiden/Amsterdam Center for Drug Research, Leiden, The Netherlands*

Abstract .....	133
I. Introduction.....	134
II. Interleukin families in atherosclerosis .....	135
A. Interleukin-1.....	136
B. Interleukin-2.....	137
C. The gp130 family.....	139
D. Granulocyte macrophage-colony-stimulating factor.....	140
E. Interleukin-10.....	141
F. Chemokines.....	141
G. Interleukin-17.....	142
III. Modulation of cytokine function as a therapeutic strategy for atherosclerosis .....	142
A. Inhibition of expression/translation of interleukins and their receptors.....	145
B. Inhibition of interleukin processing.....	147
C. Neutralization of proinflammatory interleukins.....	147
D. Interleukin receptor antagonists .....	149
E. Up-regulation of anti-inflammatory interleukins.....	150
F. Inhibition of interleukin signaling.....	152
G. Inhibition of interleukin-induced gene expression.....	153
IV. Discussion .....	154
References .....	156

**Abstract**—Interleukins are considered to be key players in the chronic vascular inflammatory response that is typical of atherosclerosis. Thus, the expression of proinflammatory interleukins and their receptors has been demonstrated in atheromatous tissue, and the serum levels of several of these cytokines have been found to be positively correlated with (coronary) arterial disease and its sequelae. In vitro studies have confirmed the involvement of various interleukins in pro-atherogenic processes, such as the up-regulation of adhesion molecules on endothelial cells, the activation of macrophages, and smooth muscle cell proliferation. Furthermore, studies in mice deficient or transgenic for specific interleukins have demonstrated that, whereas some interleukins are indeed intrinsically pro-atherogenic, others may have anti-atherogenic

qualities. As the roles of individual interleukins in atherosclerosis are being uncovered, novel anti-atherogenic therapies, aimed at the modulation of interleukin function, are being explored. Several approaches have produced promising results in this respect, including the transfer of anti-inflammatory interleukins and the administration of decoys and antibodies directed against proinflammatory interleukins. The chronic nature of the disease and the generally pleiotropic effects of interleukins, however, will demand high specificity of action and/or effective targeting to prevent the emergence of adverse side effects with such treatments. This may prove to be the real challenge for the development of interleukin-based anti-atherosclerotic therapies, once the mediators and their targets have been delineated.

Address correspondence to: Jan H. von der Thüsen, Division of Biopharmaceutics, Leiden/Amsterdam Center for Drug Research, Gorlaeus Laboratories, Leiden University, Einsteinweg 55, P.O. Box 9502, 2300 RA Leiden, The Netherlands. E-mail: thuesen@lacdr.leidenuniv.nl

Article, publication date, and citation information can be found at <http://pharmrev.aspetjournals.org>.

DOI: 10.1124/pr.55.1.5.

## I. Introduction

Atherosclerosis remains, despite a recent decline, the most common cause of death in the Western world. The disease course of atherosclerosis is characterized by its chronicity, and progression in its initial stages is particularly insidious. Chronic inflammation is the pathological hallmark of atherosclerosis (Ross, 1986, 1993a, 1999), and inflammatory processes are instrumental in all stages of this disease. Even prior to the development of detectable intimal lesions, the expression pattern of the endothelium has been shown to be inflammatory in nature, conforming to the response-to-injury hypothesis as first postulated by the late Russell Ross (Ross and Glomset, 1973). Thus, in lesion-prone sites of the arterial tree, the endothelial expression of adhesion molecules is up-regulated, reflecting endothelial dysfunction secondary to unfavorable hemorheology (Nakashima et al., 1998) and/or hypercholesterolemia (Rosenfeld, 1991; Li et al., 1993; Sakai et al., 1997; Nakashima et al., 1998). In turn, this leads to the adhesion, extravasation, and intimal accumulation of circulating leukocytes (Nageh et al., 1997; Gerszten et al., 1998; Nakashima et al., 1998; Ramos et al., 1999; Dong et al., 2000), and thus to the development of the earliest detectable lesion—the fatty streak—which consists solely of lipid-laden macrophages and T lymphocytes (Stary et al., 1994). These cell types are also present in more advanced plaques, in addition to smooth muscle cells and extracellular lipid and matrix deposits (Stary et al., 1994, 1995). The cellular constituents of the atherosclerotic lesion are thought to participate actively in the propagation of inflammation and, eventually, plaque destabilization (Ross, 1999; Sukhova et al., 1999). As well as contributing to the bulk of the lesion, plaque cells are involved in the production and degradation of extracellular matrix and contribute toward the formation of a necrotic lesion core by the elaboration of toxic mediators. These cellular functions are partly autonomous but to a large extent subject to autocrine and paracrine control mechanisms. A plethora of mediators has been shown to be involved in intercellular signaling in atheromatous tissue, including small molecules such as nitric oxide (Ignarro et al., 1999; Li and Forstermann, 2000), lipid mediators such as eicosanoids and sterols (Hajjar and Pomerantz, 1992; Edwards and Ericsson, 1999; Schnaper et al., 2000), and polypeptides such as cytokines (Frostegard et al., 1999; Meager, 1999).

Whereas fatty streaks are now known to develop even in utero under the influence of maternal hypercholesterolemia (Napoli et al., 1997), plaques rarely give rise to symptoms before the sixth or seventh decade of life. If primary prevention is to be the cardinal aim, the protracted nature of lesion development will necessitate a therapeutic strategy with a comparably prolonged duration of effectivity. In conjunction with the as yet perfunctory levels of prognostic accuracy for the identification of

patients at risk of symptomatic atherosclerosis, this poses stringent demands with respect to the tolerability of any preventive intervention, including the use of immunomodulatory therapies.

The rate of atherogenesis largely depends on the level of exposure to major risk factors, including a positive family history, hypercholesterolemia, smoking, diabetes mellitus, and hypertension. Although the avoidance of risk factors undoubtedly constitutes the most rewarding approach to the prevention of atherosclerosis, it has thus far been frustrated by inadequate patient compliance and the influence of genetic factors in determining an individual's predisposition to atherosclerosis. This has led to the introduction of a variety of pharmacological interventions, including the widespread use of an extremely effective class of lipid-lowering drugs: the HMG-CoA reductase inhibitors, or so-called *statins* (Braunstein et al., 2001). Despite recent concerns regarding the induction of rhabdomyolysis, a rare and potentially lethal side effect of statin usage, these drugs continue to be the mainstay of most cholesterol-lowering regimens. In several clinical prevention trials (e.g., CARE; Ridker et al., 1998), statins have also been found to exert additional, lipid-independent, anti-inflammatory effects. These may contribute significantly to their anti-atherogenic properties, and this has indeed been corroborated in recent animal studies (Williams et al., 1998). Indeed, immunomodulation could be an attractive paradigm for the development of therapeutic alternatives to statins in atherosclerosis prevention. This may be of particular benefit to those whose lipid levels are (partially) unresponsive to statin therapy; as in a substantial number of patients in the U.S. National Cholesterol Education Program, LDL<sup>1</sup> cholesterol levels cannot be attained by statin monotherapy alone (Brown et al., 1998).

To enable rational drug design aimed at immunomodulation in atherosclerosis, the pivotal inflammatory processes involved in this disease need to be delineated. In this regard, extensive efforts have been devoted to outlining the involvement of cytokines, because these

<sup>1</sup> Abbreviations: LDL, low density lipoprotein; TNF, tumor necrosis factor; TNFR, TNF receptor; IL, interleukin; IFN $\gamma$ , interferon  $\gamma$ ; Th, T helper cell; MCP-1, monocyte chemoattractant protein-1; RANTES, regulated on activation normal T cell expressed and secreted; MIP-1, macrophage inflammatory protein-1; ICE, IL-1 $\beta$ -converting enzyme; MAPK, mitogen-activated protein kinase; NF- $\kappa$ B, nuclear factor- $\kappa$ B; AP-1, activating protein-1; ICAM-1, intercellular adhesion molecule 1; VCAM-1, vascular cell adhesion molecule-1; SMC, smooth muscle cell; MMP, matrix metalloproteinase; SOCS, suppressor of cytokine signaling; LPS, lipopolysaccharide; GM-CSF, granulocyte-macrophage colony-stimulating factor; G-CSF, granulocyte-colony-stimulating factor; Jak, Janus kinase; STAT, signal transducer and activator of transcription; ODN, oligodeoxynucleotide; PKC, protein kinase C; PDGF, platelet-derived growth factor; TGF $\beta$ , transforming growth factor  $\beta$ ; AAV, adeno-associated virus; IKK, I $\kappa$ B kinase; iNOS, inducible nitric-oxide synthase; WIN 67694, Z-Val-Ala-Asp-CH<sub>2</sub>O(CO)[2,6-CI2]Ph; SB 203580, 4-(4-fluorophenyl)-2-(4-methylsulfinylphenyl)-5-(4-pyridyl)-1H-imidazole; VE 13,045, carbobenzyloxy-Val-Ala-Asp(O-et)-CH<sub>2</sub>O-dichlorobenzoate.

cell-regulatory proteins are known to be key players in the initiation and control of inflammation in general. The term “cytokine” was first coined in the 1970s and encompasses a large number of (glyco)proteins involved in cell-to-cell signaling. Cytokines are conventionally classified by assignment to one of six families: interleukins, the tumor necrosis factor family, interferons, colony-stimulating factors, growth factors, and chemokines (Henderson and Higgs, 2000). Considerable overlap between these families exists, however, and alternative methods of subdivision have been suggested. Depending on the aim of classification it may be preferable to distinguish cytokines with an essentially proinflammatory mode of action [including tumor necrosis factor (TNF), interleukin-12 (IL-12), IL-18, and interferon  $\gamma$  (IFN $\gamma$ )] from those with largely anti-inflammatory properties (including IL-4, IL-10, IL-13, and the endogenous IL-1 receptor antagonist, IL-1ra) or T helper cell type I (Th1; including IL-2, IFN $\gamma$ , and TNF) from T helper cell type II (Th2; including IL-3, IL-4, IL-5, IL-6, IL-10, and IL-13) cytokines. Alternatively, it may be desirable to identify cytokines according to their major function, such as those effecting chemoattraction [chemokines, including monocyte chemoattractant protein-1 (MCP-1), RANTES, macrophage inflammatory protein-1 (MIP-1), IL-8, and IL-16] or on the basis of receptor sequence homology (e.g., those employing the gp130 signal transduction protein, such as IL-6, IL-11, IL-12, oncostatin M, and cardiotrophin-1). Nonetheless, a substantial degree of pleiotropism in cytokine effector functions makes most of these subdivisions somewhat arbitrary.

Members of each conventional cytokine family have been found to be involved in atherosclerosis, and all cell types present in the atherosclerotic plaque are capable of producing and responding to cytokine mediators. It is conceivable, therefore, that intervention in cytokine signaling could provide effective prevention and/or treatment of atherosclerosis, and proof-of-principle data to this effect have been obtained in a variety of *in vitro* and *in vivo* studies, although this has not yet yielded clinically applicable protocols. In this review, we shall focus mainly on interleukins in our aim to outline the results that have been achieved to date in delineating the pathophysiological role and the therapeutic potential of cytokines in atherosclerosis. In addition, we shall discuss the potential of the modulation of cytokine activity as a therapeutic approach to the primary and secondary prevention of atherosclerosis. Following an overview of the roles ascribed to a variety of interleukins in the pathogenesis of atherosclerosis, we shall describe recent progress in this field and perceived future opportunities.

## II. Interleukin Families in Atherosclerosis

By definition, interleukins are produced mainly by leukocytes and exert their effects mainly *on* leukocytes. Endothelial cells and smooth muscle cells, however, also

express a variety of interleukins and/or their respective receptors, and their effects in atherogenesis are therefore by no means restricted to macrophages and T cells. Thus far, more than 30 major members of the interleukin family have been identified, and the majority of these have been shown to play a role in atherogenesis. As applies to cytokines in general, it is possible to subdivide the interleukins into families according to the homology of their amino acid sequences or the homology of the receptor complexes to which they bind (Fig. 1). Of these subgroups, the gp130 receptor family comprises principally pro-atherogenic interleukins, but most other families have both anti- and pro-atherogenic members (e.g., IL-1 family, IL-2 family, and  $\gamma$ c receptor family). It has not proved feasible to pinpoint an interleukin that acts as the cardinal culprit in the atherosclerotic process. On the contrary, it seems rather more likely that the delicate balance between pro- and anti-inflammatory signals that generally serves to keep inflammation in check, goes awry in atherosclerosis, leading to a self-perpetuating mechanism of lesion formation (Ross, 1993b; Tedgui and Mallat, 2001). Considering the extensive interplay of soluble mediators in the atherosclerotic plaque, however, it may prove possible to devise an anti-atherosclerotic therapy aimed at modifying the effect of a single interleukin, provided that due attention is paid to the mechanisms of redundancy, which have been shown to exist in cytokine signaling. In doing so, candidate interleukins cannot be identified solely by virtue of a demonstrated systemic or local modulation of their expression in the course of atherogenesis. On the contrary, it is of paramount importance to determine whether cytokine responses that have been observed in relation to the development of atherosclerosis are *compensatory* to, *contributory* to, or merely *associated* with this disease. Making this distinction will require well designed intervention studies in animal models, in which the effect of attenuation or administration of a particular interleukin can be evaluated. The currently favored approach entails the up- or down-regulation of interleukin expression in atherosclerosis-prone mouse strains by means of gene insertion (“transgenics”) or gene deletion (“knockouts”), respectively. Administration of an interleukin or its ablation by specific antibodies/antagonists, however, can also provide valuable data regarding its role in atherogenesis. When pertinent, the results of such studies will be discussed in the next section.

Since the effects exerted by cytokines may differ significantly depending on their local environment, it will also be necessary to distinguish between the role of systemic and local variations in cytokine levels. This type of information could in the future be derived from cell- or organ-specific gene overexpression through the use of specific promoters and gene deletion by means of the cre-lox system (Perkins, 2002) or by comparison of

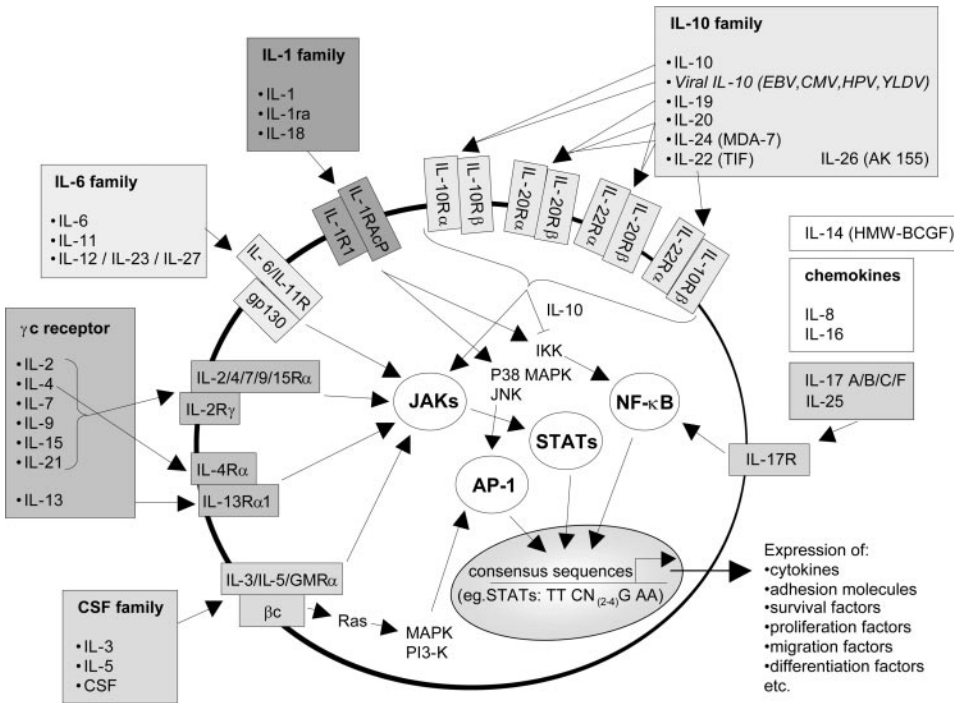


FIG 1. Schematic representation of the receptor specificity and mechanism of action of interleukin families thought to be involved in atherogenesis. Most receptors have been found to consist of heterodimeric complexes, frequently incorporating an interleukin-specific chain in addition to a common chain that is shared by the interleukin family members (including IL-2R $\gamma$ ,  $\beta$ c, and gp130). Receptor activation initiates intracellular phosphorylation cascades that are mediated by kinases (including p38 MAPK, c-Jun N-terminal kinase, and JAKs), resulting in the activation and/or nuclear translocation of transcription factors (including AP-1, STATs, NF- $\kappa$ B). Binding of these factors to DNA consensus sequences, in conjunction with the required cofactors, effects the expression of specific patterns of pro- and/or anti-inflammatory mediators.

the effects of local and systemic administration of cytokines.

### A. Interleukin-1

The IL-1 family comprises four proteins that share considerable sequence homology and contain a  $\beta$ -pleated sheet structure (Dinarello, 1997): IL-1 $\alpha$ , IL-1 $\beta$ , IL-1 receptor antagonist (IL-1Ra), and IL-18 (also known as IFN $\gamma$ -inducing factor). Release of mature IL-1 $\alpha$  requires extracellular calpain-mediated cleavage of a pro-IL-1 $\alpha$ , whereas mature IL-1 $\beta$  is derived proteolytically from pro-IL-1 $\beta$  by intracellular IL-1 $\beta$ -converting enzyme (ICE or caspase-1) activity. Upon binding of IL-1 $\alpha$  or IL-1 $\beta$  to the IL-1 receptor type I (IL-1RI), IL-1R accessory protein (IL-1RIAcP) is recruited by the receptor complex, and intracellular signal transduction is triggered through a p38 mitogen-activated protein kinase (MAPK)-activated phosphorylation cascade. Due to extensive signal amplification, minute amounts of IL-1 can have considerable biological activity, and as little as 1 ng/kg intravenous IL-1 $\beta$  causes symptoms in humans. The signaling cascade culminates in the nuclear translocation of the transcription factors nuclear factor kappa B (NF- $\kappa$ B) and activating protein-1 (AP-1) and the ensuing transcription of a variety of proinflammatory genes, including autocrine amplification of IL-1 production (Suzuki et al., 1989). In addition to the IL-1RI, IL-1 may also bind to the so-called type II interleukin-1 receptor, the expression of which appears to be regulated

by IL-4 (Colotta et al., 1993b). Binding of IL-1 to this receptor does not result in cellular activation, and IL-1RII is therefore presumed to act as a decoy that negatively regulates IL-1 activity.

A further member of the IL-1 cytokine family, IFN $\gamma$ -inducing factor, has been termed IL-18, on the basis of its pleiotropic Th1-inducing effects (Ushio et al., 1996). It has been assigned to the IL-1 family on the grounds of sequence homology (26% with IL-1 $\beta$ ) and similarity of the IL-18 receptor to IL-1R (Torigoe et al., 1997; Dinarello, 1999). Like IL-1 $\beta$ , IL-18 is dependent on ICE for proteolytic processing, and on nuclear translocation of NF- $\kappa$ B for transcriptional activation.

Owing to its proinflammatory effects on endothelial cells (Jirik et al., 1989; Loppnow and Libby, 1989a,b; Sironi et al., 1989; Suzuki et al., 1989; Sica et al., 1990b; Bochner et al., 1991; Clinton et al., 1992; Collins et al., 1995; Garcia et al., 2000), smooth muscle cells (Loppnow and Libby, 1989a, 1990; Wang et al., 1991; Clinton et al., 1992; Braun et al., 1995; Stanford et al., 2000), and macrophages (Sica et al., 1990b), and due to its production by all of these cell types in atherosclerotic lesions (Moyer et al., 1991; Tipping and Hancock, 1993; Galea et al., 1996), IL-1 was one of the first cytokines to be considered instrumental in the propagation of vessel wall inflammation in atherosclerosis. It is thought to facilitate early lesion formation by increasing leukocyte adhesion to endothelial cells (Bevilacqua et al., 1985; Wang et al., 1995) and mediating leukocyte transmigration.

tion (Moser et al., 1989; Furie and McHugh, 1989). Subsequently, locally produced IL-1 may serve to maintain an inflammatory milieu by autocrine and paracrine stimulation of cytokine (Jirik et al., 1989; Loppnow and Libby, 1989a,b, 1990, 1992; Sironi et al., 1989; Sica et al., 1990a,b; Wang et al., 1991; Clinton et al., 1992; Li et al., 1995; Taki et al., 1999; Garcia et al., 2000; Stanford et al., 2000) and adhesion molecule expression (Osborn et al., 1989; Bochner et al., 1991; Braun et al., 1995; Collins et al., 1995). In the advanced plaque, IL-1-induced up-regulation of matrix metalloproteinases may destabilize the proteinaceous scaffold of the cap and thereby have a hand in plaque rupture (Galis et al., 1995; Libby et al., 1995); this hypothesis is corroborated clinically by the fact that a particular IL-1 $\beta$  gene polymorphism has been found to be associated with myocardial infarction in chlamydia pneumoniae seropositive patients (Moriyama et al., 2001), and that pericardial fluid levels of IL-1 $\beta$  are raised in patients with unstable angina pectoris (Oyama et al., 2001).

Because the IL-18 signal transduction cascade is similar to that activated by IL-1, it is perhaps unsurprising that IL-18 has also been found to up-regulate the expression of intercellular adhesion molecule 1 (ICAM-1) and cytokines by monocytes, including IL-1 $\beta$ , IL-6, and IL-8 (Dinarello, 1999), and the production of vascular cell adhesion molecule-1 (VCAM-1) by endothelial cells (Vidal-Vanaclocha et al., 2000). It is, therefore, entirely conceivable that IL-18 may have pro-atherogenic properties, and Mallat et al. (2001a) have indeed demonstrated IL-18 in atherosclerotic plaques in human carotids, which is primarily localized to macrophages. They found the corresponding receptor, IL-18R, to be expressed on endothelial cells and macrophages and barely present on SMCs. These findings have subsequently been confirmed histologically and in vitro by Gerdes et al. (2002), who also demonstrated the functionality of the IL-18 receptor on these cells through IL-18-mediated induction of pro-atherogenic factors, including IL-6, IL-8, ICAM-1, and matrix metalloproteinases. In addition, the serum level of IL-18 has recently been identified as a strong predictor of cardiovascular death in stable and unstable angina (Blankenberg et al., 2002). The pro-atherogenic effects of IL-18 are thought to be mediated by IFN $\gamma$ , since the induction of atherosclerosis by exogenous IL-18 is abrogated by IFN $\gamma$  deficiency in apolipoprotein E knockout (apoE $^{-/-}$ ) mice (Whitman et al., 2002). A role for IL-18 in plaque destabilization was suggested by the up-regulation of IL-18 mRNA levels in symptomatic and ulcerative atherosclerotic plaques (Mallat et al., 2001a).

In comparison with the proinflammatory reprobates of the IL-1 family, IL-1ra appears positively angelic. IL-1ra displays affinity for the IL-1R, but it does not induce a cellular response; it is therefore believed to be an endogenous inhibitor of IL-1 signaling (Dinarello, 1997). IL-1ra is produced by monocytes (Arend et al.,

1990), macrophages (Janson et al., 1991), and smooth muscle cells (Beasley et al., 1995). Recombinant intracellular IL-1ra has been shown to counteract the IL-1-induced production of IL-6, IL-8, and monocyte chemoattractant protein by human endothelial cells (Bertini et al., 1992), and to inhibit smooth muscle cell proliferation (Porreca et al., 1993). Moreover, vascular inflammation is the major phenotypic characteristic of IL-1ra-deficient mice (Nicklin et al., 2000), whereas atherogenesis is reduced in IL-1ra transgenic mice on a high fat diet (Devlin et al., 2002), and fatty streak formation is reduced in apoE $^{-/-}$  mice by IL-1ra administration (Elhage et al., 1998). IL-1ra has been found to be present in carotid atherosclerotic plaques (Gottsater et al., 2002), and the relevance of IL-1ra to human atherosclerosis is underscored by the fact that certain IL-1ra alleles are associated with coronary artery disease (Francis et al., 1999) and restenosis (Kastrati et al., 2000; Francis et al., 2001).

### B. Interleukin-2

This family of cytokines encompasses a group of interleukins which share a common receptor subunit, the "common  $\gamma$  chain" ( $\gamma$ c chain), which acts in unison with a subtype specific  $\alpha$  chain to initiate the signaling cascade. As the common receptor subunit was initially discovered in relation to IL-2, it has also been termed the "IL-2 receptor  $\gamma$  chain" (Takeshita et al., 1990), and the group of cytokines that interact with this receptor has consequently been termed the "IL-2 family" (Leonard and Lin, 2000). The members of this interleukin family are primarily involved in T cell development and activation, and mutations of the  $\gamma$ c chain cause X-linked severe combined immunodeficiency in humans (Noguchi et al., 1993b) and lead to thymic hypoplasia in mice (Cao et al., 1995).

In addition to IL-2, the family includes IL-4 (Russell et al., 1993), IL-7 (Noguchi et al., 1993a), IL-9 (Russell et al., 1994), IL-15 (Giri et al., 1994a), and IL-21 (Vosshenrich and Di Santo, 2001). All members interact with receptor complexes consisting of an interleukin-specific  $\alpha$  chain and the common  $\gamma$ c chain (Fig. 1). Moreover, the IL-4  $\alpha$  chain is also a component of the IL-13 receptor complex (Zurawski et al., 1993), and for purposes of classification, we shall include IL-13 in this interleukin family. A substantial degree of functional redundancy is extolled by the IL-2 family members, which is comprehensible in view of considerable overlap in their signaling pathways. Thus, Janus kinase 1 (Jak1) and Jak3 have been found to be activated by the subtype-specific chains and the constant  $\gamma$ c chain, respectively (Miyazaki et al., 1994; Russell et al., 1994; Leonard and Lin, 2000), which ultimately cascades into the activation of transcription by the common downstream effector molecules "signal transducer and activator of transcription" 5a (Stat5a), Stat5b, and Stat3 (Lin et al., 1995; Lin and Leonard, 2000). IL-4 and IL-13 are somewhat distinct in

activating Jak2 and Stat-6 via a  $\gamma$  chain-independent pathway (Palmer Crocker et al., 1996).

IL-2 (Arbustini et al., 1991; Frostegard et al., 1999) and the IL-2R receptor (Kishikawa et al., 1993) are expressed in atheromatous tissue, but a direct causal role for IL-2 in atherogenesis remains to be proven. Nonetheless, serum IL-2 levels have been found to be elevated in ischemic heart disease (Mazzone et al., 1999) and especially unstable angina pectoris (Mizia-Stec et al., 2002), and the risk of acute myocardial infarction is increased following IL-2 treatment for cancer (Kragel et al., 1990). A possible explanation for the presumed pro-atherogenic effect of IL-2 may lie in its ability to induce a T helper cell shift toward a Th1 phenotype. T cells have been shown to be present in atherosclerotic lesions (Hansson et al., 1988), and Th1 cells, in particular, are believed to actively promote atherogenesis (de Boer et al., 1999; Frostegard et al., 1999; Huber et al., 2001; Laurat et al., 2001; Song et al., 2001). In its capacity as an autocrine stimulator of Th1 cell differentiation and proliferation (Kurt-Jones et al., 1987; Harel-Bellan et al., 1988), IL-2 may promote the expansion and activation of this T cell subset, and, consequently, plaque development.

Conversely, IL-4 is known to promote Th2-type responses (partly by autocrine activation) and to exert immunosuppressive effects on macrophages, including the suppression of proinflammatory cytokine production and the stimulation of IL-1 $\alpha$  elaboration (Paul, 1991). This cytokine is therefore considered to be potentially anti-atherogenic. The highly pleiotropic effects of IL-4, however, reserve a rather more complicated role for IL-4 in atherosclerosis. Thus, whereas mice deficient in Stat6, which is one of the mediators activated by IL-4, develop larger atherosclerotic lesions than their wild-type counterparts (Huber et al., 2001), IL-4 deficient mice do not display increased susceptibility to diet-induced atherosclerosis (George et al., 2000a). They have even been found to be relatively resistant to the acceleration of fatty streak formation by heat shock protein 65 or mycobacterium tuberculosis (George et al., 2000b). Similarly, reconstitution with IL-4-deficient bone marrow in LDLr $^{-/-}$  mice reduces atherosclerotic lesion formation in the aortic arch and the thoracic aorta compared with reconstitution with wild-type bone marrow (King et al., 2002). Although IL-4 expression in atherosclerotic plaques appears to be limited (Uyemura et al., 1996), among the pro-atherogenic effects of IL-4 we may count the up-regulation of P-selectin (Khew-Goodall et al., 1999) and 15-lipoxygenase (Lee et al., 2001b) expression by endothelial cells, VCAM-1 (Barks et al., 1997) and matrix metalloproteinase 1 (MMP-1) (Sasaguri et al., 1998) expression by vascular smooth muscle cells, and the augmentation of CD36 receptor expression (Feng et al., 2000) and cholesterol esterification (Cornicelli et al., 2000) in macrophages. On the other hand, IL-4 has also been shown to inhibit smooth muscle cell

proliferation (Vadiveloo et al., 1994; Sasaguri et al., 1998) and macrophage adhesiveness (Elliott et al., 1991). The net effect of IL-4 in atherosclerosis thus still hangs in the balance, and it may vary with the stage of the disease.

IL-9 was initially identified as a mast cell and T cell growth factor (Renauld et al., 1990) and has subsequently been shown to lead to exaggerated Th2-type inflammatory responses (Godfraind et al., 1998; McLane et al., 1998) and thymic lymphomas (Renauld et al., 1994) in IL-9 transgenic mice. IL-9 is not entirely independent in its actions, however, since IL-9 production by T lymphocytes requires IL-2-mediated stimulation (Houssiau et al., 1992), and the mitogenic effect of IL-9 on T lymphocytes requires their preactivation (Uyttenhove et al., 1988). In a murine model of Gram-negative bacterial shock, IL-9 led to suppression of TNF $\alpha$ , IL-12, and IFN $\gamma$ , possibly mediated by an induction of IL-10 expression (Grohmann et al., 2000). In agreement with this study, IL-9 has been found to induce the expression of the intracellular cytokine signal inhibitors cytokine-inducible SH2-containing protein, suppressor of cytokine signaling (SOCS)-2 and SOCS-3 (Lejeune et al., 2001). SOCS-3, in particular, may impair signaling by pro-atherogenic cytokines that act through the gp130 receptor, including IL-6 and IL-12. Some of the activities of IL-9 may also be mediated by its induction of IL-22 (IL-TIF), which shares 22% sequence homology with IL-10 (Dumoutier et al., 2000). Although its role in atherosclerosis has thus far not been elucidated, it appears that IL-9 may be potentially anti-atherogenic through a deflection of the immune response from a Th1 to a Th2 type. Albeit that a caveat needs to be added, as overzealous stimulation of Th2 responses may well prove to be detrimental in the later stages of atherosclerosis. Thus, mast cells have been identified in advanced plaques (Kaartinen et al., 1994a; Jeziorska et al., 1997) and are presumed to promote plaque instability by the secretion of chymase (Kaartinen et al., 1994b; Kovanen, 1997) and the stimulation of calcification (Jeziorska et al., 1998). Their stimulation may promote, rather than impede, the development of atherosclerotic complications.

IL-15 is produced by a variety of cells, including monocytes (Musso et al., 1999) and endothelial cells (Oppenheimer-Marks et al., 1998; Krishnaswamy et al., 1999), and has an activity profile similar to IL-2, without sharing sequence homology (Waldmann and Tagaya, 1999). IL-15 mediates extravasation of lymphocytes through its stimulatory and chemotactic effects on natural killer cells (Carson et al., 1994; Allavena et al., 1997) and T lymphocytes (Giri et al., 1995; Sancho et al., 1999) and by the up-regulation of hyaluronan on the endothelium (Estess et al., 1999). Recently, atherosclerotic lesions in humans and apoE $^{-/-}$  mice were found to contain IL-15-responsive T cells as well as IL-15 itself, which colocalizes with oxidized LDL-positive macrophages (Houtkamp et al., 2001, Wuttge et al., 2001). IL-15 may

therefore accelerate atherogenesis by promoting the recruitment and antigen-independent induction of T lymphocytes.

Despite sharing only 20 to 25% sequence homology and differing from IL-4 in lacking an effect on T cell function (Zurawski and de Vries, 1994), IL-13 is highly akin to IL-4 with respect to its immunomodulatory properties (Opal and DePalo, 2000), which is likely to be attributable to IL-4R-mediated Stat6 activation by both cytokines (Hart et al., 1999). In monocytes, IL-13 attenuates the expression of a wide range of inflammatory cytokines, including IL-1, IL-6, IL-8, IL-10, IL-12, MIP-1 $\alpha$ , granulocyte-macrophage colony-stimulating factor (GM-CSF), granulocyte colony-stimulating factor (G-CSF), IFN $\alpha$ , and TNF $\alpha$ , while up-regulating the expression of IL-1ra (de Waal Malefyt et al., 1993; Mijatovic et al., 1997). Nitric oxide production is inhibited by IL-13 in macrophages (Doherty et al., 1993; Bogdan et al., 1997) and smooth muscle cells (Ruetten and Thiemermann, 1997). The properties of IL-13 are not exclusively anti-inflammatory, however, as exemplified by the IL-13-mediated potentiation of IL-8 receptor expression, 15-lipoxygenase expression, and LDL oxidation by monocytes (Nassar et al., 1994; Folcik et al., 1997; Bonechi et al., 2000), and of IL-8 and MCP-1 release in response to IL-1 $\alpha$  or TNF $\alpha$  in SMCs (Jordan et al., 1997). Moreover, IL-13 is known to enhance the transmigration of leukocytes by stimulating the endothelial expression of adhesion molecules (Bochner et al., 1995; Ying et al., 1997) and chemotactic factors (Goebeler et al., 1997). In analogy with IL-4, the overall effect of IL-13 in atherosclerosis is still controvertible.

The complex actions of IL-2 family members in the vascular wall are depicted in Fig. 2.

### C. The gp130 Family

The common receptor subunit shared by the members of this family of cytokines, gp130, was first discovered as a signal transducer for IL-6 (Hibi et al., 1990). The other factors to employ this receptor subunit in combination with their own specific subunit, are IL-11 (Yin et al., 1993), IL-12 (Chua et al., 1994), leukemia inhibitory factor (Gearing et al., 1991), oncostatin M (Gearing et al., 1992), cardiotrophin-1 (Ip et al., 1992), ciliary neurotrophic factor (Pennica et al., 1995), and neurotrophin-1/B cell-stimulating factor-3 (Senaldi et al., 1999) (Fig. 1). Following gp130 binding, the Janus kinases Jak1, Jak3, and Tyk2 and the transcription factors Stat1 and Stat3 are phosphorylated (Heinrich et al., 1998). In this review, we shall restrict the discussion to the interleukin members of the gp130 family.

In addition, two novel heterodimeric interleukins with an activity profile similar to IL-12 have recently been identified. IL-23 is composed of a p19 subunit and the p40 subunit of IL-12 (Oppmann et al., 2000), and this cytokine acts through a receptor composed of IL-12R $\beta$ 1 and a novel cytokine receptor subunit, IL-23R (Parham

et al., 2002). IL-27 is made up of an IL-12 p40-related and an IL-12 p35-related protein and binds to the gp130-related receptor WSX-1/TCCR (Pflanz et al., 2002).

Endothelial cells, smooth muscle cells, and macrophages are capable of elaborating IL-6, and its expression has been observed in atherosclerotic lesions in humans, hypercholesterolemic rabbits, and apoE-deficient mice (Ikeda et al., 1992; Kishikawa et al., 1993; Seino et al., 1994; Rus et al., 1996; Sukovich et al., 1998; Schiefer et al., 2000). Although the endothelium is largely unresponsive to IL-6 (Podor et al., 1989), addition of the soluble IL-6R $\alpha$  subunit (sIL-6R) enables endothelial cells to mount an inflammatory response to IL-6, by interacting with membrane-bound gp130 (Jones et al., 2001). This process has been termed "trans-signaling", and it may lead to increased endothelial cell adhesiveness by the up-regulation of E-selectin, ICAM-1, and VCAM-1, and the release of inflammatory mediators, including MCP-1, IL-8, and IL-6 itself (Modur et al., 1997; Romano et al., 1997). Thus, sIL-6R present in serum and/or elaborated locally by cells in the intima may serve to augment endothelial adhesion and extravasation of leukocytes into the atherosclerotic plaque. Monocytes and macrophages, on the other hand, produce IL-6R autonomously and therefore do not depend on ambient sIL-6R levels for IL-6-mediated modulation of gene expression (Akira and Kishimoto, 1996). The effector functions of IL-6 in cells of the monocyte/macrophage lineage include the differentiation of monocytes to macrophages (Chomarat et al., 2000), the up-regulation of acute phase response gene expression in hepatocytes and macrophages (Perlmutter, 1989), and the priming of macrophages for enhanced TNF $\alpha$  production in response to lipopolysaccharide (LPS) administration (Cochran and Finch-Arietta, 1992). In smooth muscle cells, IL-6 induces proliferation directly (Nabata et al., 1990; Ikeda et al., 1991) and indirectly through the initiation of an autocrine loop mediated by the up-regulation of gp130 (Klouche et al., 1999). In addition, smooth muscle cells are stimulated by IL-6 to express ICAM-1 (Ikeda et al., 1993) and to evolve into foam cells (Klouche et al., 2000).

Whereas homozygous deletion of gp130 in mice leads to intrauterine death due to myocardial hypoplasia (Yoshida et al., 1996), IL-6-deficient mice develop normally despite an attenuated acute phase response and impaired cellular immunity to virus infection (Kopf et al., 1994). This is a reflection of the functional redundancy in gp130-mediated signaling and thus of the extent to which the other members of the gp130 family can take over IL-6-mediated functions. IL-6 was initially described as a lymphocyte stimulatory factor but has since been found to exert a plethora of inflammatory effects (Hirano et al., 1990). With the possible exception of IL-1, IL-6 is the cytokine with the most extensively studied pro-atherogenic profile. Causality has been established through the exacerbation of early atherosclerosis by recombinant IL-6 in various atherosclerosis-prone murine

models (Huber et al., 1999). Interestingly, the progression of atherosclerotic lesions to an advanced phenotype appears to be inhibited by IL-6 in apoE-deficient mice, uncovering a potentially biphasic mode of action in atherogenesis (Elhage et al., 2001), which is perhaps partly explained by its observed anti-inflammatory properties (Barton et al., 1996; Xing et al., 1998) and its inhibition of macrophage class A scavenger receptor expression (Liao et al., 1999). Nonetheless, inhibition of IL-6 signaling may be considered to constitute an attractive therapeutic strategy for the prevention of coronary heart disease (Stein and Kung Sutherland, 1998; Yudkin et al., 2000).

Clinically, elevated levels of IL-6 and its hepatic by-product C-reactive protein (Verma et al., 2002) are associated with increased risks of coronary and peripheral atherosclerosis (Erren et al., 1999; Mazzone et al., 1999; Flex et al., 2002; Bermudez et al., 2002; Kato et al., 2002; Stenvinkel et al., 2002; van der Meer et al., 2002), myocardial infarction (Ridker et al., 2000b; Ikeda et al., 2001), and the risk of death of patients with cardiovascular disease (Volpato et al., 2001), and IL-6 has been suggested to mediate the pro-atherogenic properties of cytomegalovirus (Blankenberg et al., 2001). In a large multicenter study, IL-6 gene polymorphisms were found to correlate with the severity of coronary artery disease and the risk of myocardial infarction (Georges et al., 2001), and carotid atherosclerosis has been shown to be independently linked with an IL-6 promoter polymorphism (Rauramaa et al., 2000; Rundek et al., 2002), as has the risk of coronary artery disease (Humphries et al., 2001). In addition, lower levels of soluble IL-6 receptor, a naturally occurring IL-6 antagonist, are linked with the risk of myocardial infarction (Ueda et al., 1999). Although these clinical findings do not establish causality, they have identified a strong association between IL-6 levels and atherosclerosis.

Despite sharing considerable redundancy with IL-6 with respect to its signaling and effector functions, IL-11 has been judged to be a more anti-inflammatory member of the gp130 family of cytokines based on the net effect of its pleiotropic actions (Schwertschlag et al., 1999; Taki et al., 1999). In macrophages, recombinant IL-11 has been found to attenuate macrophage expression of TNF $\alpha$ , IL-1 $\beta$ , IL-12, and nitric oxide following an LPS challenge (Trepicchio et al., 1996; Leng and Elias et al., 1997). These effects are direct and mediated by NF- $\kappa$ B down-regulation (Trepicchio et al., 1997), as is IL-11-mediated attenuation of smooth muscle cell proliferation and cytokine production (Zimmerman et al., 2002). In endothelial cells, IL-11 provides protection against immune-mediated injury (Mahboubi et al., 2000), and inhibits apoptosis through up-regulation of survivin (Mahboubi et al., 2001). In CD4<sup>+</sup> lymphocytes, IL-11 has been found to induce a shift from a Th1 to a Th2 phenotype (Bozza et al., 2001). This effect has been put to use in immunomodulatory treatment employing IL-11 in

psoriasis (Trepicchio et al., 1999) and Crohn's disease (Sands et al., 1999), and it may also offer therapeutic possibilities in the setting of atherosclerosis.

Activated monocytes are the primary source of IL-12 (D'Andrea et al., 1992), a cytokine that induces proliferation (Gately et al., 1991) and a shift toward a Th1 expression pattern in lymphocytes (Hsieh et al., 1993). IL-12 was originally implicated in atherosclerosis by Uyemura et al. (1996), who observed an abundance of p40 mRNA and IL-12 p70 protein in atherosclerotic lesions, and up-regulation of IL-12 production by monocytes following the addition of highly oxidized LDL. Subsequently, atherosclerotic lesions in apoE-deficient mice were found to contain IL-12, and their progression to be accelerated by daily injections of recombinant IL-12 (Lee et al., 1999). Conversely, a selective defect of macrophage IL-12 synthesis due to 12/15-lipoxygenase deficiency reduces lesion formation in atherosclerosis-prone Apobec-1<sup>-/-</sup>/ApoE<sup>-/-</sup> mice (Zhao et al., 2002). In clinical practice, raised serum levels of IL-12 have been found to be associated with acute myocardial infarction (Zhou et al., 2001a).

#### D. Granulocyte-Macrophage Colony-Stimulating Factor

The genes encoding the members of this family—IL-3, IL-5, and GM-CSF—are clustered on the human chromosome 5 (van Leeuwen et al., 1989) (Fig. 1). Their products bind to receptor complexes consisting of a common  $\beta$  chain ( $\beta$ c) and a cytokine-specific  $\alpha$  chain (Hayashida et al., 1990; Kitamura et al., 1991), resulting in the activation of JAK/STAT, the ras/MAPK, and the phosphatidylinositol-3 kinase pathway (Guthridge et al., 1998). The primary effector functions to be identified for this family are the promotion of hematopoietic proliferation, survival, and differentiation, which is confirmed by the invariable occurrence of a myeloproliferative disorder in human common  $\beta$  chain transgenic mice (Nishinakamura et al., 1995). Since mice deficient for IL-3, IL-5, or GM-CSF suffer from pulmonary alveolar proteinosis, signaling via receptors involving the common  $\beta$  chain is thought to exert additional pleiotropic actions on mature cells of the monocyte/macrophage lineage (D'Andrea et al., 1998).

Indeed, IL-3 has been found to stimulate adhesion (Elliott et al., 1990) and c-jun expression in monocytes (Mufson et al., 1992). It is elaborated by activated T lymphocytes in atheromatous tissue and acts on smooth muscle cells to increase migration and proliferation (Brizzi et al., 2001). Moreover, receptors for IL-3 are also present on endothelial cells (Colotta et al., 1993a), which respond to the binding of IL-3 by increased proliferation, by augmented adhesion molecule, major histocompatibility complex II, and cytokine production, and by participating in angiogenesis in vivo (Brizzi et al., 1993; Korpelainen et al., 1995; Dentelli et al., 1999). IL-3 is thus believed to play a role in the early stages of atherogenesis by facilitating leukocyte extravasation and in



advanced lesions by augmenting macrophage activation, smooth muscle cell accumulation, and neovascularization of the plaque.

The involvement of IL-5 in the stimulation of B cell and eosinophil responses has been meticulously documented, with the aid of, *inter aliter*, IL-5 transgenic (Tominaga et al., 1991), and IL-5-deficient mouse models (Kopf et al., 1996). Its role in atherosclerosis remains uncharted territory, however. IL-5 is produced by endothelial cells (Krishnaswamy et al., 1999), but their expression of the IL-5 $\alpha$  receptor subunit is limited (Colotta et al., 1993a). IL-5 expression appears to be scanty in advanced human atherosclerotic plaques, and is associated with the presence of eosinophils (Frostegard et al., 1999). Because IL-5 is an archetypal Th2 lymphokine, it may activate mast cells in the atherosclerotic plaque, which have been associated with the development of unstable lesions and plaque rupture (Kaartinen, 1994a,b, 1996a,b, 1998; Kovanen et al., 1995). Notwithstanding its low prevalence, the significance of locally produced IL-5 may thus increase in importance with the age of the lesion, and this could lead to destabilization of the atheroma.

#### E. Interleukin-10

This family comprises a sizeable array of mammalian and viral molecules that possess a considerable degree of sequence similarity with its founder member, IL-10 (Rich and Kupper, 2001; Volk et al., 2001). These include IL-19, IL-20, IL-22, IL-24/MDA-7, IL-26/AK155, and the IL-10 homologs encoded by Epstein-Barr virus, cytomegalovirus, herpesvirus papio, and Yaba-like disease virus (Fickenscher et al., 2002; Wolk et al., 2002; Fig. 1).

IL-10 was initially identified as a cytokine synthesis inhibitory factor (CSIF) (Fiorentino et al., 1989), but has subsequently been found to be a pleiotropic immunoregulatory cytokine that is secreted by a wide variety of cells, including lymphocytes and monocytes/macrophages (Lalani et al., 1997b; Moore et al., 2001). IL-10 signaling is mediated by Jak1 and Stat3 and entails the down-regulation of NF- $\kappa$ B activity (Schottelius et al., 1999). Its effector functions include induction of a shift of T cell cytokine expression from a Th1 to a Th2 profile (Fiorentino et al., 1989), and attenuation of the production of proinflammatory cytokines by macrophages (Bogdan et al., 1991; de Waal Malefyt et al., 1991a; Lang et al., 2002) and polymorphonuclear neutrophils (Cassatella et al., 1993). In addition, IL-10 effects differentiation of monocytes to macrophages (Allavena et al., 1998), suppression of antigen-presenting activity (de Waal Malefyt et al., 1991b), a decline in the release of reactive nitrogen and oxygen intermediates (Gazzinelli et al., 1992; Mallat et al., 1999a; Haddad and Fahlman, 2002), and inhibition of ICAM-1 expression (Song et al., 1997). Monocyte adhesion to endothelial cells is attenuated by IL-10 through modulation of monocyte CD18 and CD62-L expression (Mtairag et al., 2001) and attenua-

tion of ICAM-1 and VCAM-1 expression on endothelial cells (Krakauer, 1995; Lindner et al., 1997). IL-10 has been found to be present in mature plaques (Uyemura et al., 1996; Mallat et al., 1999a) and is thought to play an active role in curbing the inflammatory milieu of the vessel wall (Tedgui and Mallat, 2001). This is supported by the observation that IL-10 knockout (IL-10 $^{-/-}$ ) mice suffer from accelerated atherosclerosis, whereas IL-10 transgenic mice are relatively protected (Pinderski-Oslund et al., 1999). Clinical poignancy is added by the fact that a hypoactive allele of the IL-10 promoter sequence increases the risk of cardiovascular events in hemodialysis patients (Girndt et al., 2002), whereas serum levels of IL-10 have been found to be decreased in patients with unstable angina compared with patients with chronic stable angina (Smith et al., 2001). Indeed, as IL-10 is known to down-regulate MMP-9 production and up-regulate tissue inhibitor of metalloproteinase-1 (TIMP-1) expression in macrophages (Lacraz et al., 1995), IL-10 may have a direct stabilizing influence on advanced plaques. Moreover, the combined weight of these data has led to extensive speculation about the therapeutic applicability of IL-10 in atherosclerosis (Terkeltaub, 1999).

#### F. Chemokines

On the basis of their chemoattractant activity for leukocytes, the interleukins IL-8 and IL-16 have been classified as chemokines (Center and Cruikshank, 1982; Mukaida et al., 1989). IL-16 has not been scrutinized in an atherosclerotic context, and any potential influence is likely to be mediated mainly by its effects on lymphocyte function, which include stimulation of migration, proliferation, and cytokine production (Cruikshank et al., 2000). IL-8, on the other hand, is well established as a pro-atherogenic factor (Reape and Groot, 1999). Its expression is induced in monocytes and macrophages following the addition of oxidized LDL and cholesterol, respectively (Terkeltaub et al., 1994; Wang et al., 1996). Atheromatous tissue has been found to contain IL-8, most of which is thought to be derived from intimal macrophages (Apostolopoulos et al., 1996; Wang et al., 1996). In addition, cytokine-stimulated vascular smooth muscle cells elaborate IL-8 (Wang et al., 1991), and endothelial cells respond to cyclic stretch by up-regulation of IL-8 production (Okada et al., 1998). Boisvert et al. (1998) have discovered an important role for macrophage-derived IL-8 in atherosclerotic lesion development, as transplantation of IL-8 $^{-/-}$  bone marrow to irradiated and atherogenic diet-fed LDLr $^{-/-}$  mice resulted in less extensive intimal macrophage accumulation than transplantation using IL-8 $^{+/+}$  donors. IL-8 is presumed to accelerate atherogenesis by increasing the endothelial adhesiveness for monocytes (Gerszten et al., 1999), by its mitogenic and chemoattractant actions on smooth muscle cells (Yue et al., 1994), and by mediating angiogenesis in the atherosclerotic plaque (Simonini et

al., 2000). Furthermore, IL-8 may cause destabilization of advanced plaques through its inhibitory effect on TIMP-1 expression in macrophages and an ensuing increase in metalloproteinase activity (Moreau et al., 1999). Interestingly, IL-8 levels have been found to be elevated in peripheral blood monocytes from hypercholesterolemic patients (Porreca et al., 1999), and serum IL-8 levels to be associated with unstable angina pectoris and acute myocardial infarction (Zhou et al., 2001a), reflecting the potential clinical relevance of IL-8-mediated functions in atherosclerosis.

### G. Interleukin-17

The term IL-17 harbors a family of proinflammatory cytokines, of which the founder member was found to be an ortholog of murine CTLA-8 (Rouvier et al., 1993) and its gene to have been captured by the T lymphotropic herpesvirus saimiri (Rouvier et al., 1993, Yao et al., 1995a,b; Fossiez et al., 1998). It is primarily produced by activated memory T cells and Th1/Th0 cells (Aarvak et al., 1999) and binds to a ubiquitously expressed receptor (Yao et al., 1995a). More recently, the variants IL-17B, IL-17C, IL-17E, IL-17F, and IL-25 have been cloned, which are considered to signal through subtype-specific receptors (Li et al., 2000b; Hymowitz et al., 2001; Lee et al., 2001a; Hurst et al., 2002). IL-17 induces the expression of proinflammatory mediators by a variety of cells, including the production of IL-6 and IL-8 by stromal cells (Yao et al., 1995a,b), ICAM-1 by fibroblasts and keratinocytes (Yao et al., 1995b; Albanesi et al., 1999), as well as IL-1 $\beta$ , IL-1ra, IL-6, IL-10, TNF $\alpha$ , prostaglandin E $_2$ , MMP-3, and MMP-9 by macrophages (Jovanovic et al., 1998, 2001). Binding of IL-17 to its receptor results in an increase in Ca $^{2+}$  influx, a decrease of intracellular cAMP levels, activation of mitogen-activated protein kinases, and stimulation of NF- $\kappa$ B activity (Jovanovic et al., 1998; Awane et al., 1999). The activity profiles of IL-17B and IL-17C differ from that of IL-17 in that they fail to induce IL-6 in fibroblasts but are capable of stimulating the release of TNF $\alpha$  and IL-1 $\beta$  from the monocytic cell line THP-1 (Li et al., 2000b). IL-17E has been shown to stimulate NF- $\kappa$ B activity and the production of IL-8 in TK-10 cells (Lee et al., 2001a). The IL-17 family has not yet been implicated in atherogenesis, but its proinflammatory effects on macrophages, the stimulation of endothelial IL-2 and MCP-1 elaboration by IL-17F (Starnes et al., 2001), the production of IL-17 by activated T cells, and the widespread expression of the IL-17 receptor make this interleukin family a potential pro-atherogenic candidate.

### III. Modulation of Cytokine Function As a Therapeutic Strategy for Atherosclerosis

From the preceding discussion it will have become evident that, despite having been thoroughly researched with respect to their basic immunological functions,

many of the interleukins identified to date have yet to be typecast on the atherosclerotic stage (Table 1). When classified according to their perceived role in atherogenesis, a large number thus remain in the "unknown" category. A similarly sizable group has been found to be pro-atherogenic, and only a small subset has been adjudicated to possess an equivocal (IL-4, IL-13) or anti-atherogenic (IL-1ra, IL-9, IL-10, IL-11) propensity. It therefore appears that the most rewarding strategies of interleukin modulation for the prevention of atherosclerosis are likely to involve the down-regulation of signaling mediated by proinflammatory cytokines. Nonetheless, due attention also needs to be paid to the intriguing therapeutic possibility of harnessing the anti-atherogenic potential of anti-inflammatory interleukins. The modulation of (patho)physiological effects exerted by cytokines that have thus far been adjudicated to have either an overtly pro- or an anti-atherogenic role on the evidence of animal intervention studies are, in the short term, the most likely candidates for the development of such strategies (Table 2; Fig. 3).

The function of interleukins is tightly regulated at a number of levels in their production, processing, and signaling cascades. Interleukins being proteins, the first step in their production necessitates the binding of nuclear transcription factors to enable gene transcription. Following mRNA translation, the production of mature molecules requires additional proteolytic processing for a number of interleukins and interleukin receptors. The ambient concentration of some interleukins is known to be negatively regulated following exposure on the cell surface or release into the surrounding extracellular

TABLE 1  
The (causative, associative, or presumed) role of interleukin family members in atherosclerosis

	Pro-Atherogenic	Equivocal	Anti-Atherogenic	Unknown
IL-1 family	<b>IL-1<math>\alpha/\beta</math></b> <b>IL-18</b>		<b>IL-1ra</b>	
IL-2 family	IL-2	IL-4	<b>IL-9</b>	IL-7 IL-21
CSF family	<i>IL-3</i>	<i>IL-13</i>		IL-5
gp130 family	<b>IL-6</b> <b>IL-12</b>		<i>IL-11</i>	IL-23 IL-27
Chemokines	<b>IL-8</b>			IL-16
IL-10 family			<b>IL-10</b>	IL-19 IL-20 IL-22 IL-24 IL-26 IL-25 IL-14
IL-17 family	<i>IL-17A/B/C/E/F</i>			
Unclassified				

Bold type represents causative role; italic type represents presumed role; light-face roman type represents associative role.

TABLE 2

Primary vascular sources, targets, and selected effects of interleukins for which a pro- or anti-atherogenic role has been established in murine intervention studies.

Primary Sources	Primary Vascular Effector Cells and Direct Effects				Murine Intervention Studies
	Endothelial Cells	Monocytes/Macrophages	Smooth Muscle Cells		
IL-1 EC, T cell, Mφ, SMC (IL-1ra: EC, Mφ, SMC)	IL-1, IL-6, IL-8, MCP-1, proliferation M-CSF, fractalkine, IL-1RI, ICAM-1, VCAM-1, E-selectin, tissue factor, heme oxygenase, apoptosis	IL-6 PDGF-AA, superoxide	IL-6, IL-8, IL-11, MCP-1, M-CSF, GM-CSF, ICAM-1, VCAM-1, iNOS, COX-2, Mn-SOD, stromelysin, interstitial collagenase, proliferation	↑	↑ vascular inflammation in IL-1ra-deficient mice ↓ atherogenesis in IL-1ra-transgenic mice
IL-6 EC, Mφ, SMC	IL-6, migration HGF proliferation	MCP-1	Proliferation	↑	rIL-6 ↓ fatty streak formation rIL-6 ↑ atherogenesis rIL-6 ↓ plaque progression ↑ atherogenesis following transfer of IL-8-transgenic bone marrow
IL-8 EC, Mφ, SMC	Monocyte adhesion	TIMP-1	Chemotaxis, proliferation	↑	rIL-12 ↑ plaque progression IL-9 ↓ atherogenesis IL-9 immunization ↑ atherogenesis
IL-12 IL-9 EC, Mφ T cell	VCAM-1	TNFα oxidative burst		↑	↑ atherogenesis in IL-10-deficient mice
IL-10 T cell, Mφ	IL-6, IL-8 VCAM-1, ICAM-1, P-selectin, E-selectin, angiogenesis	TIMP-1, differentiation, phagocytosis	Cytokine production, TNFα	↑	PLA <sub>2</sub> , proliferation
		ICAM-1, CD18, CD62-L, tissue factor		↓	↓ atherogenesis in IL-10-transgenic mice
		iNOS		↓	↓ atherogenesis following transfer of IL-10-transgenic bone marrow
		MMP-9			plasmid-mediated/adenoviral IL-10 gene transfer ↓ atherogenesis
		MHC II, B7, NF-κB activation, activation, proliferation			

Cox-2, cyclooxygenase-2; EC, endothelial cell; HGF, hepatic growth factor; M-CSF, monocyte colony-stimulating factor; PLA<sub>2</sub>, phospholipase A<sub>2</sub>; SR-A, scavenger receptor-A; TIMP-1, tissue inhibitor of metalloproteinase-1.

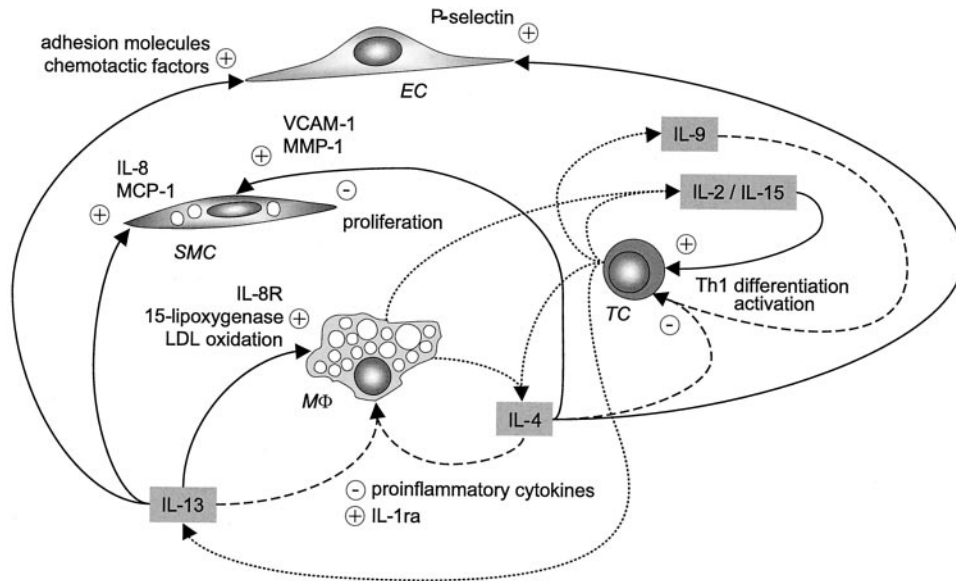


FIG 2. Overview of the complex actions of IL-2 family members in the vascular wall. All members are produced by cellular constituents of the atherosclerotic plaque (dotted line: EC, endothelial cell; SMC, smooth muscle cell; MΦ, macrophage; TC, T cell) and exert effects that are considered to be either pro-atherogenic (continuous line) or anti-atherogenic (dashed line).

space. This may involve neutralization of interleukins by binding to a specific antibody or to a soluble form of its corresponding receptor.

Interleukin molecules that escape endogenous regulation mechanisms can bind to their target receptor and thus initiate a signaling sequence. The abundance of the membrane-bound form of interleukin receptors may be controlled by endocytosis and degradation via the ubiquitin-proteasome system. The signaling cascade is frequently rather complex and often shares redundancy with those activated by other members of a particular interleukin family. A varied array of pathways has been found to convey interleukin signaling to the nucleus, frequently involving receptor-mediated activation of kinases (including Jaks, Tyks, and MAPKs) and subsequent activation of nuclear transcription factors (includ-

ing STATs, NF-κB, and AP-1) (Fig. 1). Intracellular signal transduction is negatively controlled by specific inhibitors of the Jak-STAT pathway that regulate its components by dephosphorylation, degradation by the ubiquitin-proteasome pathway, and binding of dominant-negative STATs. Signaling eventually culminates in the transcriptional activation of a cytokine-specific set of genes, the products of which mediate the biological functions of the cytokine in question by intracellular, autocrine, paracrine and endocrine mechanisms.

In theory, any step in the production and effector pathways of a particular interleukin may be considered to represent a potential target for therapies aimed at modulating its biological activity (Fig. 4). In practice, various approaches are not yet feasible due to a lack of detailed understanding of the mechanisms involved.

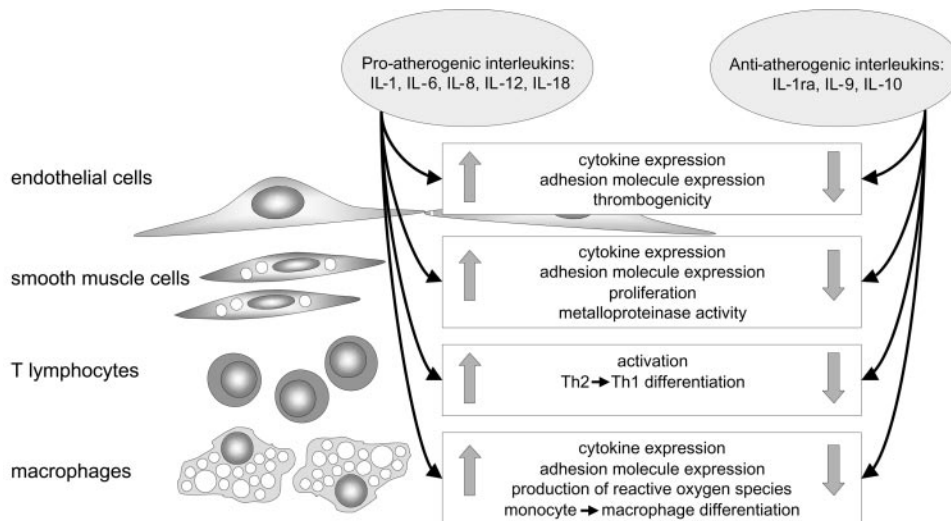


FIG 3. Primary vascular target cells and summarized actions of proven pro- and anti-atherogenic interleukins.

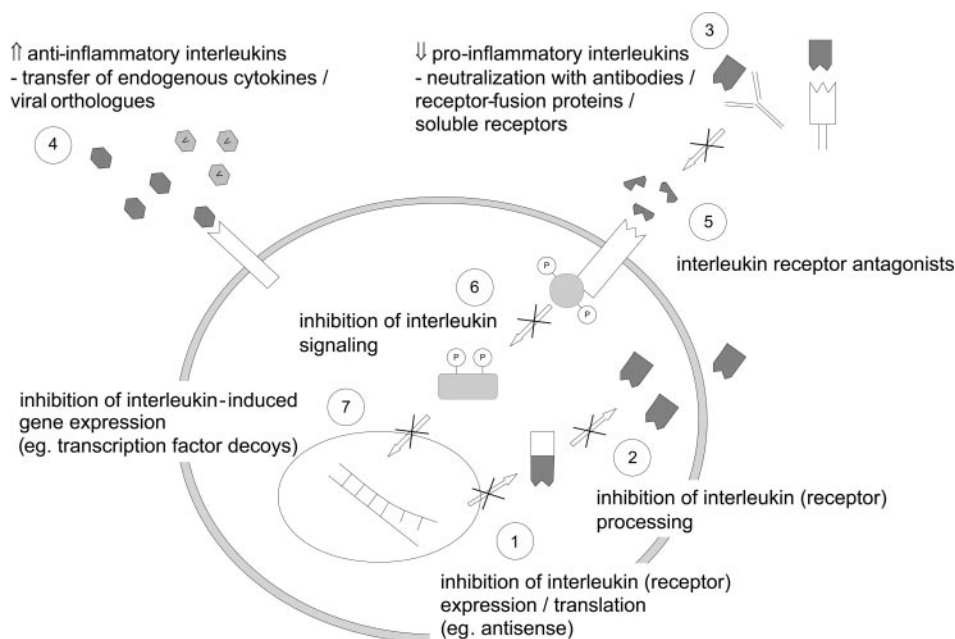


FIG 4. Depiction of potential strategies for the modification of interleukin activity as a therapy for atherosclerosis.

Moreover, the specificity of such interventions is frequently limited by considerable redundancy in interleukin processing and signaling pathways. Although this may be desirable if the goal of the intervention is a general reduction of proinflammatory signaling, a more subtle change of cellular functions may require direct alteration of extracellular interleukin levels or interleukin-receptor interaction. In the specific case of atherosclerosis, the more difficult hurdles on the course to the clinical use of cytokine modulation therapy are hidden in the insidious and chronic nature of the atherosclerotic process (Ross 1986, 1993a, 1999). Inherent in this observation is the need for any strategy aimed at *primary* prevention to be comparably chronic in its duration of action. In view of the high prevalence of the disease and its still poorly predictable course, such a strategy would also need to be safe, effective, and affordable. Most of the interleukin-based treatments that have been conceived thus far do not answer these demands. In the meantime, it may be more realistic to focus on a remedy that is capable of effecting *secondary* or *tertiary* prevention. An example of the latter is the phenotypic stabilization of unstable atherosclerotic atheromata to avert the risk of plaque rupture and fatal thrombosis. This may be achievable by the use of a short, and possibly localized, course of anti-interleukin therapy.

In this review, we discuss examples of techniques directed at modulating each of the steps described above. We shall pay particular attention to methods that have been shown to hold promise for the prevention of atherosclerosis or those that interfere with the function of interleukins thought to be involved in atherogenesis.

#### A. Inhibition of Expression / Translation of Interleukins and Their Receptors

The foremost approach to the specific inhibition of interleukin (receptor) expression and translation has been the use of short strands of (modified) nucleotides that are complimentary to stretches of mRNA encoding the target protein. This is thought to lead to formation of DNA:RNA duplexes and subsequent degradation of the mRNA sequence by RNaseH. The advent of this oligonucleotide-based “antisense” therapy was hailed as the dawn of a new era of highly specific and effective treatments for a variety of diseases, ranging from cancer to hypertension (Raizada et al., 2000; Lebedeva and Stein, 2001). This unbridled optimism has been somewhat deflated in recent years, however, as it has transpired that the mechanism of action of antisense molecules is frequently less specific and far more complex than originally conceived (Lebedeva and Stein, 2001). Moreover, unmodified oligonucleotides are rapidly degraded *in vivo*, and efficient transfection of target cells with antisense constructs has proved difficult. Nonetheless, several studies describing the antisense-mediated down-regulation of interleukin production have been reported (Crooke, 2000).

IL-1 $\alpha$  is known to inhibit endothelial cell proliferation, and thereby to promote the type of endothelial injury that is thought to precipitate atherogenesis (Ross, 1986). Furthermore, IL-1 $\alpha$  is an autocrine stimulator of adhesion molecule expression, including ICAM-1 and E-selectin, and the up-regulation of these molecules by hypoxic endothelial cells has been found to be mediated by IL-1 $\alpha$  (Shreeniwas et al., 1992). Antisense oligodeoxynucleotides (ODNs) directed against IL-1 $\alpha$  have been

found to prevent endothelial cell senescence, to prolong their life span, and to hinder adhesion molecule production *in vitro* (Maier et al., 1990; Maier and Ragnotti, 1993). Moreover, the IL-1 $\alpha$ -mediated up-regulation of cyclooxygenase expression in endothelial cells has been shown to be limited by the addition of ODNs directed against protein kinase C (PKC), which is a mediator in the signal transduction pathway that leads to IL-1 $\alpha$  induction (Hsu et al., 1999). Because interleukin-1 also affects smooth muscle cell function, Hsu et al. (1999) transfected vascular smooth muscle cells *in vitro* with an Epstein-Barr virus-derived vector expressing IL-1 antisense transcripts, which repressed the expression of matrix genes such as type I collagen and fibronectin by smooth muscle cells and prolonged their life span. In macrophages, more specifically the macrophage-like cell line U937, the expression of IL-1 $\beta$  can also be down-regulated by means of antisense techniques employing phosphorothioate oligonucleotides (Yahata et al., 1996).

The platelet-derived growth factor (PDGF)-mediated up-regulation of IL-6 in smooth muscle cells can be attenuated by antisense ODNs directed against this pro-atherogenic interleukin (Roth et al., 1995). This has been shown to inhibit cell division, and has thus established IL-6 as a mediator of PDGF-induced smooth muscle cell proliferation. The feasibility of antisense-mediated inhibition of IL-6 expression in the vessel wall has been demonstrated by *ex vivo* pressure-mediated transfection of naked oligonucleotides into human saphenous vein explants, which resulted in 70 to 75% inhibition of IL-6 expression, as measured 2 h after the transfection procedure (Mann et al., 1999).

Chemokine function has also been successfully repressed by antisense techniques. Thus, the role of IL-8 as a monocyte-derived angiogenic factor was revealed *in vitro* by the inhibition of monocyte-induced angiogenic activity following the administration of an IL-8 antisense oligonucleotide (Koch et al., 1992), and pretreatment of human pulmonary artery endothelial cells with antisense against MCP-1 has been shown to reduce TNF $\alpha$ -induced *trans*-endothelial monocyte migration (Maus et al., 2000).

Rather than inhibiting the production of interleukins themselves, antisense strategies could also be deployed against interleukin signaling by altering the expression of the relevant receptor. Indeed, ODNs directed against the IL-1 receptor have been shown to diminish IL-1-stimulated prostaglandin E<sub>2</sub> synthesis in murine and human fibroblasts (Burch and Mahan, 1991), and *in vivo* applicability was confirmed by the finding that subcutaneous injection of IL-1 receptor antisense in mice decreased neutrophil accumulation at sites of IL-1 injection.

Intriguingly, ODNs containing cytidine phosphate guanosine motifs have been identified as potent stimulators of Th1 type responses, and this type of aspecific effect needs to be taken into account during the design of

anti-inflammatory antisense sequences (Chu et al., 1997). In a drive to enhance the specificity as well as the efficacy, tolerability, and duration of action of antisense-mediated mRNA cleavage, the attention has turned to the use of ribozymes. These are RNA molecules with intrinsic endonuclease activity, which bind to target RNA in a base pair-specific fashion, and subsequently catalyze the cleavage of this RNA strand by facilitating the hydrolysis of phosphodiester bonds (Zaug et al., 1986; James and Gibson, 1998). Indeed, stable expression of ribozymes aimed at IL-1 $\beta$  and ICE can effect a dramatic decrease in the steady-state levels of their target mRNAs in the monocytic cell line THP-1 (Leavitt et al., 2000) and minimized hammerhead ribozymes have been shown to be active against IL-2 (Sioud et al., 1997). *In vivo* efficacy and *in vitro* reduction of TNF-induced IL-6 production has been demonstrated for IL-6 ribozymes (Mahieu et al., 1994). The first cardiovascular target to have been successfully inhibited by ribozyme therapy directed against cytokine expression is transforming growth factor  $\beta$  (TGF $\beta$ ) production in smooth muscle cells (Su et al., 2000). *In vivo*, TGF $\beta$  ribozymes have been shown to reduce neointima formation in a rat model of vascular injury (Yamamoto et al., 2000).

Although ribozymes may prove to be more effective and specific than antisense oligonucleotides due to their enzymatic mode of action, they share similar limitations to their biological activity. Thus, efficient cellular transfection is difficult to achieve, and once it has occurred, the duration of action is curtailed by a short intracellular half-life. Both problems have been extensively addressed, to varying degrees of success. Cellular uptake has been increased by the use of lipid, peptide, and polymer delivery systems, and nuclease-mediated degradation has been inhibited by chemical modifications of the oligonucleotide backbone (Morishita et al., 1994; Hughes et al., 2001; Lebedeva and Stein, 2001). Circumventing both disadvantages in a single approach may be possible by cloning ribozymes into an expression vector that affords avid transfection of target cells in addition to an extended duration of expression. These are characteristics of viral vectors—retroviruses, adenoviruses, and adeno-associated viruses (AAVs) being the main protagonists. These viral vectors have all been used as a carrier for ribozymes, but AAVs are particularly promising as they combine the main advantage of adenoviruses, i.e., high efficiency of transduction, with the prolonged expression due to integration of transgenes in the genome that is typical of retroviruses (Monahan and Samulski, 2000). AAVs have been shown to be capable of transducing endothelial and vascular smooth muscle cells *in vitro*, but expression *in vivo* is confined to the adventitia (Lynch et al., 1997). Therefore, to constitute a useful gene transfer vehicle in the prevention of atherosclerosis, the issue of targeting accuracy needs to be addressed, for instance by selecting the most suitable virus serotype, or by using an adenovirus/AAV hybrid

system (Recchia et al., 1999; Monahan and Samulski, 2000).

### B. Inhibition of Interleukin Processing

As mentioned, the production of mature forms of various cytokines requires proteolytic processing of inactive precursors. This is exemplified by the conversion of pro-IL-1 $\beta$  and pro-IL-18 by ICE/caspase-1 (Wilson et al., 1994; Tone et al., 1997), and the cleavage of membrane-bound TNF $\alpha$  by TNF $\alpha$ -converting enzyme (Black et al., 1997). Interestingly, the shedding of the soluble form of several interleukin receptors has also been found to require metalloproteinase activity, including IL-1R, IL-2R, and IL-6R (Mullberg et al., 1995, 1997). With respect to atherosclerosis, interference with proteinase-dependent processing may constitute an attractive strategy to attenuate the release of active IL-1 $\beta$  or to inhibit sIL-6R-mediated transfer of IL-6 sensitivity to cells that do not themselves express IL-6R (Jones et al., 2001).

Several naturally occurring ICE inhibitors have been described (Croucher et al., 2000). Thus, cowpox virus protein A (CrmA) protects cells infected by cowpox from immunological clearance by preventing the release of IL-1 $\beta$  (Ray et al., 1992), whereas the baculovirus protein p35 performs a similar function in baculovirus-infected cells (Bump et al., 1995). Smooth muscle cells produce an endogenous ICE inhibitor, which has been identified as serpin proteinase inhibitor 9 (PI-9) (Schonbeck et al., 1997; Young et al., 2000). Interestingly, protein levels of this enzyme have been found to be decreased in unstable plaques in conjunction with a reciprocal up-regulation of IL-1 $\beta$ , suggesting an endogenous anti-inflammatory role for constitutive PI-9 expression. Consequently, inhibition of caspase-1 activity might be an effective strategy in the prevention of lesion destabilization. In considering the feasibility of therapeutic ICE inhibition, one might either opt to capitalize on the potency of naturally occurring antagonists, or one could interpret their action as a paradigm for the development of synthetic ICE inhibitors (Livingston, 1997). Several such compounds have been developed that display activity *in vitro*. This includes the down-regulation by WIN 67694 of the LPS-induced release of IL-1 $\beta$  by murine macrophages (Miller et al., 1995), and the reduction of human myocardial ischemic dysfunction in an *ex vivo* organ culture model by YVAD (Pomerantz et al., 2001). *In vivo*, a single intraperitoneal dose of VE-13,045 administered after an LPS challenge reduced murine IL-1 $\beta$  serum levels by 50 to 70% (Ku et al., 1996). Prior to clinical use, however, the specificity for caspase-1 of the compound in question needs to be warranted, in view of the degree of conservation that has been found to exist between the active sites of the caspase family members. Moreover, due heed should be paid to the potentially detrimental inhibition of caspase-1-mediated apoptosis, which could contribute toward tissue hyperplasia or even neoplasia.

### C. Neutralization of Proinflammatory Interleukins

The biological activity of interleukins is partially regulated by anti-cytokine antibodies, soluble cytokine receptors, and cytokine-binding proteins, the elaboration of which is frequently controlled by the interleukin concerned (Heaney and Golde, 1998; Slifka and Whitton, 2000). Soluble interleukin receptors are produced by alternative splicing of mRNA or by proteolytic cleavage of full-length receptors. For instance, IL-1 activity is inhibited by the soluble type II IL-1 receptor (Giri et al., 1990), which is shed from neutrophils in response to proinflammatory stimuli, including TNF, IL-13, and endotoxin (Colotta et al., 1994; Giri et al., 1994b). Its pathophysiological roles are thought to include the limitation of IL-1 activity in sepsis (Giri et al., 1994b). Whereas the plasma level of soluble IL-2R has been deemed to be a marker for T cell activation in ischemic heart disease (Simon et al., 2001), high levels of sIL-2R paradoxically reduce the relative risk of lesion instability, which is known to be associated with increased inflammatory activity in the plaque (Blum et al., 1995; Takeshita et al., 1997; Simon et al., 2001). Moreover, *in vitro* studies have evidenced the inhibition of IL-2-induced activation of peripheral mononuclear cells by sIL-2R (Zorn et al., 1994). Soluble IL-4-binding proteins are known to occur in mice (Fernandez-Botran and Vitetta, 1990) and humans (Fanslow et al., 1993). The benefit of sIL-4R in preventing IL-4-mediated inflammatory responses has been demonstrated in a murine model of asthma (Henderson et al., 2000), and its administration has been found to be safe and to stabilize lung functions in patients with moderate asthma (Renz, 1999).

For some soluble interleukin receptors, however, the effects are rather less clear-cut (Heaney and Golde, 1998). The *trans*-signaling activity conferred by sIL-6R has already been discussed (Jones et al., 2001), as well as its role in endothelial cell activation (Modur et al., 1997; Romano et al., 1997). By contrast, the soluble receptor subunit for another member of the IL-6 family, IL-11, has been found to antagonize IL-11 activity (Curtis et al., 1997). Likewise, the soluble form of the gp130 subunit shared by the IL-6 family of receptors is thought to inhibit IL-6 mediated signaling by binding the IL-6/sIL-6R complex (Narazaki et al., 1993; Muller-Newen et al., 1998; Jostock et al., 2001).

Antagonistic binding proteins have recently also been found for IL-13, IL-18, and IL-22 (Zhang et al., 1997; Xu et al., 1998b; Novick et al., 1999). IL-18-binding protein (IL-18bp) has been characterized as a modulator of the Th1 response on the basis of its ability to inhibit IL-18-mediated up-regulation of IFN $\gamma$ , IL-8, NF- $\kappa$ B, and VCAM-1 (Reznikov et al., 2000; Vidal-Vanaclocha et al., 2000). The human IL-18bp gene encodes at least four isoforms (Kim et al., 2000b), and its expression is increased by IFN $\gamma$  in a range of human cell lines (Muhl et

al., 2000). Serum levels of IL-18bp are raised in septic patients, with a concomitant decrease in free IL-18, and its roles are therefore presumed to include the provision of negative feedback in states of high inflammatory activity (Novick et al., 2001). Recently, Mallat et al. (2001b) have demonstrated the anti-atherogenic potential of IL-18bp. They have found electrotransfer of an expression plasmid encoding murine IL-18bp to attenuate atherosclerotic lesion development in the aorta of apoE<sup>-/-</sup> mice. This treatment also resulted in changes in plaque composition, comprising a decrease in inflammatory cell content and an increase of smooth muscle cell and collagen content of the lesion. IL-18bp therefore appears to have a beneficial effect on plaque stability as well as plaque progression. Moreover, IL-18bp may promote ischemia-induced neovascularization by inhibiting the anti-angiogenic role of IL-18, and its administration could therefore also aid postinfarction myocardial recovery (Mallat et al., 2002). Interestingly, poxvirus proteins have been identified that share considerable sequence homology with human IL-18bp. These inhibit virus elimination by the host's immune system by binding IL-18, attenuating IL-18-induced IFN $\gamma$  production, and impairing natural killer cell cytotoxicity (Born et al., 2000; Calderara et al., 2001) and may be promising anti-atherogenic agents in their own right. Similar protective functions appear to be served by the viral capture and modification of other cytokine receptor genes (Spriggs, 1996; McFadden et al., 1998), including the IL-1R (Spriggs et al., 1992) and IL-8R (Rosenkilde et al., 1999), and the chemokine binding proteins M-T1 and M-T7 (Upton et al., 1992; Graham et al., 1997; Lalani et al., 1997a). The latter is an IFN $\gamma$ R homolog and has been used successfully in the attenuation of angioplasty-induced neointima formation in rat carotid arteries (Liu et al., 2000). A 38-kDa glycopeptide encoded by the tanapox virus binds IL-2, IL-5, and IFN $\gamma$ , and inhibits the TNF $\alpha$ -induced expression of E-selectin, VCAM-1, and ICAM-1 by tanapox virus-infected primary endothelial cells (Paulose et al., 1998).

Other cytokines have also been targeted by soluble receptor therapy, the foremost example being the antagonism of TNF $\alpha$ . TNF $\alpha$  has been suggested to be pro-atherogenic by virtue of its presence in atherosclerotic lesions and its proinflammatory effects on all cell types involved in atherogenesis, including the up-regulation of adhesion molecules, chemoattractants, cytokines, and growth factors (LeBoeuf and Schreyer, 1998). Although systemic TNF $\alpha$  levels are not correlated with an increased propensity to atherosclerosis, the level of TNF $\alpha$  is an independent risk factor for the occurrence of acute coronary events in patients with coronary artery disease (Ridker et al., 2000a; Sack, 2002). Most importantly, however, TNF $\alpha$  levels are known to be raised in congestive heart failure and to exacerbate heart failure in murine models, probably due to excessive myocardial remodeling (Bradham et al., 2002).

TNFR-IgG fusion proteins have proven their worth in reducing the TNF $\alpha$ -mediated induction of proinflammatory interleukins, including IL-1 $\beta$  and IL-6 (Abraham et al., 1994; Kubota et al., 2000; Kadokami et al., 2001). Two TNF $\alpha$  blockers have recently been evaluated in clinical trials: etanercept (Enbrel), a fusion protein of the soluble form of the TNFR and the Fc portion of human immunoglobulin IgG1 and infliximab (Remicade), a chimeric IgG1 monoclonal antibody that contains a murine binding site for TNF $\alpha$ . Despite encouraging results in early clinical studies, in which subcutaneous etanercept administration appeared to be safe and to result in improvement of cardiac function in patients with advanced heart failure (Bozkurt et al., 2001), a large-scale phase II/III trial (RENEWAL) has recently been prematurely discontinued due to a lack of benefit (Louis et al., 2001). The introduction of infliximab as a therapy for rheumatoid arthritis, on the other hand, has been marred by the recent report of a case of sudden death in a patient without heart failure following a single 200 mg infusion (de' Clari et al., 2002). Moreover, a phase II clinical trial investigating the use of infliximab in advanced congestive heart failure has been placed on hold after the death of seven patients in the treatment group.

The experience with infliximab, in particular, may point to a potentially protective effect of TNF $\alpha$  in heart failure. Thus, TNF $\alpha$  has been found to induce protein synthesis in cardiac myocytes (Hiraoka et al., 2001) and to lead to inflammatory autoregulation by means of the translocation of functionally inactive NF- $\kappa$ B p50 homodimers (Haudek et al., 2001).

In an atherosclerotic context, blockade of TNF $\alpha$  by administration of soluble TNFR has been found to accelerate endothelial recovery after balloon angioplasty of rat carotid arteries (Krasinski et al., 2001). Because endothelial damage is thought to be an important process in atherogenesis and atherosclerotic plaque erosion, the inhibition of TNF $\alpha$ -mediated impairment of endothelial function could yield considerable merit. In an analogous approach, adenovirus-mediated transfer of a secreted TGF $\beta$  type II receptor has been demonstrated to inhibit luminal loss after percutaneous transluminal coronary angioplasty of porcine coronary arteries (Kingston et al., 2001). Specific targeting to inflammatory tissues may refine such gene transfer approaches, as demonstrated by gene vectors in which the TNFR-IgG fusion protein sequence has been placed under the control of a serum amyloid A promoter. Serum amyloid A levels increase dramatically in inflammatory conditions, and the plasmid-mediated expression of such a construct has been shown to be activated *in vitro* by IL-1 $\beta$  and TNF $\alpha$  (Rygg et al., 2001). Nonetheless, TNF $\alpha$  also exerts potentially anti-atherogenic functions, including the inhibition of lipoprotein lipase (Tengku-Muhammad et al., 1996) and the attenuation of macrophage scavenger receptor activity (van Lenten and Fogelman, 1992; Hsu et



al., 1996; Schreyer et al., 1996). In addition, TNF $\alpha$  deficiency has no effect on atherogenesis in apoE $^{-/-}$  mice (Schreyer et al., 2002), whereas TNFR1 deficiency even predisposes to atherosclerosis (Schreyer et al., 1996). As is the case with respect to heart failure, controversy thus still shrouds antagonism of TNF $\alpha$  activity as a treatment for atherosclerosis, which is also borne out by the fact that administration of TNF-binding protein in apoE $^{-/-}$  mice attenuates fatty streak formation in females, whereas it has no effect in male mice (Elhage et al., 1998).

Virus-encoded interleukin and interleukin receptor homologs are also thought to function as antigens and haptens, respectively, in the generation of autoantibodies against a series of interleukins that have been found to occur naturally in healthy humans and certain disease states (Bendtzen et al., 1998), including antibodies against IL-1 $\alpha$  (Bendtzen et al., 1989, 1994), IL-6 (Hansen et al., 1991; Bendtzen et al., 1994), and IL-10 (Bendtzen et al., 1994). The (patho)physiological role of these antibodies remains somewhat unclear, although most neutralize their target interleukins in vitro (Svenson et al., 1992; Hansen et al., 1993, 1995). Several have also been found to attenuate interleukin activity in vivo, and anti-cytokine therapy by means of monoclonal antibodies has also been investigated in the context of atherosclerosis. An important role has been assigned to CD40L-CD40 interactions in the pathogenesis of atherosclerosis (Mach et al., 1997). Accordingly, treatment with anti-CD40L antibody reduces de novo atherogenesis in atherosclerosis-prone mice (Mach et al., 1998) and cardiac allograft arteriopathy in a murine heterotopic cardiac transplant model (Wang et al., 2002) and has also been found to alter the histological appearance of pre-existing atherosclerotic lesions toward a more stabilized phenotype (Lutgens et al., 2000; Schonbeck et al., 2000). By contrast, antibody-mediated neutralization of TGF $\beta$  signaling accelerates atherogenesis in apoE $^{-/-}$  mice, and leads to the development of a more inflammatory plaque phenotype (Mallat et al., 2001c). Despite being a CD40-inducible protein (Zan et al., 1998), TGF $\beta$  thus appears to have anti-atherogenic properties, and its inhibition would therefore be undesirable in the prevention of atherogenesis.

The chronic nature of atherosclerosis and the generally rapid clearance of administered antibodies, however, would necessitate repeated parenteral administration to ensure prolonged efficacy. Eliciting an endogenous antibody response by immunization with the cytokine in question may circumvent this problem. A humoral immune response has previously been shown to be mounted against most therapeutically administered recombinant interleukin preparations (Revoltella, 1998), and this observation has paved the way for the introduction of intentional interleukin immunization. Svenson et al. (2000) have immunized mice with recombinant murine IL-1 $\alpha$  in conjunction with purified pro-

tein derivative of tuberculin, which resulted in the development of IL-1 $\alpha$  neutralizing autoantibodies that attenuate the expression of IL-6 in vivo. Alternatively, a synthetic interleukin receptor antagonist may be used as an antigen for the induction of autoimmunity against interleukins, as has been demonstrated for IL-6 (Ciapponi et al., 1997), or vaccination may be conducted with a DNA vaccine encoding antigenic epitopes of the cytokine concerned (Youssef et al., 1998). Thus, rats have been found to mount a protracted immune response to Fas ligand after a course of vaccinations with FasL cDNA (Wildbaum et al., 2000). The resulting autoantibodies inhibited the production of TNF $\alpha$  by cultured T lymphocytes in vitro and provided protection against experimental autoimmune encephalomyelitis in vivo.

In considering the therapeutic scope of humoral anti-interleukin immune response induction, however, one needs to take into account that some anti-cytokine antibodies have been found to stabilize cytokine functions rather than solely neutralizing their activity (Bendtzen et al., 1990; Wendling et al., 1993). Antibodies to IL-3, IL-4, and IL-7 have thus been demonstrated to form complexes with their target interleukins, which prolongs their in vivo half-life (Finkelman et al., 1993). This has led to the realization that the efficacy of monoclonal anti-interleukin therapy constitutes a balance between the neutralization avidity and the rate of clearance of the formed complex. These characteristics may partly depend on the specific epitope recognized by the antibody, and meticulous preclinical assessment of complex clearance is therefore indicated prior to clinical evaluation.

#### D. Interleukin Receptor Antagonists

Endogenous regulation of interleukin activity also occurs at the level of ligand-receptor interaction. A major exponent of this type of modulation is the control of IL-1 signaling by the endogenous IL-1 receptor antagonist, IL-1ra (Arend et al., 1998; Smith, 2000). First discovered in the 1980s (Arend et al., 1985), this factor has been extensively studied as a potential anti-inflammatory compound. Systemic treatment with IL-1ra has been proven to be beneficial in the treatment of rheumatoid arthritis in animal models and in humans, as judged by histological and clinical improvement (Bresnihan et al., 1998; Cunnane et al., 2001). As discussed above, IL-1ra has also been suggested as an important protective factor in atherogenesis (Francis et al., 1999) and restenosis (Kastrati et al., 2000), and its administration is currently under scrutiny as a potential anti-atherogenic therapy. Elhage et al. (1998) have demonstrated that subcutaneous injection of IL-1ra by means of an osmotic pump (25 mg/kg/day for 1 month) leads to a significant reduction in fatty streak formation in the aortic sinus of apoE $^{-/-}$  mice on an atherogenic diet (Elhage et al., 1998). Short-term treatment with IL-1ra has been found to be well tolerated. Due to the central role of IL-1 in the

immune response, however, long-term systemic treatment with an inhibitor of this factor may not be desirable. It is encouraging, therefore, that local gene therapeutic approaches involving IL-1ra have provided promising results in the attenuation of cerebral, pancreatic, and articular inflammation in animal models (Yang et al., 1997; Fernandes et al., 1999; Giannoukakis et al., 1999) and are currently awaiting evaluation in clinical trials (Del Vecchio et al., 2001).

In lieu of naturally occurring antagonists, inhibitors of interleukin receptors have also been developed by synthetic means. Phage display techniques have led to the development of AF12198, a 15-mer peptide with nanomolar affinity for the human type I IL-1 receptor, which does not bind to the human type II receptor (Akeson et al., 1996), and inhibits IL-1-induced ICAM-1 expression by endothelial cells in vitro. Moreover, it downregulates IL-6 induction in cynomolgus monkeys and is thus considered to be the first small molecule to show IL-1 receptor antagonist activity in vivo.

In general, the development of small interleukin receptor antagonists has proved difficult, however, due to the complex and multipoint high-affinity interactions between interleukin receptors and their ligands. A more rewarding strategy has been the mutation of existing ligands. IL-6 ligand-receptor interaction can be blocked by IL-6 variants that have been mutated to display increased affinity for IL-6R and decreased binding to gp130 (Sun et al., 1997; Devlin et al., 1998; Honemann et al., 2001). Due to their interference with gp130 interaction, these IL-6 receptor antagonists may also function as IL-11 antagonists (Sun et al., 1997). IL-12, in its active form, consists of two disulfide-bonded subunits, p40 and p35, and synthetic antagonists have been devised for human (Ling et al., 1995) and murine IL-12 (Gillesen et al., 1995) by homodimerization of the IL-12 p40 subunit. The p40 homodimer acts as a potent IL-12 antagonist in vitro, reduces the murine Th1 type response to endotoxin in vitro (Gately et al., 1996), and protects mice from septic shock following LPS injection (Mattner et al., 1997). Considering the importance that has been assigned to Th1-mediated processes in atherosclerosis, this approach may also hold promise in the prevention of atherogenesis.

The possibility of attenuating interleukin binding by introducing a blocking antibody response to its receptor has also been explored. In vitro, binding of IL-2 to IL-2R can be inhibited by the addition of humanized antibodies that are bispecific for anti-IL-2 receptor  $\alpha$  and  $\beta$  (Pilson et al., 1997). In addition to its inhibitory activity on IL-2 signaling, this antibody displays activity against IL-15, possibly by virtue of competing for the shared IL-2 $\beta$  receptor subunit. In a monkey model of autoimmune uveitis, this antibody has been demonstrated to markedly reduce inflammation after twice-weekly intravenous injections for 4 weeks (Guex-Crosier et al., 1997). Furthermore, antagonism of IL-2 by means of anti-

IL-2R antibodies, including the commercial preparations basiliximab and dacluzimab, has proven an effective addition to the immunosuppressive regimen following renal allograft transplantation (Vincenti et al., 1998; Onrust and Wiseman, 1999). This therapeutic efficacy is believed to also be partly due to inhibition of IL-15-mediated responses (Boelaars-van Haperen et al., 2001). With respect to other interleukins, antibody blockade of IL-4R and IL-6R has been found to alleviate antigen-induced airway hyperresponsiveness and collagen-induced arthritis, respectively (Gavett et al., 1997; Takagi et al., 1998; Mihara et al., 2001), and antibody directed at IL-18R reduces LPS-induced inflammation and mortality in mice (Xu et al., 1998a).

Opsionization and complement activation are believed to contribute to the mechanism of action of interleukin-receptor antibodies, and the ensuing elimination of cells expressing the relevant receptor may attenuate inflammatory pathways elicited by interleukin binding. In analogy, the specificity of interleukin binding has been employed in devising a "Trojan horse" strategy for the targeting of cytotoxic compounds. This entails the administration of fusion proteins consisting of an interleukin and a toxic polypeptide domain, as used in the transfer of pseudomonas exotoxin to IL-4R-expressing breast carcinoma cells (LeMaistre et al., 1998) and of diphtheria toxin to IL-2R-expressing lymphomas (Leland et al., 2000). Significant toxic side effects may limit this type of therapy to acutely life-threatening and incurable diseases (Bagel et al., 1998). This would almost certainly exclude atherosclerosis as a candidate ailment, although it could be applicable in a short-term strategy for the prevention of restenosis following angioplasty of atherosclerotic lesions. Thus, it is interesting to note that Miller et al. (1996) have found atherosclerotic vascular thickening in rabbits following aortic balloon angioplasty to be reduced by an interleukin-2 receptor-specific fusion protein, termed DAB<sub>486</sub>-IL-2, in which the receptor binding domain of diphtheria toxin had been replaced by a human IL-2 sequence. DAB<sub>486</sub>-IL-2 was administered for 10 days following angioplasty (0.1 mg/kg/day i.v.), found to be well tolerated for the duration of the experiment, and to result in complete inhibition of lesion formation compared with controls.

#### *E. Up-Regulation of Anti-Inflammatory Interleukins*

As has been discussed in a previous section, several interleukins have been ascribed a putative anti-atherogenic role, including IL-9, IL-10, IL-11, and potentially IL-4 and IL-13 (Table 1). All of these cytokines are known to induce a Th2 type cytokine response, and have been implicated in the pathogenesis of Th2-mediated diseases (Barnes, 2001a). Consistently, their inhibition has been suggested as a potential treatment for these conditions, including asthma (Henderson et al., 2000; Barnes 2001b; Zhou et al., 2001b). Overexpression of these interleukins, on the other hand, has been specu-

lated to ameliorate a variety of Th1-mediated inflammatory conditions, such as rheumatoid arthritis, septic shock, and atherosclerosis. Their up-regulation may therefore hold promise as a therapeutic modality in these diseases, and several studies to this effect have been reported.

Endotoxin-elicited shock has been used as a model for the evaluation of the role and the therapeutic potential of all of these interleukins in Th1-mediated inflammation. The protective effect of IL-4 has been studied in a murine model of Gram-negative septic shock following *Pseudomonas aeruginosa* infection (Giampietri et al., 2000). Mortality was found to be reduced by IL-4 treatment, correlating with a decrease in TNF $\alpha$  elaboration. Similarly beneficial effects have been found after prophylactic injections of recombinant IL-9 in this model (Grohmann et al., 2000). This effect is accompanied by a reduction in TNF $\alpha$ , IL-12 p40, and IFN $\gamma$  levels, and appears to be IL-9-specific, as heat-inactivated IL-9 did not improve survival rates. Circulating IL-10 levels were found to be markedly augmented by IL-9 injection, and this may be partly responsible for an indirect suppression of proinflammatory cytokine expression, as IL-10 itself also reduces TNF $\alpha$  production and lethality in murine endotoxemia (Gerard et al., 1993). Conversely, IL-9 production in mast cells is greatly stimulated by IL-10, closing a potent positive feedback loop (Stassen et al., 2000). IL-11, on the other hand, inhibits LPS-induced up-regulation of TNF $\alpha$ , IL-1 $\beta$ , and IFN $\gamma$  by an IL-10-independent mechanism in vivo, and has been found to result in a 60% inhibition of LPS-induced elaboration of TNF $\alpha$ , IL-1 $\beta$ , IL-12 p40, and nitric oxide by murine peritoneal macrophages in vitro (Trepicchio et al., 1997). Moreover, IL-11 reduces lung TNF $\alpha$  levels and neutrophil sequestration, and improves pulmonary vasomotor function in a model of LPS-induced lung injury (Sheridan et al., 1999). IL-13 even leads to a paradoxical decrease in IL-10 levels following intraperitoneal LPS injection, despite TNF $\alpha$ , IFN $\gamma$ , and IL-12 attenuation, and is therefore also presumed to exert its protective effect in endotoxemic shock through an IL-10-independent pathway (Muchamuel et al., 1997).

The chondroprotective and anti-colitic properties of anti-inflammatory interleukins have also been evaluated. Locally applied recombinant human IL-4 and IL-10 attenuated cartilage degradation and mononuclear cell activity in human rheumatoid synovium that had been engrafted subcutaneously to SCID CB17 mice. Moreover, IL-10, but not IL-4, decreased the expression of ICAM-1 by synovial cells in this model (Jorgensen et al., 1998). IL-11 also significantly reduced the severity of collagen-induced arthritis in mice (Walmsley et al., 1998), and possibly of rheumatoid arthritis in humans (Moreland et al., 2001). In a rat model of inflammatory bowel disease, intraperitoneal adenoviral transfer of IL-4 has been found to significantly inhibit tissue dam-

age, serum and colon IFN $\gamma$  levels, and myeloperoxidase activity in the distal colon (Hogaboam et al., 1997).

Surprisingly little is known about the atheroprotective role of these interleukins, and IL-10 is undoubtedly the most extensively studied candidate in this respect. A relative deficiency of IL-10 signaling has been implicated in the pathogenesis of a variety of chronic autoimmune conditions, including rheumatoid arthritis, Crohn's disease, multiple sclerosis, and psoriasis. Promising results have been obtained in studies addressing the therapeutic potential of IL-10 administration in animal models of these diseases (Croxford et al., 1998; Kim et al., 2000a; Lubberts et al., 2000), and the outcomes of early clinical trials have been encouraging with respect to safety and efficacy, but these require confirmation on a larger scale (van Deventer et al., 1997; Asadullah et al., 1999; Colombel et al., 2001). The advantageous potential of IL-10 in dampening the inflammatory background of atherosclerosis is strongly suggested by several in vitro and animal studies (Terkeltaub, 1999). Thus, atherogenesis is decreased in IL-10 transgenic mice on a high-fat diet, whereas IL-10 knockout (IL-10 $-/-$ ) mice display an increased atherogenic tendency (Pinderski-Oslund et al., 1999), which is ameliorated by plasmid-mediated transfer of IL-10 (Mallat et al., 1999b). Furthermore, transfer of bone marrow from IL-10 transgenic mice to LDLr $-/-$  mice inhibits atherosclerosis by altering the phenotype of the resident lymphocyte and macrophage populations in the atherosclerotic plaque (Pinderski et al., 2002).

We have recently demonstrated that de novo collar-induced atherogenesis in LDLr $-/-$  mice (von der Thüsen et al., 2001b) is inhibited by adenovirus-mediated overexpression of human IL-10 (hIL-10), following both systemic and local transfer (von der Thüsen et al., 2001a) (Fig. 5). Although we found overexpression of hIL-10 to be immunomodulatory, as evidenced by monocyte deactivation, it also resulted in marked serum cholesterol lowering. The anti-atherogenic effect of systemic hIL-10 may therefore be considered to be bipartite in this hypercholesterolemic animal model. Local immunomodulation, however, is thought to be solely responsible for the attenuation of atherosclerotic plaque formation (44.9%,  $P < 0.05$ ) that was observed after in vivo endothelial hIL-10 transduction with the same vector. We have used a similar approach in the evaluation of IL-9 as an atheroprotective agent and found daily injections of IL-9 protein (1  $\mu$ g/mouse/day i.p.) for 5 weeks to reduce carotid collar-induced atherosclerosis by 65% in LDLr $-/-$  ( $P < 0.01$ ) (Kuiper et al., 2001). An explanation for this finding may lie in the IL-9-mediated up-regulation of inhibitors of interleukin signaling, in addition to its enhancement of IL-10 production (Lejeune et al., 2001).

The atheroprotective nature of IL-10 cannot be considered to be a foregone conclusion, as local injection of an IL-10 expression plasmid inhibits angiogenesis in a

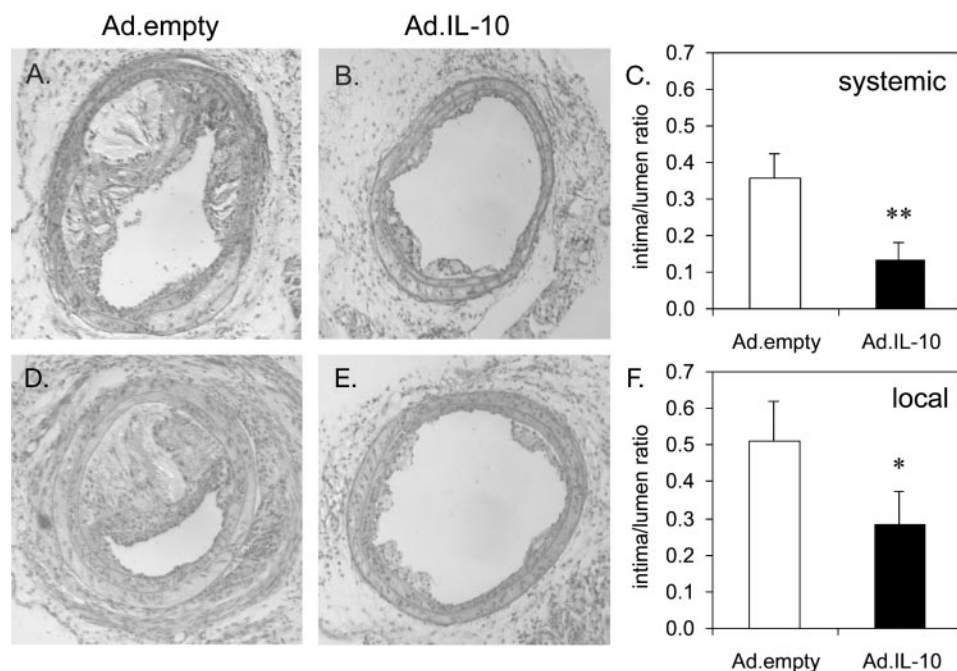


FIG 5. Effects of adenovirus-mediated gene transfer of the anti-inflammatory cytokine IL-10 on atherosclerosis in LDLr<sup>-/-</sup> mice. Both systemic (hepatic) (A–C) and local (endothelial) (D–F) overexpression result in a decrease in atherosclerotic plaque surface area and complexity, which is reflected by a marked attenuation of the degree of stenosis (C+F) \*,  $P < 0.05$ ; \*\*,  $P < 0.02$ . Adapted from von der Thüsen et al., 2001a.

mouse model of hindlimb ischemia (Silvestre et al., 2000), and administration of IL-10 protein augments arterial disease in murine heart transplants (Furukawa et al., 1999). The effect of IL-10 application, as a protein or following gene transfer, may eventually be found to depend on the stage of the disease, the mode of transfer and the dosing regimen. Furthermore, it may be possible to tailor the pleiotropic actions of IL-10 to the use as an anti-atherogenic agent by the use of viral IL-10 homologs. Thus, the Epstein-Barr virus BRCF-1 gene product (vIL-10) has been found to share 84% amino acid sequence identity but only a limited number of the pleiotropic actions of hIL-10. Perhaps most importantly, it lacks the immunostimulatory properties of human IL-10, while sharing its inhibitory activity with respect to cytokine synthesis and macrophage activation (Ding et al., 2000). Furthermore, vIL-10 has been found to lead to augmented and more prolonged expression following adenovirus-mediated transfer in mice in comparison with its human counterpart (Minter et al., 2001), and to effectively reduce endothelial expression of E-selectin, P-selectin, and ICAM-1 in rats following adenovirus-mediated transfer (Henke et al., 2000). The application of vIL-10 may eventually prove to be preferable to hIL-10 if treatment is primarily aimed at providing an anti-inflammatory stimulus, as is the case in the prevention of atherosclerosis.

#### F. Inhibition of Interleukin Signaling

The effector functions of all interleukins depend on the activation of intracellular signaling cascades involving, *inter alia*, Jaks, Tyks, and STATs (Leonard and Lin,

2000; Touw et al., 2000). These pathways are negatively regulated by endogenous signaling inhibitors, including the SH2-containing phosphatase, SOCS, and protein inhibitor of activated STAT families (Chung et al., 1997; Starr et al., 1997; Liu et al., 1998; Naka et al., 1999), of which the expression is partly controlled by interleukins themselves. These are considered to play a pivotal role in the cross-regulation of interleukin function, as Th2 cytokines have been found to lead to the expression of negative regulators of Th1 cytokines, and vice versa. Moreover, interleukins may also up-regulate inhibitors of their own signaling cascades and are therefore subject to negative feedback loops. Thus, IL-4 activity is controlled by SOCS-1 (SSI-1), which is elaborated in response to interferons as well as IL-4 itself (Naka et al., 1997; Dickensheets et al., 1999; Losman et al., 1999), and the immunosuppressive and autoregulatory effects of IL-9, IL-10, and IL-11 are thought to be partly mediated by the up-regulation of SOCS-3, which inhibits STAT5-mediated signaling (Auernhammer and Melmed, 1999; Cassatella et al., 1999; Donnelly et al., 1999; Lejeune et al., 2001).

The administration of inhibitors of cytokine signaling could have beneficial effects in atherosclerosis. The essential role of tyrosine kinases in cytokine signaling has prompted the evaluation of tyrosine kinase inhibitors as therapeutic agents. In this respect, a group of compounds called “tyrphostins” has been shown to have anti-proliferative and anti-inflammatory properties *in vitro* and *in vivo* that are thought to be mediated by tyrosine kinase inhibition (Levitzki, 1990). Platelet-derived growth factor is among the cytokines to be inhib-

ited by the tyrphostins, and these have therefore been speculated to be effective against smooth muscle cell-mediated pathological processes. The latter includes injury-induced neointima formation, and application of the tyrphostin AG-17 by means of a perivascular controlled release implant has been found to inhibit intimal hyperplasia in injured rat carotid arteries (Golomb et al., 1996). In addition, Huynh et al. (1998) have found *ex vivo* incubation of jugular veins with AG-51 to reduce post-operative intimal hyperplasia by 49%, following their placement as an interposition graft in rabbit carotid arteries. A relative lack of pharmacological and functional selectivity, however, may limit the applicability of these inhibitors as immunomodulatory compounds aimed at diverting the cytokine response from a Th1 to a Th2 expression pattern. SB 203580, for example, in addition to its originally described inhibitory activity for p38 MAPK, also attenuates stress-activated protein kinases and c-Jun N-terminal kinases (Cuenda et al., 1995; Clerk and Sugden, 1998). SB 203580 inhibits TNF and IL-1 expression and protects mice from collagen-induced arthritis (Owens and Lumb, 2000) but also attenuates IL-4, IL-5, and IL-13, and virtually blocks IL-10 production (Koprak et al., 1999). The latter effect, in particular, is considered undesirable in the setting of atherosclerosis. It is conceivable, however, that specificity of protein kinase inhibition may in the future be achieved by the transfer of endogenous inhibitors, including SOCSs by gene therapy or protein administration. As an example, plasmid-mediated overexpression of SOCS-1 has recently been found to inhibit cytokine-induced CD40 expression in macrophages by blocking IFN $\gamma$ -mediated STAT-1 $\alpha$  activation (Wesemann et al., 2002).

#### G. Inhibition of Interleukin-Induced Gene Expression

Interleukin signaling eventually culminates in the sequence-specific binding of DNA by activated transcription factors and the ensuing up-regulation of target gene transcription. Some of these transcription factors are considered potentially rewarding substrates for immunomodulatory therapy, including NF- $\kappa$ B, AP-1 and the STAT proteins, by virtue of their capacity to integrate converging signals from various proinflammatory cytokines and other inflammatory stimuli (Collins, 1993; Touw et al., 2000; Tedgui and Mallat, 2001). Several methods have been employed in the attenuation of transcription factor activity, based on the characteristic bispecific affinity of these molecules for regulatory proteins and DNA.

The inhibition of vascular NF- $\kappa$ B-regulated transcription, in particular, is presumed to hold anti-atherogenic potential. Activated NF- $\kappa$ B has been identified in smooth muscle cells, macrophages, and endothelial cells in the atherosclerotic lesion (Brand et al., 1996). Functional significance for NF- $\kappa$ B in atherogenesis has been deduced from its colocalization with the expression of

NF- $\kappa$ B target genes in plaques (Brand et al., 1996) and the association of its expression in coronary atherosclerotic lesions with unstable angina (Wilson et al., 2002). The role of NF- $\kappa$ B as a causal mediator in atherosclerosis remains unclear, however, which is partly due to the intrauterine lethality associated with p65- and I $\kappa$ B $\alpha$ -deficiency in mice (Collins and Cybulsky, 2001). The regulation of NF- $\kappa$ B activity depends on the extent of binding to its naturally occurring inhibitors, including I $\kappa$ B $\alpha$ , I $\kappa$ B $\beta$ , I $\kappa$ B $\gamma$ , and BCL3 (Ghosh and Baltimore, 1990; Finco and Baldwin, 1995). Phosphorylation of I $\kappa$ B by the I $\kappa$ B kinase (IKK) complex, containing IKK $\alpha$ , IKK $\beta$ , and IKK $\gamma$  (NEMO), leads to I $\kappa$ B ubiquitination and proteasome-mediated degradation. This enables the nuclear translocation of unbound NF- $\kappa$ B and subsequent activation of NF- $\kappa$ B-dependent transcription. In endothelial cells, NF- $\kappa$ B is thought to play an essential role in the regulation of adhesion molecule expression in response to inflammatory stimuli, including cytokines (Collins et al., 1995; De Caterina et al., 2001). The endothelial NF- $\kappa$ B/I $\kappa$ B system is presumed to be primed in endothelial cells in lesion-prone arterial sites, as evidenced by increased expression of the p65 (RelA) NF- $\kappa$ B subunit, I $\kappa$ B $\alpha$ , and I $\kappa$ B $\beta$ , prior to plaque development and NF- $\kappa$ B activation (Hajra et al., 2000). The attenuation of I $\kappa$ B activity by IKK up-regulation has been identified as a pivotal step in endothelial activation (Read et al., 1994; Bennett et al., 1996; Johnson et al., 1996), whereas the inhibition of endothelial adhesion molecule expression by nitric oxide has been found to be mediated by I $\kappa$ B $\alpha$  (Spiecker et al., 1997). The recognition of the physiological importance of this inhibitory pathway has prompted the evaluation of the anti-atherogenic properties of I $\kappa$ B $\alpha$  administration. Adenoviral transfer of I $\kappa$ B $\alpha$  has thus been found to effect down-regulation of inflammatory genes in endothelial cells, including VCAM-1, IL-1, IL-6, and IL-8 (Wrighton et al., 1996), and to lead to inhibition of monocyte adhesion and transmigration on TNF $\alpha$ -activated endothelium (Weber et al., 1999). In addition, TNF $\alpha$ -induced endothelial expression of adhesion molecules (E-selectin and ICAM-1) and chemokines (MCP-1) is attenuated by retrovirus-mediated introduction of a proteolysis-resistant I $\kappa$ B $\alpha$  mutant, I $\kappa$ B $\Delta$ N, and the addition of pharmacological inhibitors of I $\kappa$ B $\alpha$  phosphorylation and proteasome degradation (Cobb et al., 1996; Pierce et al., 1997; Lockyer et al., 1998; Hipp et al., 2002).

NF- $\kappa$ B may also modulate atherogenesis by regulating the transcription of inflammatory genes in monocytes/macrophages and smooth muscle cells (Ghosh and Baltimore, 1990; Bourcier et al., 1997). In macrophages, the LPS-stimulated production of proinflammatory cytokines and inducible nitric-oxide synthase (iNOS) have been found to be reduced by adenoviral overexpression of I $\kappa$ B $\alpha$  and the administration of proteasome inhibitors, respectively (Griscavage et al., 1996; Bondeson et al., 1999). In vascular smooth muscle cells, liposomal deliv-

ery of purified I $\kappa$ B $\alpha$  peptide attenuates TNF $\alpha$ -induced proliferation (Selzman et al., 1999), and overexpression of I $\kappa$ B $\alpha$  diminishes the elaboration of the matrix metalloproteinases MMP-1, MMP-3, and MMP-9, which may have plaque-stabilizing consequences *in vivo* (Bond et al., 2001). In addition, decoy oligonucleotides to NF- $\kappa$ B binding sites have been used to counteract NF- $\kappa$ B-mediated transcriptional activation and have displayed effectiveness in inhibiting graft coronary artery disease of rat cardiac allografts following *ex vivo* pressure-mediated delivery (Feeley et al., 2000). Other interleukin-activated transcription factors to have been successfully inhibited *in vitro* by the decoy approach include STAT1 (Ohtsubo et al., 2000), STAT6 (Wang et al., 2000), and AP-1 (Morishita et al., 1998). Inhibition of AP-1-mediated transcription, in particular, effectively reduced joint destruction in a murine model of collagen-induced arthritis (Shiozawa et al., 1997).

Attenuation of NF- $\kappa$ B activity is also presumed to constitute a physiological feedback mechanism in inflammatory homeostasis. Thus, several potentially anti-atherogenic interleukins reduce NF- $\kappa$ B activity by variably increasing I $\kappa$ B $\alpha$  transcription (IL-4) (Donnelly et al., 1993; Abu-Amer, 2001), preventing I $\kappa$ B degradation (IL-10 and IL-13) (Lentsch et al., 1997) or increasing the expression of BCL3, a protein with close homology to I $\kappa$ B proteins (IL-4 and IL-9) (Richard et al., 1999). Interestingly, NF- $\kappa$ B inhibition has evolved as a viral strategy of immune response evasion, exemplified by the adenovirus-encoded E1A protein (Kalvakolanu, 1999). The up-regulation of IL-6 by TNF $\alpha$  and IL-1 is inhibited by E1A due to its prevention of NF- $\kappa$ B p65-p50 heterodimer formation; although this leaves monomeric p50 to bind to the  $\kappa$ B element in the IL-6 promoter, this does not induce transcription (Janaswami et al., 1992). Moreover, E1A negatively regulates Stat1, Stat2, and Stat3 activity, and thereby attenuates IL-6-mediated gene expression (Takeda et al., 1994). An I $\kappa$ B homolog, A238L, is encoded by the African swine fever virus, and this has been shown to inhibit the production of proinflammatory cytokines in macrophages, allowing persistent viral infection (Powell et al., 1996). It may prove possible to exploit these anti-inflammatory traits in the prevention of atherosclerosis by overexpression or protein administration of the interleukins or viral proteins concerned.

Synthetic compounds with inhibitory activity for NF- $\kappa$ B have also been described. High throughput cell-based screening has led to the discovery of SP100030, a T cell-specific NF- $\kappa$ B and AP-1 inhibitor (Gerlag et al., 2000). SP100030 attenuates IL-2, IL-8, and TNF $\alpha$  production in T cell lines and alleviates disease progression in a murine model of collagen-induced arthritis. Finally, it has recently transpired that the pharmacological effects of several anti-inflammatory compounds, including salicylates, are partly derived from their inhibition of I $\kappa$ B phosphorylation and degradation (Schwenger et al., 1998; Young, 1998). This knowledge may aid the devel-

opment of derivatives of these drugs that are specifically targeted toward the inhibition of cytokine-induced inflammation.

Once interleukin-mediated transcription of inflammatory genes has occurred, antisense technology could be employed to interfere specifically with their translation. Interleukin-1 stimulated up-regulation of granulocyte-macrophage and granulocyte colony-stimulating factor gene expression in endothelial cells has been successfully inhibited by antisense ODNs (Segal et al., 1992). Moreover, the expression of endothelial adhesion molecules can be inhibited by the application of phosphorothioate oligonucleotides directed against ICAM-1, VCAM-1, and E-selectin (Bennett et al., 1994). Antisense-mediated down-regulation of endothelial ICAM-1 expression on monocytes reduces endothelial adhesiveness for leukocytes, which may be advantageous in atherogenesis (Steidl et al., 2000). The applicability of ICAM-1 inhibition by means of antisense has been demonstrated *in vivo*, as it has been shown to be effective in the prevention of cardiac allograft or lung isograft failure in mice and rats, respectively (Stepkowski et al., 1994; Toda et al., 2000). In clinical studies, the phosphorothioate ICAM-1 antisense preparation ISIS 2302 has been found to be well tolerated and to significantly lower the need for steroid treatment in Crohn's disease (Yacyszyn et al., 1998).

The previously mentioned caveats that apply to antisense therapy in general (Lebedeva and Stein, 2001) are evidently also poignant with respect to its use in the inhibition of interleukin-induced gene expression. The doubts that have been raised about the sequence-specific nature of ODN-mediated effects, the poor transfection efficiency, and the short half-life of ODNs *in vivo* will need to be addressed to warrant their applicability in the prevention of atherosclerosis.

#### IV. Discussion

Cytokines are being increasingly recognized as a potentially rewarding therapeutic target in a wide variety of diseases. For example, of the over 600 clinical gene therapy trials currently completed, ongoing or pending worldwide, those concerned with the transfer of cytokine genes constitute the largest category (Gene Therapy Clinical Trials, 2002). Most of these involve the application of immunostimulatory cytokines for the treatment of neoplastic and infectious diseases. Of the protocols addressing vascular diseases (51 in total), the overwhelming majority is intended to stimulate revascularization in peripheral and coronary ischemia by cytokine overexpression, largely employing the angiogenic growth factors fibroblast growth factor, PDGF, and vascular endothelial growth factor. While these hold promise for the treatment of atherosclerosis-related ischemia, it will have transpired from the preceding discussion that cytokine-directed therapy in general, and interleu-

kin-based treatment specifically, is still in its infancy as a means for the prevention of the onset and progression of atherogenesis per se. A lack of understanding of their involvement in atherogenesis currently prevents the use of some interleukins as targets for immunomodulation, including the members of the IL-10 family IL-19, IL-20, and IL-22 (Table 1). Other interleukins are overtly pleiotropic in their actions, and attenuating or augmenting their effects may be detrimental or beneficial, depending on the stage of atherosclerosis (IL-4, IL-13). Yet others have been attributed primarily anti- (IL-1ra, IL-9, IL-11, IL-10) or pro-atherogenic (including IL-1, IL-2, IL-6, IL-18) properties, and their modulation could therefore represent the most readily applicable approach to immunotherapy in atherosclerosis. This type of therapy may prove to be an effective alternative to currently used treatment protocols (e.g., lipid-lowering drugs) but could also be useful as an adjunctive to conventional pharmacotherapy. It is possible to conceive of several obstacles that may have impeded the development of such immunomodulatory strategies. Some of these are related to specific pathogenic features associated with atherosclerosis, others to the systemic and local consequences of immunomodulation, and yet others to purely technical aspects of interleukin therapy.

The chronic nature of atherosclerosis has doubtlessly hampered the evolution of adequate disease prevention strategies in general and also remains a significant obstacle to the preventive use of interleukin-based treatments. In considering the relative benefit of long-term use of the latter, one needs to pay attention to its cost, the practicality of its dosing regimen, and, most importantly, potential side effects.

All interleukins possess roles that are certainly not restricted to atherosclerosis, and their actions are frequently pivotal to several aspects of the immune system. Interleukins orchestrate defense mechanisms against a wide range of pathogens and tumor cells, in addition to playing a key role in various forms of nonimmune inflammation, and undiscerning diversions of the interleukin response will therefore invariably compromise one or more of these functions. For instance, whereas the inhibition of signaling by IL-2, IL-6, and IL-12 may be beneficial in the context of atherosclerosis, these factors have been implicated as potent antitumor agents (Maini et al., 1997), and attenuation of IL-2 signaling, in particular, may increase the risk of neoplasia. Conversely, Th2 cytokines are considered to have anti-atherogenic potential, but their role in the pathogenesis of autoimmune diseases is also well documented. Prolonged up-regulation of these factors, although tolerable in the short-term, may have deleterious consequences for the development or progression of *inter alia*, asthma, diabetes mellitus, systemic lupus erythematosus, and rheumatoid arthritis (Lafaille, 1998; Romagnani, 2000). Thus, whereas inhibition of atherogenesis in murine models has been achieved by application of the IL-1

antagonist IL-1ra (Elhage et al., 1998), the anti-inflammatory interleukin IL-10 (von der Thüsen et al., 2001a), and the interleukin-binding protein IL-18bp (Mallat et al., 2001b), these treatments still require long-term toxicological evaluation before beginning clinical trials. This type of untargeted systemic immunomodulation may eventually be limited to short-term treatments aimed at, for instance, the induction of regression or stabilization of existing atherosclerotic plaques. Proof-of-principle data to this effect have been obtained in ApoE<sup>-/-</sup> mice, in which the administration of antibodies to the cytokine CD40L has been seen to result in a stabilized plaque phenotype (Lutgens et al., 2000; Schonbeck et al., 2000). These studies indicate the potential benefits of short-term immunomodulatory treatment, and could serve as a paradigm for the development of similar strategies in humans.

For prolonged treatment, it may be desirable to restrict the action radius of therapeutic compounds to the atherosclerotic lesion and/or to ensure specificity of action for the atherosclerotic process. This will require the identification of marker molecules and cytokine signaling pathways, which are more or less specific for atherosclerosis, and these efforts may be greatly aided by the advent of DNA array and phage display technology (Faber et al., 2001; Houston et al., 2001; Monajemi et al., 2001). Thus, employing phage display techniques, we have recently identified a peptide sequence that specifically binds human P-selectin (Molenaar et al., 2001). This adhesion molecule is up-regulated on the endothelium of atherosclerosis-prone sites, and high affinity ligands for P-selectin may therefore serve as efficient tools for the targeting of viral and nonviral drug delivery vehicles to the developing atherosclerotic plaque. Such techniques may eventually also be extended to the targeting of specific cellular subsets in the atherosclerotic lesion to enhance therapeutic efficacy and reduce the risk of bystander effects.

Alternatively, site-directed targeting could be achieved by mechanical means. The development of local application catheters has recently been intensified, opening up possibilities for the intravascular instillation of therapeutic compounds. Due to the invasive nature of such techniques, this approach will demand preparations with an extended duration of action or therapeutics that have a lasting effect on atherogenesis or restenosis even with a single dosage regimen. Viral expression vectors may be used in achieving prolonged up-regulation of anti-inflammatory interleukins, such as (v)IL-10 (Kim et al., 2000a; Minter et al., 2001; von der Thüsen et al., 2001a), interleukin antagonists, such as IL-1ra and soluble TNF receptor (Giannoukakis et al., 1999; Kim et al., 2001), inhibitors of NF- $\kappa$ B signaling, such as I $\kappa$ B $\alpha$  (Wrighton et al., 1996; Bondeson et al., 1999; Weber et al., 1999), and antisense oligonucleotides and ribozymes directed against proinflammatory interleukins and interleukin-induced genes. Although ex-

tended transgene expression has been found to occur with some adenoviruses, including Ad-IL-10 (>200 days, unpublished data), the use of AAVs or Ad/AAV hybrids may be preferable in accomplishing this goal (Lynch et al., 1997; Recchia et al., 1999; Monahan and Samulski, 2000).

Barring the development of preparations with an extended duration of action, however, repeated administration will continue to be required to sustain therapeutic efficacy. This may elicit a humoral response to the protein concerned, which could severely compromise its potency and aggravate side effects. A further drawback of repeated administration is the fact that most currently available preparations require parenteral administration, which limits their tolerability and thus the likelihood of patient compliance. Paradoxically, to reduce the need for repeated administration, it may be possible to induce long-term immunomodulation by deliberately opting for active immunization by viral or nonviral means. The possibility of (DNA) vaccination as a method of raising neutralizing antibody responses against inflammatory interleukins has been discussed (Revoltella, 1998; Svenson et al., 2000), as has the possibility of interleukin stabilization and half-life extension by these antibodies (Finkelman et al., 1993). It should be noted, however, that extended duration of effectivity could also be regarded as a disadvantage, due to the relative irreversibility of such therapies in case of the occurrence of deleterious side effects.

The use of smaller synthetic compounds may reduce the need for parenteral administration and could therefore constitute a practical alternative to the transfer of entire interleukin molecules or anti-interleukin (receptor) antibodies. The examples discussed in this review include inhibitors of interleukin processing (Livingston, 1997), tyrosine kinase activity (Golomb et al., 1996; Huynh et al., 1998), proteasome function (Bondeson et al., 1999; Richard et al., 1999), p38 MAPK (Cuenda et al., 1995; Clerk and Sugden, 1998), and NF- $\kappa$ B (Gerlag et al., 2000). A drawback of many of these drugs, however, is their lack of pharmacological and functional specificity, partly due to the involvement of their molecular targets as downstream mediators in convergent signaling cascades, which is perhaps best exemplified by NF- $\kappa$ B. Careful toxicological evaluation will therefore be required before their clinical introduction. The use of (modified) endogenous inhibitors, including SH2-containing phosphatase, SOCS, and protein inhibitor of activated STAT, may eventually provide the required selectivity.

The production of interleukins and most of their inhibitors is currently a rather costly undertaking. Despite recent progress in recombinant protein production technology and therapeutic antibody expression technology (Maini et al., 1997), this situation is unlikely to change in the foreseeable future, making widespread prophylactic protein treatment prohibitively expensive.

With a view to these health economic implications and the minimization of potential side effects, it is imperative that treatment be confined to susceptible patients. Accurate tools for the identification of patients who may benefit most from such therapies, are therefore required. Refinement of genetic, biochemical, and radiological markers of predisposition to (complications of) atherosclerosis may provide important prognostic clues. The discovery of correlations between atherosclerotic events and interleukin-related polymorphisms, in particular, including those found for IL-1 (Momiya et al., 2001), IL-1ra (Francis et al., 1999, 2001; Kastrati et al., 2000), and IL-6 (Rauramaa et al., 2000; Georges et al., 2001), may facilitate the identification of suitable patients, whereas improved magnetic resonance imaging and ultrasound imaging of existing plaques will provide an impetus for the noninvasive determination of plaque "vulnerability" to rupture (Fayad and Fuster, 2001; Choudhury et al., 2002).

In summary, the currently available methods of modulation of interleukin-mediated inflammatory pathways are not yet suited to the widespread prevention of atherosclerosis. Substantial investigative efforts are still required with respect to target identification and the definition of suitable patient populations. Technical aspects of compound specificity, duration of action, and mode of transfer await additional improvement, but the first promising signs are looming on the horizon, because several techniques have been successfully validated in animal models. The initial aims of such therapies are likely to include the lasting stabilization of pre-existing plaques by short-term cytokine immunomodulation, which possibly represents the most readily achievable objective in clinical practice in the near future.

#### References

- Aarvak T, Chabaud M, Miossec P, and Natvig JB (1999) IL-17 is produced by some proinflammatory Th1/Th0 cells but not by Th2 cells. *J Immunol* **162**:1246–1251.
- Abraham E, Coulson WF, Schwartz MD, and Allbee J (1994) Effects of therapy with soluble tumour necrosis factor receptor fusion protein on pulmonary cytokine expression and lung injury following haemorrhage and resuscitation. *Clin Exp Immunol* **98**:29–34.
- Abu-Amer Y (2001) IL-4 abrogates osteoclastogenesis through STAT6-dependent inhibition of NF- $\kappa$ B. *J Clin Invest* **107**:1375–1385.
- Akeson AL, Woods CW, Hsieh LC, Bohnke RA, Ackermann BL, Chan KY, Robinson JL, Yanofsky SD, Jacobs JW, Barrett RW, and Bowlin TL (1996) AF12198, a novel low molecular weight antagonist, selectively binds the human type I interleukin (IL)-1 receptor and blocks in vivo responses to IL-1. *J Biol Chem* **271**:30517–30523.
- Akira S and Kishimoto T (1996) Role of interleukin-6 in macrophage function. *Curr Opin Hematol* **3**:87–93.
- Albanesi C, Cavani A, and Girolomoni G (1999) IL-17 is produced by nickel-specific T lymphocytes and regulates ICAM-1 expression and chemokine production in human keratinocytes: synergistic or antagonist effects with IFN- $\gamma$  and TNF- $\alpha$ . *J Immunol* **162**:494–502.
- Allavena P, Giardino G, Bianchi G, and Mantovani A (1997) IL-15 is chemotactic for natural killer cells and stimulates their adhesion to vascular endothelium. *J Leukoc Biol* **61**:729–735.
- Allavena P, Piemonti L, Longoni D, Bernasconi S, Stoppacciaro A, Ruco L, and Mantovani A (1998) IL-10 prevents the differentiation of monocytes to dendritic cells but promotes their maturation to macrophages. *Eur J Immunol* **28**:359–369.
- Apostolopoulos J, Davenport P, and Tipping PG (1996) Interleukin-8 production by macrophages from atheromatous plaques. *Arterioscler Thromb Vasc Biol* **16**:1007–1012.
- Arbustini E, Grasso M, Diegoli M, Pucci A, Bramero M, Ardissino D, Angoli L, de Servi S, Bramucci E, Mussini A, et al. (1991) Coronary atherosclerotic plaques with and without thrombus in ischemic heart syndromes: a morphologic, immunohistochemical and biochemical study. *Am J Cardiol* **68**:36B–50B.
- Arend WP, Joslin FG, and Massoni RJ (1985) Effects of immune complexes on



production by human monocytes of interleukin 1 or an interleukin 1 inhibitor. *J Immunol* **134**:3868–3875.

Arend WP, Malyak M, Guthridge CJ, and Gabay C (1998) Interleukin-1 receptor antagonist: role in biology. *Annu Rev Immunol* **16**:27–55.

Arend WP, Welgus HG, Thompson RC, and Eisenberg SP (1990) Biological properties of recombinant human monocyte-derived interleukin 1 receptor antagonist. *J Clin Invest* **85**:1694–1697.

Asadullah K, Docke WD, Ebeling M, Friedrich M, Belbe G, Audring H, Volk HD, and Sterry W (1999) Interleukin 10 treatment of psoriasis: clinical results of a phase 2 trial. *Arch Dermatol* **135**:187–192.

Auernhammer CJ and Melmed S (1999) Interleukin-11 stimulates proopiomelanocortin gene expression and adrenocorticotropic secretion in corticotroph cells: evidence for a redundant cytokine network in the hypothalamo-pituitary-adrenal axis. *Endocrinology* **140**:1559–1566.

Awane M, Andres PG, Li DJ, and Reinecker HC (1999) NF-kappa B-inducing kinase is a common mediator of IL-17-, TNF-alpha- and IL-1 beta-induced chemokine promoter activation in intestinal epithelial cells. *J Immunol* **162**:5337–5344.

Bagel J, Garland WT, Breneman D, Holick M, Littlejohn TW, Crosby D, Faust H, Fivenson D, and Nichols J (1998) Administration of DAB389IL-2 to patients with recalcitrant psoriasis: a double-blind, phase II multicenter trial. *J Am Acad Dermatol* **38**:938–944.

Barks JL, McQuillan JJ, and Iademarco MF (1997) TNF-alpha and IL-4 synergistically increase vascular cell adhesion molecule-1 expression in cultured vascular smooth muscle cells. *J Immunol* **159**:4532–4538.

Barnes PJ (2001a) Th2 cytokines and asthma: an introduction. *Respir Res* **2**:64–65.

Barnes PJ (2001b) Cytokine-directed therapies for asthma. *J Allergy Clin Immunol* **108**:S72–S76.

Barton BE, Shortall J, and Jackson JV (1996) Interleukins 6 and 11 protect mice from mortality in a staphylococcal enterotoxin-induced toxic shock model. *Infect Immun* **64**:714–718.

Beasley D, McGuiggan ME, and Dinarello CA (1995) Human vascular smooth muscle cells produce an intracellular form of interleukin-1 receptor antagonist. *Am J Physiol* **269**:C961–C968.

Bendtsen K, Hansen MB, Diamant M, Ross C, and Svenson M (1994) Naturally occurring autoantibodies to interleukin-1 alpha, interleukin-6, interleukin-10 and interferon-alpha. *J Interferon Res* **14**:157–158.

Bendtsen K, Hansen MB, Ross C, and Svenson M (1998) High-avidity autoantibodies to cytokines. *Immunol Today* **19**:209–211.

Bendtsen K, Svenson M, Fomsgaard A, and Poulsen LK (1989) Native inhibitors (autoantibodies) of IL-1 alpha and TNF. *Immunol Today* **10**:222.

Bendtsen K, Svenson M, Jonsson V, and Hippe E (1990) Autoantibodies to cytokines—friends or foes? *Immunol Today* **11**:167–169.

Bennett BL, Lacosn RG, Chen CC, Cruz R, Wheeler JS, Kletzian RF, Tomasselli AG, Heinrichson RL, and Manning AM (1996) Identification of signal-induced IkappaB-alpha kinases in human endothelial cells. *J Biol Chem* **271**:19680–19688.

Bennett CF, Condon TP, Grimm S, Chan H, and Chiang MY (1994) Inhibition of endothelial cell adhesion molecule expression with antisense oligonucleotides. *J Immunol* **152**:3530–3540.

Bermudez EA, Rifai N, Buring JE, Manson JE, and Ridker PM (2002) Relation between markers of systemic vascular inflammation and smoking in women. *Am J Cardiol* **89**:1117–1119.

Bertini R, Sironi M, Martin-Padura I, Colotta F, Rambaldi S, Bernasconi S, Ghezzi P, Haskill SJ, Liu D, and Mantovani A (1992) Inhibitory effect of recombinant intracellular interleukin 1 receptor antagonist on endothelial cell activation. *Cytokine* **4**:44–47.

Bevilacqua MP, Pober JS, Wheeler ME, Cotran RS, and Gimbrone MA (1985) Interleukin 1 acts on cultured human vascular endothelium to increase the adhesion of polymorphonuclear leukocytes, monocytes and related leukocyte cell lines. *J Clin Invest* **76**:2003–2011.

Black RA, Rauch CT, Kozlosky CJ, Peschon JJ, Slack JL, Wolfson MF, Castner BJ, Stocking KL, Reddy P, Srinivasan S, et al. (1997) A metalloproteinase disintegrin that releases tumour-necrosis factor-alpha from cells. *Nature (Lond)* **385**:729–733.

Blankenbreg S, Rupprecht HJ, Bickel C, Espinola-Klein C, Rippin G, Hafner G, Ossendorf M, Steinhagen K, and Meyer J (2001) Cytomegalovirus infection with interleukin-6 response predicts cardiac mortality in patients with coronary artery disease. *Circulation* **103**:2915–2921.

Blankenberg S, Tiret L, Bickel C, Peetz D, Cambien F, Meyer J, and Rupprecht HJ (2002) Interleukin-18 is a strong predictor of cardiovascular death in stable and unstable angina. *Circulation* **106**:24–30.

Blum A, Sclarovsky S, and Shohat B (1995) T lymphocyte activation in stable angina pectoris and after percutaneous transluminal coronary angioplasty. *Circulation* **91**:20–22.

Bochner BS, Klunk DA, Sterbinsky SA, Coffman RL, and Schleimer RP (1995) IL-13 selectively induces vascular cell adhesion molecule-1 expression in human endothelial cells. *J Immunol* **154**:799–803.

Bochner BS, Lusinskas FW, Gimbrone MA, Newman W, Sterbinsky SA, Derser-Anthony CP, Klunk D, and Schleimer RP (1991) Adhesion of human basophils, eosinophils and neutrophils to interleukin 1-activated human vascular endothelial cells: contributions of endothelial cell adhesion molecules. *J Exp Med* **173**:1553–1557.

Boelaars-van Haperen MJAM, Baan CC, van Riemsdijk IC, Ijzermans JNM, and Weimar W (2001) Treatment with the chimeric anti-IL-2R alpha basiliximab affects both the IL-2 and IL-15 signalling pathways after clinical kidney transplantation. *Transplant Proc* **33**:1007–1008.

Bogdan C, Thuring H, Dlaska M, Rollinghoff M, and Weiss G (1997) Mechanism of suppression of macrophage nitric oxide release by IL-13: influence of the macrophage population. *J Immunol* **159**:4506–4513.

Bogdan C, Vodovotz Y, and Nathan C (1991) Macrophage deactivation by interleukin 10. *J Exp Med* **174**:1549–1555.

Boisvert WA, Santiago R, Curtiss LK, and Terkeltaub RA (1998) A leukocyte homologue of the IL-8 receptor CXCR-2 mediates the accumulation of macrophages in

atherosclerotic lesions of LDL receptor-deficient mice. *J Clin Invest* **101**:353–363.

Bond M, Chase AJ, Baker AH, and Newby AC (2001) Inhibition of transcription factor NF-kappa B reduces matrix metalloproteinase-1, -3 and -9 production by vascular smooth muscle cells. *Cardiovasc Res* **50**:556–565.

Bondeo J, Browne KA, Brennan FM, Foxwell BM, and Feldmann M (1999) Selective regulation of cytokine induction by adenoviral gene transfer of IkappaBalpha into human macrophages: lipopolysaccharide-induced, but not zymosan-induced, proinflammatory cytokines are inhibited, but IL-10 is nuclear factor-kappaB independent. *J Immunol* **162**:2939–2945.

Bonecchi R, Facchetti F, Dusi S, Luini W, Lissandrini D, Sirmelink M, Locati M, Bernasconi S, Allavena P, Brandt E, et al. (2000) Induction of functional IL-8 receptors by IL-4 and IL-13 in human monocytes. *J Immunol* **164**:3862–3869.

Born TL, Morrison LA, Esteban DJ, VandenBos T, Thebeau LG, Chen N, Spriggs MK, Sims JE, and Buller RM (2000) A poxvirus protein that binds to and inactivates IL-18 and inhibits NK cell response. *J Immunol* **164**:3246–3254.

Bourcier T, Sukhova G, and Libby P (1997) The nuclear factor kappa-B signaling pathway participates in dysregulation of vascular smooth muscle cells in vitro and in human atherosclerosis. *J Biol Chem* **272**:15817–15824.

Bozkurt B, Torre-Amione G, Warren MS, Whitmore J, Soran OZ, Feldman AM, and Mann DL (2001) Results of targeted anti-tumor necrosis factor therapy with etanercept (ENBREL) in patients with advanced heart failure. *Circulation* **103**:1044–1047.

Bozza M, Bliss JL, Dorner AJ, and Trepicchio WL (2001) Interleukin-11 modulates Th1/Th2 cytokine production from activated CD4(+) T cells. *J Interferon Cytokine Res* **21**:21–30.

Bradham WS, Bozkurt B, Gunasinghe H, Mann D, and Spinale FG (2002) Tumor necrosis factor-alpha and myocardial remodeling in progression of heart failure: a current perspective. *Cardiovasc Res* **53**:822–830.

Brand K, Page S, Rogler G, Bartsch A, Brandl R, Knuechel R, Page M, Kaltschmidt C, Baeuerle PA, and Neumeier D (1996) Activated transcription factor nuclear factor-kappa B, is present in the atherosclerotic lesion. *J Clin Invest* **97**:1715–1722.

Braun M, Pietsch P, Felix SB, and Baumann G (1995) Modulation of intercellular adhesion molecule-1 and vascular cell adhesion molecule-1 on human coronary smooth muscle cells by cytokines. *J Mol Cell Cardiol* **27**:2571–2579.

Braunstein JB, Cheng A, Cohn G, Aggarwal M, Nass CM, and Blumenthal RS (2001) Lipid disorders—justification of methods and goals of treatment. *Chest* **120**:979–988.

Bresnihan B, Alvaro-Gracia JM, Cobby M, Doherty M, Domljan Z, Emery P, Nuki G, Pavelka K, Rau R, Rozman B, et al. (1998) Treatment of rheumatoid arthritis with recombinant human interleukin-1 receptor antagonist. *Arthritis Rheum* **41**:2196–2204.

Brizzi MF, Formato L, Dentelli P, Rosso A, Pavan M, Garbarino G, Pegoraro M, Camussi G, and Pegoraro L (2001) Interleukin-3 stimulates migration and proliferation of vascular smooth muscle cells—a potential role in atherogenesis. *Circulation* **103**:549–554.

Brizzi MF, Garbarino G, Rossi PR, Pagliardi GL, Arduino C, Avanzi GC, and Pegoraro L (1993) Interleukin 3 stimulates proliferation and triggers endothelial-leukocyte adhesion molecule 1 gene activation of human endothelial cells. *J Clin Invest* **91**:2887–2892.

Brown AS, Baker-Arkema RG, Yellen L, Henley RW, Guthrie R, Campbell CF, Koren M, Wok, McLain R, and Black DM (1998) Treating patients with documented atherosclerosis to National Cholesterol Education Program-recommended low-density-lipoprotein cholesterol goals with atorvastatin, fluvastatin, lovastatin and simvastatin. *J Am Coll Cardiol* **32**:665–672.

Bump NJ, Hackett M, Huginin M, Seshagiri S, Brady K, Chen P, Ferenz C, Franklin S, Ghayur T, Li P, et al. (1995) Inhibition of ICE family proteases by baculovirus antiapoptotic protein p35. *Science (Wash DC)* **269**:1885–1888.

Burch RM and Mahan LC (1991) Oligonucleotides antisense to the interleukin 1 receptor mRNA block the effects of interleukin 1 in cultured murine and human fibroblasts and in mice. *J Clin Invest* **88**:1190–1196.

Calderara S, Xiang Y, and Moss B (2001) Orthopoxvirus IL-18 binding proteins: Affinities and antagonist activities. *Virology* **279**:22–26.

Cao X, Shores EW, Hu Li J, Anver MR, Kelsall BL, Russell SM, Drago J, Noguchi M, Grinberg A, and Bloom ET (1995) Defective lymphoid development in mice lacking expression of the common cytokine receptor gamma chain. *Immunity* **2**:223–238.

Carson WE, Giri JG, Lindemann MJ, Linett ML, Ahdieh M, Paxton R, Anderson D, Eisenmann J, Grabstein K, and Caligiuri MA (1994) Interleukin (IL) 15 is a novel cytokine that activates human natural killer cells via components of the IL-2 receptor. *J Exp Med* **180**:1395–1403.

Cassatella MA, Gasperini S, Bovolenta C, Calzetti F, Vollebregt M, Scapini P, Marchi M, Suzuki R, Suzuki A, and Yoshimura A (1999) Interleukin-10 (IL-10) selectively enhances C15/SOCS3 mRNA expression in human neutrophils: evidence for an IL-10-induced pathway that is independent of STAT protein activation. *Blood* **94**:2880–2889.

Cassatella MA, Meda L, Bonora S, Ceska M, and Constantin G (1993) Interleukin 10 (IL-10) inhibits the release of proinflammatory cytokines from human polymorphonuclear leukocytes. Evidence for an autocrine role of tumor necrosis factor and IL-1 beta in mediating the production of IL-8 triggered by lipopolysaccharide. *J Exp Med* **178**:2207–2211.

Center DM and Cruikshank W (1982) Modulation of lymphocyte migration by human lymphokines. I. Identification and characterization of chemoattractant activity for lymphocytes from mitogen-stimulated mononuclear cells. *J Immunol* **128**:2563–2568.

Chomarar P, Banchereau J, Davoust J, and Palucka AK (2000) IL-6 switches the differentiation of monocytes from dendritic cells to macrophages. *Nat Immunol* **1**:510–514.

Choudhury RP, Fuster V, Badimon JJ, Fisher EA, and Fayad ZA (2002) MRI and characterization of atherosclerotic plaque: emerging applications and molecular imaging. *Arterioscler Thromb Vasc Biol* **22**:1065–1074.

- Chu RS, Targoni OS, Krieg AM, Lehmann PV, and Harding CV (1997) CpG oligodeoxynucleotides act as adjuvants that switch on T helper 1 (Th1) immunity. *J Exp Med* **186**:1623–1631.
- Chua AO, Chizzonite R, Desai BB, Truitt TP, Nunes P, Minetti LJ, Warriar RR, Presky DH, Levine JF, Gately MK, and Gubler U (1994) Expression cloning of a human IL-12 receptor component. A new member of the cytokine receptor superfamily with strong homology to gp130. *J Immunol* **153**:128–136.
- Chung CD, Liao J, Liu B, Rao X, Jay P, Berta P, and Shuai K (1997) Specific inhibition of Stat3 signal transduction by PIAS3. *Science (Wash DC)* **278**:1803–1805.
- Ciapponi L, Maione D, Scoumanne A, Costa P, Hansen MB, Svenson M, Bendtzen K, Alonzi T, Paonessa G, Cortese R, et al. (1997) Induction of interleukin-6 (IL-6) autoantibodies through vaccination with an engineered IL-6 receptor antagonist. *Nat Biotechnol* **15**:997–1001.
- Clerk A and Sugden PH (1998) The p38-MAPK inhibitor, SB203580, inhibits cardiac stress-activated protein kinases/c-Jun N-terminal kinases (SAPKs/JNKs). *FEBS Lett* **426**:93–96.
- Clinton SK, Underwood R, Hayes L, Sherman ML, Kufe DW, and Libby P (1992) Macrophage colony-stimulating factor gene expression in vascular cells and in experimental and human atherosclerosis. *Am J Pathol* **140**:301–316.
- Cobb RR, Felts KA, Parry GC, and Mackman N (1996) Proteasome inhibitors block VCAM-1 and ICAM-1 gene expression in endothelial cells without affecting nuclear translocation of nuclear factor-kappa B. *Eur J Immunol* **26**:839–845.
- Cochran FR and Finch-Arietta MB (1992) Interleukin-6 can prime THP-1 macrophages for enhanced production of tumor necrosis factor-alpha in response to LPS. *Immunopharmacology* **23**:97–103.
- Collins T (1993) Endothelial nuclear factor-kappa B and the initiation of the atherosclerotic lesion. *Lab Invest* **68**:499–508.
- Collins T and Cybulsky MI (2001) NF-kappa B: pivotal mediator or innocent bystander in atherogenesis? *J Clin Invest* **107**:255–264.
- Collins T, Read MA, Neish AS, Whitley MZ, Thanos D, and Maniatis T (1995) Transcriptional regulation of endothelial cell adhesion molecules: NF-kappa B and cytokine-inducible enhancers. *FASEB J* **9**:899–909.
- Colombel JF, Rutgeerts P, Malchow H, Jacyna M, Nielsen OH, Rask-Madsen J, Van Deventer S, Ferguson A, Desreumaux P, Forbes A, et al. (2001) Interleukin 10 (Tenovil) in the prevention of postoperative recurrence of Crohn's disease. *Gut* **49**:42–46.
- Colotta F, Bussolino F, Polentarutti N, Guglielmetti A, Sironi M, Bocchietto E, De Rossi M, and Mantovani A (1993a) Differential expression of the common beta and specific alpha chains of the receptors for GM-CSF, IL-3 and IL-5 in endothelial cells. *Exp Cell Res* **206**:311–317.
- Colotta F, Re F, Muzio M, Bertini R, Polentarutti N, Sironi M, Giri JG, Dower SK, Sims JE, and Mantovani A (1993b) Interleukin-1 type II receptor: a decoy target for IL-1 that is regulated by IL-4. *Science (Wash DC)* **261**:472–475.
- Colotta F, Re F, Muzio M, Polentarutti N, Minty A, Caput D, Ferrara P, and Mantovani A (1994) Interleukin-13 induces expression and release of interleukin-1 decoy receptor in human polymorphonuclear cells. *J Biol Chem* **269**:12403–12406.
- Cornicelli JA, Butteiger D, Rateri DL, Welch K, and Daugherty A (2000) Interleukin-4 augments acetylated LDL-induced cholesterol esterification in macrophages. *J Lipid Res* **41**:376–383.
- Crooke ST (2000) Oligonucleotide-based drugs in the control of cytokine synthesis, in *Novel Cytokine Inhibitors* (Higgs GA and Henderson B eds) pp 83–101, Birkhäuser Verlag AG, Basel.
- Croucher PI, Hoken I, and Hargreaves PG (2000) Inhibiting cytokine-processing enzymes, in *Novel Cytokine Inhibitors* (Higgs GA and Henderson B eds) pp 103–122, Birkhäuser Verlag AG, Basel.
- Croxford JL, Triantaphyllopoulos K, Podhajec OL, Feldmann M, Baker D, and Chernajovsky Y (1998) Cytokine gene therapy in experimental allergic encephalomyelitis by injection of plasmid DNA-cationic liposome complex into the central nervous system. *J Immunol* **160**:5181–5187.
- Cruikshank WW, Kornfeld H, and Center DM (2000) Interleukin-16. *J Leukoc Biol* **67**:757–766.
- Cuenda A, Rouse J, Doza YN, Meier R, Cohen P, Gallagher TF, Young PR, and Lee JC (1995) SB 203580 is a specific inhibitor of a MAP kinase homologue which is stimulated by cellular stresses and interleukin-1. *FEBS Lett* **364**:229–233.
- Cunnane G, Madigan A, Murphy E, FitzGerald O, and Bresnihan B (2001) The effects of treatment with interleukin-1 receptor antagonist on the inflamed synovial membrane in rheumatoid arthritis. *Rheumatology* **40**:62–69.
- Curtis DJ, Hilton DJ, Roberts M, Murray L, Nicola N, and Begley CG (1997) Recombinant soluble interleukin-11 (IL-11) receptor alpha-chain can act as an IL-11 antagonist. *Blood* **90**:4403–4412.
- D'Andrea R, Rengaraju M, Valiante NM, Chehimi J, Kubin M, Aste M, Chan SH, Kobayashi M, Young D, Nickbarg E, et al. (1992) Production of natural killer cell stimulatory factor (interleukin 12) by peripheral blood mononuclear cells. *J Exp Med* **176**:1387–1398.
- D'Andrea RJ, Harrison-Findik D, Butcher CM, Fennie J, Blumbergs P, Bartley P, McCormack M, Jones K, Rowland R, Gonda TJ, and Vadas MA (1998) Dysregulated hematopoiesis and a progressive neurological disorder induced by expression of an activated form of the human common beta chain in transgenic mice. *J Clin Invest* **102**:1951–1960.
- de Boer OJ, van der Wal AC, Verhagen CE, and Becker AE (1999) Cytokine secretion profiles of cloned T cells from human aortic atherosclerotic plaques. *J Pathol* **188**:174–179.
- De Caterina R, Bourcier T, Laufs U, La Fata V, Lazzerini G, Neish AS, Libby P, and Liao JK (2001) Induction of endothelial-leukocyte interaction by interferon-gamma requires coactivation of nuclear factor-kappa B. *Arterioscler Thromb Vasc Biol* **21**:227–232.
- de' Clari F, Salani I, Safwan E, and Giannacco A (2002) Sudden death in a patient without heart failure after a single infusion of 200 mg infliximab: does TNF-alpha have protective effects on the failing heart, or does infliximab have direct harmful cardiovascular effects? *Circulation* **105**:E183.
- de Waal Malefyt R, Abrams J, Bennett B, Figdor CG, and de Vries JE (1991a) Interleukin 10 (IL-10) inhibits cytokine synthesis by human monocytes: an autoregulatory role of IL-10 produced by monocytes. *J Exp Med* **174**:1209–1220.
- de Waal Malefyt R, Figdor CG, Huijbens R, Mohan-Peterson S, Bennett B, Cuijpepper J, Dang W, Zurawski G, and de Vries JE (1993) Effects of IL-13 on phenotype, cytokine production and cytotoxic function of human monocytes. Comparison with IL-4 and modulation by IFN-gamma or IL-10. *J Immunol* **151**:6370–6381.
- de Waal Malefyt R, Haanen J, Spits H, Roncarolo MG, te Velde A, Figdor C, Johnson K, Kastelein R, Yssel H, and de Vries JE (1991b) Interleukin 10 (IL-10) and viral IL-10 strongly reduce antigen-specific human T cell proliferation by diminishing the antigen-presenting capacity of monocytes via down-regulation of class II major histocompatibility complex expression. *J Exp Med* **174**:915–924.
- Del Vecchio MA, Georgescu HI, McCormack JE, Robbins PD, and Evans CH (2001) Approaches to enhancing the retroviral transduction of human synovocytes. *Arthritis Res* **3**:259–263.
- Dentelli P, Del-Sorbo L, Rosso A, Molinar A, Garbarino G, Camussi G, Pegoraro L, and Brizzi MF (1999) Human IL-3 stimulates endothelial cell motility and promotes in vivo new vessel formation. *J Immunol* **163**:2151–2159.
- Devlin CM, Kuriakose G, Hirsch E, and Tabas I (2002) Genetic alterations of IL-1 receptor antagonist in mice affect plasma cholesterol level and foam cell lesion size. *Proc Natl Acad Sci USA* **99**:6280–6285.
- Devlin RD, Reddy SV, Savino R, Ciliberto G, and Roodman GD (1998) IL-6 mediates the effects of IL-1 or TNF, but not PTHrP or 1, 25(OH)2D3, on osteoclast-like cell formation in normal human bone marrow cultures. *J Bone Miner Res* **13**:393–399.
- Dickensheets HL, Venkataraman C, Schindler U, and Donnelly RP (1999) Interferons inhibit activation of STAT6 by interleukin 4 in human monocytes by inducing SOCS-1 gene expression. *Proc Natl Acad Sci USA* **96**:10800–10805.
- Dinarello CA (1997) Interleukin-1. *Cytokine Growth Factor Rev* **8**:253–265.
- Dinarello CA (1999) IL-18: A T-H1-inducing, proinflammatory cytokine and new member of the IL-1 family. *J Allergy Clin Immunol* **103**:11–24.
- Ding Y, Qin L, Kotenko SV, Pestka S, and Bromberg JS (2000) A single amino acid determines the immunostimulatory activity of interleukin 10. *J Exp Med* **191**:213–224.
- Doherty TM, Kastelein R, Menon S, Andrade S, and Coffman RL (1993) Modulation of murine macrophage function by IL-13. *J Immunol* **151**:7151–7160.
- Dong ZM, Brown AA, and Wagner DD (2000) Prominent role of P-selectin in the development of advanced atherosclerosis in apoE-deficient mice. *Circulation* **101**:2290–2295.
- Donnelly RP, Crofford LJ, Freeman SL, Buras J, Remmers E, Wilder RL, and Fenton MJ (1993) Tissue-specific regulation of IL-6 production by IL-4. Differential effects of IL-4 on nuclear factor-kappa B activity in monocytes and fibroblasts. *J Immunol* **151**:5603–5612.
- Donnelly RP, Dickensheets H, and Finbloom DS (1999) The interleukin-10 signal transduction pathway and regulation of gene expression in mononuclear phagocytes. *J Interferon Cytokine Res* **19**:563–573.
- Dumoutier L, Louahed J, and Renauld JC (2000) Cloning and characterization of IL-10-related T cell-derived inducible factor (IL-TIF), a novel cytokine structurally related to IL-10 and inducible by IL-9. *J Immunology* **164**:1814–1819.
- Edwards PA and Ericsson J (1999) Sterols and isoprenoids: Signaling molecules derived from the cholesterol biosynthetic pathway. *Annu Rev Biochem* **68**:157–185.
- Elhage R, Clamens S, Besnard S, Mallat Z, Tedgui A, Arnal JF, Maret A, and Bayard F (2001) Involvement of interleukin-6 in atherosclerosis but not in the prevention of fatty streak formation by 17 beta-estradiol in apolipoprotein E-deficient mice. *Atherosclerosis* **156**:315–320.
- Elhage R, Maret A, Pieraggi MT, Thiers JC, Arnal JF, and Bayard F (1998) Differential effects of interleukin-1 receptor antagonist and tumor necrosis factor binding protein on fatty-streak formation in apolipoprotein E-deficient mice. *Circulation* **97**:242–244.
- Elliott MJ, Gamble JR, Park LS, Vadas MA, and Lopez AF (1991) Inhibition of human monocyte adhesion by interleukin-4. *Blood* **77**:2739–2745.
- Elliott MJ, Vadas MA, Cleland LG, Gamble JR, and Lopez AF (1990) IL-3 and granulocyte-macrophage colony-stimulating factor stimulate two distinct phases of adhesion in human monocytes. *J Immunol* **145**:167–176.
- Erren M, Reinecke H, Junker R, Fobker M, Schulte H, Schurek JO, Kropf J, Kerber S, Breithardt G, Assmann G, and Cullen P (1999) Systemic inflammatory parameters in patients with atherosclerosis of the coronary and peripheral arteries. *Arterioscler Thromb Vasc Biol* **19**:2355–2363.
- Estes P, Nandi A, Mohamadzadeh M, and Siegelman MH (1999) Interleukin 15 induces endothelial hyaluronan expression in vitro and promotes activated T cell extravasation through a CD44-dependent pathway in vivo. *J Exp Med* **190**:9–19.
- Faber BCG, Cleutjens KBJM, Niessen RLJ, Aarts PLJV, Boon W, Greenberg AS, Kitslaar PJEHM, Tordoir JHM, and Daemen MJAP (2001) Identification of genes potentially involved in rupture of human atherosclerotic plaques. *Circ Res* **89**:547–554.
- Fanslow WC, Spriggs MK, Rauch CT, Clifford KN, Macduff BM, Ziegler SF, Schooley KA, Mohler KM, March CJ, and Armitage RJ (1993) Identification of a distinct low-affinity receptor for human interleukin-4 on pre-B cells. *Blood* **81**:2998–3005.
- Fayad ZA and Fuster V (2001) Clinical imaging of the high-risk or vulnerable atherosclerotic plaque. *Circ Res* **89**:305–316.
- Feeley BT, Miniati DN, Park AK, Hoyt EG, and Robbins RC (2000) Nuclear factor-kappa B transcription factor decoy treatment inhibits graft coronary artery disease after cardiac transplantation in rodents. *Transplantation* **70**:1560–1568.
- Feng J, Han J, Pearce SF, Silverstein RL, Gotto AM, Hajjar DP, and Nicholson AC (2000) Induction of CD36 expression by oxidized LDL and IL-4 by a common signaling pathway dependent on protein kinase C and PPAR-gamma. *J Lipid Res* **41**:688–696.
- Fernandes J, Tardif G, Martel-Pelletier J, Lascau-Coman V, Dupuis M, Moldovan F, Sheppard M, Krishnan BR, and Pelletier JP (1999) In vivo transfer of interleukin-1 receptor antagonist gene in osteoarthritic rabbit knee joints: prevention of osteoarthritis progression. *Am J Pathol* **154**:1159–1169.
- Fernandez-Botran R and Vitetta ES (1990) A soluble, high-affinity, interleukin-4

- binding protein is present in the biological fluids of mice. *Proc Natl Acad Sci USA* **87**:4202–4206.
- Fickenscher H, Hor S, Kupers H, Knappe A, Wittmann S, and Sticht H (2002) The interleukin-10 family of cytokines. *Trends Immunol* **23**:89–96.
- Finco TS and Baldwin AS (1995) Mechanistic aspects of NF-kappa B regulation: the emerging role of phosphorylation and proteolysis. *Immunology* **3**:263–272.
- Finkelman FD, Madden KB, Morris SC, Holmes JM, Boiani N, Katona IM, and Maliszewski CR (1993) Anti-cytokine antibodies as carrier proteins. Prolongation of in vivo effects of exogenous cytokines by injection of cytokine-anti-cytokine antibody complexes. *J Immunol* **151**:1235–1244.
- Fiorentino DF, Bond MW, and Mosmann TR (1989) Two types of mouse T helper cell. IV. Th2 clones secrete a factor that inhibits cytokine production by Th1 clones. *J Exp Med* **170**:2081–2095.
- Flex A, Gaetani E, Pola R, Santoliquido A, Aloi F, Papaleo P, Dal Lago A, Pola E, Serricchio M, Tondi P, and Pola P (2002) The -174 G/C polymorphism of the interleukin-6 gene promoter is associated with peripheral artery occlusive disease. *Eur J Vasc Endovasc Surg* **24**:264–268.
- Folcik VA, Aamir R, and Cathcart MK (1997) Cytokine modulation of LDL oxidation by activated human monocytes. *Arterioscler Thromb Vasc Biol* **17**:1954–1961.
- Fossiez F, Banchereau J, Murray R, Van Kooten C, Garrone P, and Lebecque S (1998) Interleukin-17. *Int Rev Immunol* **16**:541–551.
- Francis SE, Camp NJ, Burton AJ, Dewberry RM, Gunn J, Stephens-Lloyd A, Cumberland DC, Gershlick A, and Crossman DC (2001) Interleukin 1 receptor antagonist gene polymorphism and restenosis after coronary angioplasty. *Heart* **86**:336–340.
- Francis SE, Camp NJ, Dewberry RM, Gunn J, Syrris P, Carter ND, Jeffery S, Kaski JC, Cumberland DC, Duff GW, and Crossman DC (1999) Interleukin-1 receptor antagonist gene polymorphism and coronary artery disease. *Circulation* **99**:861–866.
- Frostgard J, Ulfgren AK, Nyberg P, Hedin U, Swedenborg J, Andersson U, and Hansson GK (1999) Cytokine expression in advanced human atherosclerotic plaques: dominance of pro-inflammatory (Th1) and macrophage-stimulating cytokines. *Atherosclerosis* **145**:33–43.
- Furie MB and McHugh DD (1989) Migration of neutrophils across endothelial monolayers is stimulated by treatment of the monolayers with interleukin-1 or tumor necrosis factor-alpha. *J Immunol* **143**:3309–3317.
- Furukawa Y, Becker G, Stinn JL, Shimizu K, Libby P, and Mitchell RN (1999) Interleukin-10 (IL-10) augments allograft arterial disease: paradoxical effects of IL-10 in. *Am J Pathol* **155**:1929–1939.
- Galea J, Armstrong J, Gadsdon P, Holden H, Francis SE, and Holt CM (1996) Interleukin-1 beta in coronary arteries of patients with ischemic heart disease. *Arterioscler Thromb Vasc Biol* **16**:1000–1006.
- Galis ZS, Muszynski M, Sukhova GK, Simon-Morrissey E, and Libby P (1995) Enhanced expression of vascular matrix metalloproteinases induced in vitro by cytokines and in regions of human atherosclerotic lesions. *Ann NY Acad Sci* **748**:501–748507.
- Garcia GE, Xia YY, Chen SZ, Wang YB, Ye RD, Harrison JK, Bacon KB, Zerwes HG, and Feng LL (2000) NF-kappa B-dependent fractalkine induction in rat aortic endothelial cells stimulated by IL-1 beta, TNF-alpha and LPS. *J Leukoc Biol* **67**:577–584.
- Gately MK, Carvajal DM, Connaughton SE, Gillessen S, Warrier RR, Kolinsky KD, Wilkinson VL, Dwyer CM, Higgins GF, Podlaski FJ, et al. (1996) Interleukin-12 antagonist activity of mouse interleukin-12 p40 homodimer in vitro and in vivo. *Ann N Y Acad Sci* **795**:1–12.
- Gately MK, Desai BB, Wolitzky AG, Quinn PM, Dwyer CM, Podlaski FJ, Familletti PC, Sinigaglia F, Chizzonite R, Gubler U, and Stern AS (1991) Regulation of human lymphocyte proliferation by a heterodimeric cytokine, IL-12 (cytotoxic lymphocyte maturation factor). *J Immunol* **147**:874–882.
- Gavett SH, O'Hearn DJ, Karp CL, Patel EA, Schofield BH, Finkelman FD, and Wills-Karp M (1997) Interleukin-4 receptor blockade prevents airway responses induced by antigen challenge in mice. *Am J Physiol* **272**:L253–L261.
- Gazzinelli RT, Oswald IP, James SL, and Sher A (1992) IL-10 inhibits parasite killing and nitrogen oxide production by IFN-gamma-activated macrophages. *J Immunol* **148**:1792–1796.
- Gearing DP, Comeau MR, Friend DJ, Gimpel SD, Thut CJ, McGourty J, Brasher KK, King JA, Gillis S, Mosley B, Ziegler SF, and Cosman D (1992) The IL-6 signal transducer, gp130: an oncostatin M receptor and affinity converter for the LIF receptor. *Science (Wash DC)* **255**:1434–1437.
- Gearing DP, Thut CJ, VandeBos T, Gimpel SD, Delaney PB, King J, Price V, Cosman D, and Beckmann MP (1991) Leukemia inhibitory factor receptor is structurally related to the IL-6 signal transducer, gp130. *EMBO J* **10**:2839–2848.
- Gene Therapy Clinical Trials (2002). *J Gene Med On-line Edition*, 2002 update (<http://www.wiley.co.uk/genetherapy/clinical>).
- George J, Mulkins M, Shaish A, Casey S, Schatzman R, Sigal E, and Harats D (2000a) Interleukin (IL)-4 deficiency does not influence fatty streak formation in C57BL/6 mice. *Atherosclerosis* **153**:403–411.
- George J, Shoenfeld Y, Gilburd B, Afek A, Shaish A, and Harats D (2000b) Requisite role for interleukin-4 in the acceleration of fatty streaks induced by heat shock protein 65 or *Mycobacterium tuberculosis*. *Circ Res* **86**:1203–1210.
- Georges JL, Loukaci V, Poirier O, Evans A, Luc G, Arveiler D, Ruidavets JB, Cambien F, and Tiret L (2001) Interleukin-6 gene polymorphisms and susceptibility to myocardial infarction: the ECTIM study. *J Mol Med* **79**:300–305.
- Gerard C, Bruyans C, Marchant A, Abramowicz D, Vandenabeele P, Delvaux A, Fiers W, Goldman M, and Velu T (1993) Interleukin 10 reduces the release of tumor necrosis factor and prevents lethality in experimental endotoxemia. *J Exp Med* **177**:547–550.
- Gerdes N, Sukhova GK, Libby P, Reynolds RS, Young JL, and Schonbeck U (2002) Expression of interleukin (IL)-18 and functional IL-18 receptor on human vascular endothelial cells, smooth muscle cells and macrophages: implications for atherogenesis. *J Exp Med* **195**:245–257.
- Gershzen RE, Lim YC, Ding HT, Snapp K, Kansas G, Dichek DA, Cabanas C, Sanchez-Madrid F, Gimbrone MA, Rosenzweig A, and Lusinskas FW (1998) Adhesion of monocytes to vascular cell adhesion molecule-1-transduced human endothelial cells—implications for atherogenesis. *Circ Res* **82**:871–878.
- Gerlag DM, Ransone L, Tak PP, Han Z, Palanki M, Barbosa MS, Boyle D, Manning AM, and Firestein GS (2000) The effect of a T cell-specific NF-kappa B inhibitor on in vitro cytokine production and collagen-induced arthritis. *J Immunol* **165**:1652–1658.
- Gerszten RE, Garcia-Zepeda EA, Lim YC, Yoshida M, Ding HA, Gimbrone MA, Luster AD, Lusinskas FW, and Rosenzweig A (1999) MCP-1 and IL-8 trigger firm adhesion of monocytes to vascular endothelium under flow conditions. *Nature (Lond)* **398**:718–723.
- Ghosh S and Baltimore D (1990) Activation in vitro of NF-kappa B by phosphorylation of its inhibitor I kappa B. *Nature (Lond)* **344**:678–682.
- Giampietri A, Grohmann U, Vacca C, Fioretti MC, Puccetti P, and Campanile F (2000) Dual effect of IL-4 on resistance to systemic gram-negative infection and production of TNF-alpha. *Cytokine* **12**:417–421.
- Giannoukakis N, Rudert WA, Ghivizzani SC, Gambotto A, Ricordi C, Trucco M, and Robbins PD (1999) Adenoviral gene transfer of the interleukin-1 receptor antagonist protein to human islets prevents IL-1beta-induced beta-cell impairment and activation of islet cell apoptosis in vitro. *Diabetes* **48**:1730–1736.
- Gillesen S, Carvajal D, Ling P, Podlaski FJ, Stremel DL, Familletti PC, Gubler U, Presky DH, Stern AS, and Gately MK (1995) Mouse interleukin-12 (IL-12) p40 homodimer: a potent IL-12 antagonist. *Eur J Immunol* **25**:200–206.
- Giri JG, Ahdieh M, Eisenman J, Shanebeck K, Grabstein K, Kumaki S, Namen A, Park LS, Cosman D, and Anderson D (1994a) Utilization of the beta and gamma chains of the IL-2 receptor by the novel cytokine IL-15. *EMBO J* **13**:2822–2830.
- Giri JG, Newton RC, and Horuk R (1990) Identification of soluble interleukin-1 binding protein in cell-free supernatants. Evidence for soluble interleukin-1 receptor. *J Biol Chem* **265**:17416–17419.
- Giri JG, Wells J, Dower SK, McCall CE, Guzman RN, Slack J, Bird TA, Shanebeck K, Grabstein KH, Sims JE, and Alderson MR (1994b) Elevated levels of shed type II IL-1 receptor in sepsis. Potential role for type II receptor in regulation of IL-1 responses. *J Immunol* **153**:5802–5809.
- Giri JG, Anderson DM, Kumaki S, Park LS, Grabstein KH, and Cosman D (1995) IL-15, a novel T cell growth factor that shares activities and receptor components with IL-2. *J Leukoc Biol* **57**:763–766.
- Girndt M, Kaul H, Sester U, Ulrich C, Sester M, Georg T, and Kohler H (2002) Anti-inflammatory interleukin-10 genotype protects dialysis patients from cardiovascular events. *Kidney Int* **62**:949–955.
- Godfraind C, Louahed J, Faulkner H, Vink A, Warnier G, Grecnis R, and Renaud JC (1998) Intraepithelial infiltration by mast cells with both connective tissue-type and mucosal-type characteristics in gut, trachea and kidneys of IL-9 transgenic mice. *J Immunol* **160**:3989–3996.
- Goebeler M, Schnarr B, Toksoy A, Kunz M, Brocker EB, Duschl A, and Gollitzer R (1997) Interleukin-13 selectively induces monocyte chemoattractant protein-1 synthesis and secretion by human endothelial cells. Involvement of IL-4R alpha and Stat6 phosphorylation. *Immunology* **91**:450–457.
- Golomb G, Fishbein I, Banai S, Mishaly D, Moscovitz D, Gertz SD, Gazit A, Poradous E, and Levitzki A (1996) Controlled delivery of a tyrosinase inhibits intimal hyperplasia in a rat carotid artery injury model. *Atherosclerosis* **125**:171–182.
- Gottsater A, Forsblad J, Matsch T, Persson K, Ljungcrantz I, Ohlsson K, Lindgarde F (2002) Interleukin-1 receptor antagonist is detectable in human carotid artery plaques and is related to triglyceride levels and Chlamydia pneumoniae IgA antibodies. *J Int Med* **251**:61–68.
- Graham KA, Lalani AS, Macen JL, Ness TL, Barry M, Liu LY, Lucas A, Clark-Lewis I, Moyer RW, and McFadden G (1997) The T1/35kDa family of poxvirus-secreted proteins bind chemokines and modulate leukocyte influx into virus-infected tissues. *Virology* **229**:12–24.
- Grisclavage JM, Wilk S, and Ignarro LJ (1996) Inhibitors of the proteasome pathway interfere with induction of nitric oxide synthase in macrophages by blocking activation of transcription factor NF-kappa B. *Proc Natl Acad Sci USA* **93**:3308–3312.
- Grohmann U, Van Snick J, Campanile F, Silla S, Giampietri A, Vacca C, Renaud JC, Fioretti MC, and Puccetti P (2000) IL-9 protects mice from Gram-negative bacterial shock: suppression of TNF-alpha, IL-12, and IFN-gamma and induction of IL-10. *J Immunol* **164**:4197–4203.
- Guex-Crosier Y, Raber J, Chan CC, Kriete MS, Benichou J, Pilson RS, Kerwin JA, Waldmann TA, Hakimi J, and Roberge FG (1997) Humanized antibodies against the alpha-chain of the IL-2 receptor and against the beta-chain shared by the IL-2 and IL-15 receptors in a monkey uveitis model of autoimmune diseases. *J Immunol* **158**:452–458.
- Guthridge MA, Stomski FC, Thomas D, Woodcock JM, Bagley CJ, Berndt MC, and Lopez AF (1998) Mechanism of activation of the GM-CSF, IL-3 and IL-5 family of receptors. *Stem Cells* **16**:301–313.
- Haddad JJ and Fahlman CS (2002) Redox- and oxidant-mediated regulation of interleukin-10: an anti-inflammatory, antioxidant cytokine? *Biochem Biophys Res Commun* **297**:163–176.
- Hajjar DP and Pomerantz KB (1992) Signal transduction in atherosclerosis: integration of cytokines and the eicosanoid network. *FASEB J* **6**:2933–2941.
- Hajra L, Evans AI, Chen M, Hyduk SJ, Collins T, and Cybulsky MI (2000) The NF-kappa B signal transduction pathway in aortic endothelial cells is primed for activation in regions predisposed to atherosclerotic lesion formation. *Proc Natl Acad Sci USA* **97**:9052–9057.
- Hansen MB, Svenson M, Diamant M, Abell K, and Bendtzen K (1995) Interleukin-6 autoantibodies: possible biological and clinical significance. *Leukemia* **9**:1113–1115.
- Hansen MB, Svenson M, Diamant M, and Bendtzen K (1991) Anti-interleukin-6 antibodies in normal human serum. *Scand J Immunol* **33**:777–781.
- Hansen MB, Svenson M, Diamant M, and Bendtzen K (1993) High-affinity IgG autoantibodies to IL-6 in sera of normal individuals are competitive inhibitors of IL-6 in vitro. *Cytokine* **5**:72–80.

- Hansson GK, Jonasson L, Lofsted B, Stemme S, Kocher O, and Gabbiani G (1988) Localization of T lymphocytes and macrophages in fibrous and complicated human atherosclerotic plaques. *Atherosclerosis* **72**:135–141.
- Harel-Bellan A, Durum S, Muegge K, Abbas AK, and Farrar WL (1988) Specific inhibition of lymphokine biosynthesis and autocrine growth using antisense oligonucleotides in Th1 and Th2 helper T cell clones. *J Exp Med* **168**:2309–2318.
- Hart PH, Bonder CS, Balogh J, Dickensheets HL, Donnelly RP, and Finlay-Jones JJ (1999) Differential responses of human monocytes and macrophages to IL-4 and IL-13. *J Leukoc Biol* **66**:575–578.
- Haudek SB, Bryant DD, and Giroir BP (2001) Differential regulation of myocardial NF kappa B following acute or chronic TNF-alpha exposure. *J Mol Cell Cardiol* **33**:1263–1271.
- Hayashida K, Kitamura T, Gorman DM, Arai K, Yokota T, and Miyajima A (1990) Molecular cloning of a second subunit of the receptor for human granulocyte-macrophage colony-stimulating factor (GM-CSF): reconstitution of a high-affinity GM-CSF receptor. *Proc Natl Acad Sci USA* **87**:9655–9659.
- Heaney ML and Golde DW (1998) Soluble receptors in human disease. *J Leukoc Biol* **64**:135–146.
- Heinrich PC, Behrmann I, Muller-Newen G, Schaper F, and Graeve L (1998) Interleukin-6-type cytokine signalling through the gp130/Jak/STAT pathway. *Biochem J* **334**:297–314.
- Henderson B and Higgs GA (2000) Targets for modulating cytokine responses in inflammatory and infectious diseases, in *Novel Cytokine Inhibitors* (Higgs GA and Henderson B eds) pp 1–8, Birkhäuser Verlag AG, Basel.
- Henderson WR, Chi EY, and Maliszewski CR (2000) Soluble IL-4 receptor inhibits airway inflammation following allergen challenge in a mouse model of asthma. *J Immunol* **164**:1086–1095.
- Henke PK, DeBrnyne LA, Strieter RM, Bromberg S, Prince M, Kadell AM, Sarkar M, Londy F, and Wakefield TW (2000) Viral IL-10 gene transfer decreases inflammation and cell adhesion molecule expression in a rat model of venous thrombosis. *J Immunol* **164**:2131–2141.
- Hibi M, Murakami M, Saito M, Hirano T, Taga T, and Kishimoto T (1990) Molecular cloning and expression of an IL-6 signal transducer, gp130. *Cell* **63**:1149–1157.
- Hipp MS, Urbich C, Mayer P, Wischhusen J, Weller M, Kracht M, and Spyridopoulos I (2002) Proteasome inhibition leads to NF-kappaB-independent IL-8 transactivation in human endothelial cells through induction of AP-1. *Eur J Immunol* **32**:2208–2217.
- Hirano T, Akira S, Taga T, and Kishimoto T (1990) Biological and clinical aspects of interleukin-6. *Immunol Today* **11**:443–449.
- Hiraoka E, Kawashima S, Takahashi T, Rikitake Y, Kitamura T, Ogawa W, and Yokoyama M (2001) TNF-alpha induces protein synthesis through PI3-kinase-Akt/PKB pathway in cardiac myocytes. *Am J Physiol Heart Circ Physiol* **280**:H1861–H1868.
- Hogaboam CM, Vallance BA, Kumar A, Addison CL, Graham FL, Gauldie J, and Collins SM (1997) Therapeutic effects of interleukin-4 gene transfer in experimental inflammatory bowel disease. *J Clin Invest* **100**:2766–2776.
- Honemann D, Chatterjee M, Savino R, Bommert K, Burger R, Gramatzki M, Dorken B, and Bargou RC (2001) The IL-6 receptor antagonist SANT-7 overcomes bone marrow stromal cell-mediated drug resistance of multiple myeloma cells. *Int J Cancer* **93**:674–680.
- Houssiau FA, Renaud JC, Fibbe WE, and Van Snick J (1992) IL-2 dependence of IL-9 expression in human T lymphocytes. *J Immunol* **148**:3147–3151.
- Houston P, Goodman J, Lewis A, Campbell CJ, and Braddock M (2001) Homing markers for atherosclerosis: applications for drug delivery, gene delivery and vascular imaging. *FEBS Lett* **492**:73–77.
- Houtkamp MA, van der Wal AC, de Boer OJ, van der Loos CM, de Boer PAJ, Moorman AFM, and Becker AE (2001) Interleukin-15 expression in atherosclerotic plaques—an alternative pathway for T-cell activation in atherosclerosis? *Arterioscler Thromb Vasc Biol* **21**:1208–1213.
- Hsieh CS, Macatonia SE, Tripp CS, Wolf SF, O'Garra A, and Murphy KM (1993) Development of TH1 CD4+ T cells through IL-12 produced by Listeria-induced macrophages. *Science (Wash DC)* **260**:547–549.
- Hsu HY, Nicholson AC, and Hajjar DP (1996) Inhibition of macrophage scavenger receptor activity by tumor necrosis factor-alpha is transcriptionally and post-transcriptionally regulated. *J Biol Chem* **271**:7767–7773.
- Hsu JY, Hsu MY, Sorger T, Herlyn M, and Levine EM (1999) Heparin/endothelial cell growth supplement regulates matrix gene expression and prolongs life span of vascular smooth muscle cells through modulation of interleukin-1. *In Vitro Cell Dev Biol Anim* **35**:647–654.
- Huber SA, Sakkinen P, Conze D, Hardin N, and Tracy R (1999) Interleukin-6 exacerbates early atherosclerosis in mice. *Arterioscler Thromb Vasc Biol* **19**:2364–2367.
- Huber SA, Sakkinen P, David C, Newell MK, and Tracy RP (2001) T helper-cell phenotype regulates atherosclerosis in mice under conditions of mild hypercholesterolemia. *Circulation* **103**:2610–2616.
- Hughes MD, Hussain M, Nawaz Q, Sayyed P, and Akhtar S (2001) The cellular delivery of antisense oligonucleotides and ribozymes. *Drug Discov Today* **6**:303–315.
- Humphries SE, Luong LA, Ogg MS, Hawe E, and Miller GJ (2001) The interleukin-6-174 G/C promoter polymorphism is associated with risk of coronary heart disease and systolic blood pressure in healthy men. *Eur Heart J* **22**:2243–2252.
- Hurst SD, Muchamuel T, Gorman DM, Gilbert JM, Clifford T, Kwan S, Menon S, Seymour B, Jackson C, Kung TT, et al. (2002) New IL-17 Family Members Promote Th1 or Th2 Responses in the Lung: In Vivo Function of the Novel Cytokine IL-25. *J Immunol* **169**:443–453.
- Huynh TT, Davies MG, Barber L, Svendsen E, and Hagen PO (1998) Local inhibition of tyrosine kinase activity markedly attenuates the development of intimal hyperplasia in experimental vein grafts. *J Surg Res* **77**:104–111.
- Hymowitz SG, Filvaroff EH, Yin J, Lee J, Cai L, Risser P, Maruoka M, Mao W, Foster J, Kelley RF, Pan G, Gurney AL, de Vos AM, and Starovskanik MA (2001) IL-17s adopt a cystine knot fold: structure and activity of a novel cytokine, IL-17F and implications for receptor binding. *EMBO J* **20**:5332–5341.
- Ignarro LJ, Cirino G, Casini A, and Napoli C (1999) Nitric oxide as a signaling molecule in the vascular system: An overview. *J Cardiovasc Pharmacol* **34**:879–886.
- Ikeda U, Ikeda M, Oohara T, Oguchi A, Kamitani T, Tsuruya Y, and Kano S (1991) Interleukin 6 stimulates growth of vascular smooth muscle cells in a PDGF-dependent manner. *Am J Physiol* **260**:H1713–H1717.
- Ikeda U, Ikeda M, Seino Y, Takahashi M, Kano S, and Shimada K (1992) Interleukin 6 gene transcripts are expressed in atherosclerotic lesions of genetically hyperlipidemic rabbits. *Atherosclerosis* **92**:213–218.
- Ikeda U, Ikeda M, Seino Y, Takahashi M, Kasahara T, Kano S, and Shimada K (1993) Expression of intercellular adhesion molecule-1 on rat vascular smooth muscle cells by pro-inflammatory cytokines. *Atherosclerosis* **104**:61–68.
- Ikeda U, Ito T, and Shimada K (2001) Interleukin-6 and acute coronary syndrome. *Clin Cardiol* **24**:701–704.
- Ip NY, Nye SH, Boulton TG, Davis S, Taga T, Li Y, Birren SJ, Yasukawa K, Kishimoto T, Anderson DJ, et al. (1992) CNTF and LIF act on neuronal cells via shared signaling pathways that involve the IL-6 signal transducing receptor component gp130. *Cell* **69**:1121–1132.
- James HA and Gibson I (1998) The therapeutic potential of ribozymes. *Blood* **91**:371–382.
- Janaswami PM, Kalvakolanu DV, Zhang Y, and Sen GC (1992) Transcriptional repression of interleukin-6 gene by adenoviral E1A proteins. *J Biol Chem* **267**:24886–24891.
- Janson RW, Hance KR, and Arend WP (1991) Production of IL-1 receptor antagonist by human in vitro-derived macrophages. Effects of lipopolysaccharide and granulocyte-macrophage colony-stimulating factor. *J Immunol* **147**:4218–4223.
- Jeziorska M, McCollum C, and Woolley DE (1997) Mast cell distribution, activation and phenotype in atherosclerotic lesions of human carotid arteries. *J Pathol* **182**:115–122.
- Jeziorska M, McCollum C, and Woolley DE (1998) Calcification in atherosclerotic plaque of human carotid arteries: associations with mast cells and macrophages. *J Pathol* **185**:10–17.
- Jirik FR, Podor TJ, Hirano T, Kishimoto T, Loskutoff DJ, Carson DA, and Lotz M (1989) Bacterial lipopolysaccharide and inflammatory mediators augment IL-6 secretion by human endothelial cells. *J Immunol* **142**:144–147.
- Johnson DR, Douglas I, Jahnke A, Ghosh S, and Pober JS (1996) A sustained reduction in IkappaB-beta may contribute to persistent NF-kappaB activation in human endothelial cells. *J Biol Chem* **271**:16317–16322.
- Jones SA, Horiuchi S, Topley N, Yamamoto N, and Fuller GM (2001) The soluble interleukin 6 receptor: mechanisms of production and implications in disease. *FASEB J* **15**:43–58.
- Jordan NJ, Watson ML, Williams RJ, Roach AG, Yoshimura T, and Westwick J (1997) Chemokine production by human vascular smooth muscle cells: modulation by IL-13. *Br J Pharmacol* **122**:749–757.
- Jorgensen C, Apparailly F, Couret I, Canovas F, Jacquet C, and Sany J (1998) Interleukin-4 and interleukin-10 are chondroprotective and decrease monocyte cell recruitment in human rheumatoid synovium in vivo. *Immunology* **93**:518–523.
- Jostock T, Mullberg J, Ozbek S, Atreya R, Blinn G, Voltz N, Fischer M, Neurath MF, and Rose-John S (2001) Soluble gp130 is the natural inhibitor of soluble interleukin-6 receptor transsignaling responses. *Eur J Biochem* **268**:160–167.
- Jovanovic DV, Di Battista JA, Martel-Pelletier J, Jolicoeur FC, He Y, Zhang M, Mineau F, and Pelletier JP (1998) IL-17 stimulates the production and expression of proinflammatory cytokines, IL-beta and TNF-alpha, by human macrophages. *J Immunol* **160**:3513–3521.
- Jovanovic DV, Di Battista JA, Martel-Pelletier J, Reboul P, He YL, Jolicoeur FC, and Pelletier JP (2001) Modulation of TIMP-1 synthesis by antiinflammatory cytokines and prostaglandin E-2 in interleukin 17 stimulated human monocytes/macrophages. *J Rheumatol* **28**:712–718.
- Kaartinen M, Penttila A, and Kovanen PT (1994a) Accumulation of activated mast cells in the shoulder region of human coronary atheroma, the predilection site of atheromatous rupture. *Circulation* **90**:1669–1678.
- Kaartinen M, Penttila A, and Kovanen PT (1994b) Mast cells of two types differing in neutral protease composition in the human aortic intima. Demonstration of tryptase- and tryptase/chymase-containing mast cells in normal intima, fatty streaks and the shoulder region of atheroma. *Arterioscler Thromb* **14**:966–972.
- Kaartinen M, Penttila A, and Kovanen PT (1996a) Mast cells accompany microvesicles in human coronary atheromas: implications for intimal neovascularization and hemorrhage. *Atherosclerosis* **123**:123–131.
- Kaartinen M, Penttila A, and Kovanen PT (1996b) Mast cells in rupture-prone areas of human coronary atheromas produce and store TNF-alpha. *Circulation* **94**:2787–2792.
- Kaartinen M, van der Wal AC, van der Loos CM, Piek JJ, Koch KT, Becker AE, and Kovanen PT (1998) Mast cell infiltration in acute coronary syndromes: implications for plaque rupture. *J Am Coll Cardiol* **32**:606–612.
- Kadokami T, McTiernan CF, Kubota T, Frye CS, Bounoutas GS, Robbins PD, Watkins SC, and Feldman AM (2001) Effects of soluble TNF receptor treatment on lipopolysaccharide-induced myocardial cytokine expression. *Am J Physiol* **280**:H2281–H2291.
- Kalvakolanu DV (1999) Virus interception of cytokine-regulated pathways. *Trends Microbiol* **7**:166–171.
- Kastrati A, Koch W, Berger PB, Mehilli J, Stephenson K, Neumann FJ, von Beckerath N, Böttiger C, Duff GW, and Schömig A (2000) Protective role against restenosis from an interleukin-1 receptor antagonist gene polymorphism in patients treated with coronary stenting. *J Am Coll Cardiol* **36**:2168–2173.
- Kato A, Odamaki M, Takita T, Maruyama Y, Kumagai H, and Hishida A (2002) Association between interleukin-6 and carotid atherosclerosis in hemodialysis patients. *Kidney Int* **61**:1143–1152.
- Khwah-Goodall Y, Wadhwa C, Stein BN, Gamble JR, and Vadas MA (1999) Stat6 activation is essential for interleukin-4 induction of P-selectin transcription in

- human umbilical vein endothelial cells. *Arterioscler Thromb Vasc Biol* **19**:1421–1429.
- Kim KN, Watanabe S, Ma Y, Thornton S, Giannini EH, and Hirsch R (2000a) Viral IL-10 and soluble TNF receptor act synergistically to inhibit collagen-induced arthritis following adenovirus-mediated gene transfer. *J Immunol* **164**:1576–1581.
- Kim SH, Eisenstein M, Reznikov L, Fantuzzi G, Novick D, Rubinstein M, and Dinarello CA (2000b) Structural requirements of six naturally occurring isoforms of the IL-18 binding protein to inhibit IL-18. *Proc Natl Acad Sci USA* **97**:1190–1195.
- King VL, Szilvassy SJ, and Daugherty A (2002) Interleukin-4 deficiency decreases atherosclerotic lesion formation in site-specific manner in female LDL receptor<sup>-/-</sup> mice. *Arterioscler Thromb Vasc Biol* **22**:456–461.
- Kingston PA, Sinha S, David A, Castro MG, Lowenstein PR, and Heagerty AM (2001) Adenovirus-mediated gene transfer of a secreted transforming growth factor- $\beta$  type II receptor inhibits luminal loss and constrictive remodeling after coronary angioplasty and enhances adventitial collagen deposition. *Circulation* **104**:2595–2601.
- Kishikawa H, Shimokama T, and Watanabe T (1993) Localization of T lymphocytes and macrophages expressing IL-1, IL-2 receptor, IL-6 and TNF in human aortic intima. Role of cell-mediated immunity in human atherogenesis. *Virchows Arch A Pathol Anat Histopathol* **423**:433–442.
- Kitamura T, Sato N, Arai K, and Miyajima A (1991) Expression cloning of the human IL-3 receptor cDNA reveals a shared beta subunit for the human IL-3 and GM-CSF receptors. *Cell* **66**:1165–1174.
- Klouche M, Bhakdi S, Hemmes M, and Rose-John S (1999) Novel path to activation of vascular smooth muscle cells: up-regulation of gp130 creates an autocrine activation loop by IL-6 and its soluble receptor. *J Immunol* **163**:4583–4589.
- Klouche M, Rose-John S, Schmiedt W, and Bhakdi S (2000) Enzymatically degraded, nonoxidized LDL induces human vascular smooth muscle cell activation, foam cell transformation and proliferation. *Circulation* **101**:1799–1805.
- Koch AE, Polverini PJ, Kunkel SL, Harlow LA, DiPietro LA, Elner VM, Elner SG, and Strieter RM (1992) Interleukin-8 as a macrophage-derived mediator of angiogenesis. *Science (Wash DC)* **258**:1798–1801.
- Kopf M, Baumann H, Freer G, Freudenberg M, Lamers M, Kishimoto T, Zinkernagel R, Bluethmann H, and Kohler G (1994) Impaired immune and acute-phase responses in interleukin-6-deficient mice. *Nature (Lond)* **368**:339–342.
- Kopf M, Brombacher F, Hodgkin PD, Ramsay AJ, Milbourne EA, Dai WJ, Ovington KS, Behm CA, Kohler G, Young IG, and Matthaei KI (1996) IL-5-deficient mice have a developmental defect in CD5<sup>+</sup> B-1 cells and lack eosinophilia but have normal antibody and cytotoxic T cell responses. *Immunity* **4**:15–24.
- Koprak S, Staruch MJ, and Dumont FJ (1999) A specific inhibitor of the p38 mitogen activated protein kinase affects differentially the production of various cytokines by activated human T cells: dependence on CD28 signaling and preferential inhibition of IL-10 production. *Cell Immunol* **192**:87–95.
- Korpelainen EI, Gamble JR, Smith WB, Dottore M, Vadas MA, and Lopez AF (1995) Interferon-gamma up-regulates interleukin-3 (IL-3) receptor expression in human endothelial cells and synergizes with IL-3 in stimulating major histocompatibility complex class II expression and cytokine production. *Blood* **86**:176–182.
- Kovanen PT (1997) Chymase-containing mast cells in human arterial intima: implications for atherosclerotic disease. *Heart Vessels (Suppl)*:12125–12127.
- Kovanen PT, Kaartinen M, and Paaonien T (1995) Infiltrates of activated mast cells at the site of coronary atheromatous erosion or rupture in myocardial infarction. *Circulation* **92**:1084–1088.
- Kragel AH, Travis WD, Steis RG, Rosenberg SA, and Roberts WC (1990) Myocarditis or acute myocardial infarction associated with interleukin-2 therapy for cancer. *Cancer* **66**:1513–1516.
- Krakauer T (1995) IL-10 inhibits the adhesion of leukocytic cells to IL-1-activated human endothelial cells. *Immunol Lett* **45**:61–65.
- Krasinski K, Spyridopoulos I, Kearney M, and Losordo DW (2001) In vivo blockade of tumor necrosis factor- $\alpha$  accelerates functional endothelial recovery after balloon angioplasty. *Circulation* **104**:1754–1756.
- Krishnaswamy G, Kelley J, Yerra L, Smith JK, and Chi DS (1999) Human endothelium as a source of multifunctional cytokines: molecular regulation and possible role in human disease. *J Interferon Cytokine Res* **19**:91–104.
- Ku G, Faust T, Lauffer LL, Livingston DJ, and Harding MW (1996) Interleukin-1 beta converting enzyme inhibition blocks progression of type II collagen-induced arthritis in mice. *Cytokine* **8**:377–386.
- Kubota T, Bounoutas GS, Miyagishima M, Kadokami T, Sanders VJ, Bruton C, Robbins PD, McTiernan CF, and Feldman AM (2000) Soluble tumor necrosis factor receptor abrogates myocardial inflammation but not hypertrophy in cytokine-induced cardiomyopathy. *Circulation* **101**:2518–2525.
- Kuiper J, von der Thüsen J, de Vos P, Fekkes ML, van Snick J, Biessen EAL, and van Berkel Th (2001) Interleukin 9 treatment of LDL receptor deficient mice inhibits atherosclerotic plaque formation (abstract). *Circulation* **104**:II-320.
- Kurt-Jones EA, Hamberg S, Ohara J, Paul WE, and Abbas AK (1987) Heterogeneity of helper/inducer T lymphocytes. I. Lymphokine production and lymphokine responsiveness. *J Exp Med* **166**:1774–1787.
- Lacruz S, Nicod LP, Chicheportiche R, Welgus HG, and Dayer JM (1995) IL-10 inhibits metalloproteinase and stimulates TIMP-1 production in human mononuclear phagocytes. *J Clin Investig* **96**:2304–2310.
- Lafaille JJ (1998) The role of helper T cell subsets in autoimmune diseases. *Cytokine Growth Factor Rev* **9**:139–151.
- Lalani AS, Graham K, Mossman K, Rajarathnam K, Clark-Lewis I, Kelvin D, and McFadden G (1997a) The purified myxoma virus gamma interferon receptor homolog M-T7 interacts with the heparin-binding domains of chemokines. *J Virol* **71**:4356–4363.
- Lalani I, Bhol K, and Ahmed AR (1997b) Interleukin-10: biology, role in inflammation and autoimmunity. *Ann Allergy Asthma Immunol* **79**:469–483.
- Lang R, Patel D, Morris JJ, Rutschman RL, and Murray PJ (2002) Shaping gene expression in activated and resting primary macrophages by IL-10. *J Immunol* **169**:2253–2263.
- Laurat E, Poirier B, Tupin E, Caligiuri G, Hansson G, K Bariety J, and Nicoletti A (2001) In vivo down-regulation of T helper cell 1 immune responses reduces atherosclerosis in apolipoprotein E-knockout mice. *Circulation* **104**:197–202.
- Leavitt MC, Yu G, Zhou C, and Barber JR (2000) Inhibition of interleukin-1 beta (IL-1 beta) production in human cells by ribozymes against IL-1 beta and IL-1 beta converting enzyme (ICE). *Antisense Nucleic Acid Drug Dev* **10**:409–414.
- Lebedeva I and Stein CA (2001) Antisense oligonucleotides: Promise and reality. *Ann Rev Pharmacol Toxicol* **41**:403–419.
- LeBoeuf RC and Schreyer SA (1998) The role of tumor necrosis factor- $\alpha$  receptors in atherosclerosis. *Trends Cardiovasc Med* **8**:131–138.
- Lee J, Ho WH, Maruoka M, Corpez RT, Baldwin DT, Foster JS, Goddard AD, Yansura DG, Vandlen RL, Wood WI, and Gurney AL (2001a) IL-17E, a novel proinflammatory ligand for the IL-17 receptor homolog IL-17Rrh1. *J Biol Chem* **276**:1660–1664.
- Lee TS, Yen HC, Pan CC, and Chau LY (1999) The role of interleukin 12 in the development of atherosclerosis in ApoE-deficient mice. *Arterioscler Thromb Vasc Biol* **19**:734–742.
- Lee YW, Kuhn H, Kaiser S, Hennig B, Daugherty A, and Toborek M (2001b) Interleukin 4 induces transcription of the 15-lipoxygenase I gene in human endothelial cells. *Journal of Lipid Research* **42**:783–791.
- Lejeune D, Demoulin JB, and Renaud JC (2001) Interleukin 9 induces expression of three cytokine signal inhibitors: cytokine-inducible SH2-containing protein, suppressor of cytokine signalling (SOCS)-2 and SOCS-3, but only SOCS-3 overexpression suppresses interleukin 9 signalling. *Biochem. J* **353**:109–116.
- Leland P, Taguchi J, Husain SR, Kreitman RJ, Pastan I, and Puri RK (2000) Human breast carcinoma cells express type II IL-4 receptors and are sensitive to antitumor activity of a chimeric IL-4-Pseudomonas exotoxin fusion protein in vitro and in vivo. *Mol Med* **6**:165–178.
- LeMaistre CF, Saleh MN, Kuzel TM, Foss F, Platanias LC, Schwartz G, Ratain M, Rook A, Freytes CO, Craig F, Reuben J, and Nichols JC (1998) Phase I trial of a ligand fusion-protein (DAB389IL-2) in lymphomas expressing the receptor for interleukin-2. *Blood* **91**:399–405.
- Leng SX and Elias JA (1997) Interleukin-11 inhibits macrophage interleukin-12 production. *J Immunol* **159**:2161–2168.
- Lentsch AB, Shanley TP, Sarma V, and Ward PA (1997) In vivo suppression of NF-kappa B and preservation of I kappa B alpha by interleukin-10 and interleukin-13. *J Clin Investig* **100**:2443–2448.
- Leonard WJ and Lin JX (2000) Cytokine receptor signaling pathways. *J Allergy Clin Immunol* **105**:877–888.
- Levitzi A (1990) Tyrosophostins—potential antiproliferative agents and novel molecular tools. *Biochem Pharmacol* **40**:913–918.
- Li H, Cybulsky MI, Gimbrone MA, and Libby P (1993) An atherogenic diet rapidly induces VCAM-1, a cytokine-regulatable mononuclear leukocyte adhesion molecule, in rabbit aortic endothelium. *Arterioscler Thromb* **13**:197–204.
- Li HG and Forstermann U (2000) Nitric oxide in the pathogenesis of vascular disease. *J Pathol* **190**:244–254.
- Li HZ, Chen J, Huang A, Stinson J, Heldens S, Foster J, Dowd P, Gurney AL, and Wood WI (2000) Cloning and characterization of IL-17B and IL-17C, two new members of the IL-17 cytokine family. *Proc Natl Acad Sci USA* **97**:773–778.
- Li J, Perrella MA, Tsai JK, Yet SF, Hsieh CM, Yoshizumi M, Patterson C, Endege WO, Zhou F, and Lee ME (1995) Induction of vascular endothelial growth factor gene expression by interleukin-1 beta in rat aortic smooth muscle cells. *J Biol Chem* **270**:308–312.
- Liao HS, Matsumoto A, Itakura H, Doi T, Honda M, Kodama T, and Geng YJ (1999) Transcriptional inhibition by interleukin-6 of the class A, macrophage scavenger receptor in macrophages derived from human peripheral monocytes and the THP-1 monocytic cell line. *Arterioscler Thromb Vasc Biol* **19**:1872–1880.
- Libby P, Sukhova G, Lee RT, and Galis ZS (1995) Cytokines regulate vascular functions related to stability of the atherosclerotic plaque. *J Cardiovasc Pharmacol* **25**:S9–S12.
- Lin JX and Leonard WJ (2000) The role of Stat5a and Stat5b in signaling by IL-2 family cytokines. *Oncogene* **19**:2566–2576.
- Lin JX, Migone TS, Tsang M, Friedmann M, Weatherbe JA, Zhou L, Yamauchi A, Bloom ET, Mietz J, John S, and Leonard WJ (1995) The role of shared receptor motifs and common Stat proteins in the generation of cytokine pleiotropy and redundancy by IL-2, IL-4, IL-7, IL-13 and IL-15. *Immunity* **2**:331–339.
- Lindner H, Holler E, Gerbitz A, Johnson JP, Bornkamm GW, and Eissner G (1997) Influence of bacterial endotoxin on radiation-induced activation of human endothelial cells in vitro and in vivo: interleukin-10 protects against transendothelial migration. *Transplantation* **64**:1370–1373.
- Ling P, Gately MK, Gubler U, Stern AS, Lin P, Hollfelder K, Su C, Pan YC, and Hakim J (1995) Human IL-12 p40 homodimer binds to the IL-12 receptor but does not mediate biologic activity. *J Immunol* **154**:116–127.
- Liu B, Liao J, Rao X, Kushner SA, Chung CD, Chang DD, and Shuai K (1998) Inhibition of Stat1-mediated gene activation by PIAS1. *Proc Natl Acad Sci USA* **95**:10626–10631.
- Liu L, Lalani A, Dai E, Seet B, Macauley C, Singh R, Fan L, McFadden G, and Lucas A (2000) The viral anti-inflammatory chemokine-binding protein M-T7 reduces intimal hyperplasia after vascular injury. *J Clin Investig* **105**:1613–1621.
- Livingston DJ (1997) In vitro and in vivo studies of ICE inhibitors. *J Cell Biochem* **64**:19–26.
- Lockyer JM, Colladay JS, Alperin-Lea WL, Hammond T, and Buda AJ (1998) Inhibition of nuclear factor-kappaB-mediated adhesion molecule expression in human endothelial cells. *Circ Res* **82**:314–320.
- Loppnow H and Libby P (1989a) Comparative analysis of cytokine induction in human vascular endothelial and smooth muscle cells. *Lymphokine Res* **8**:293–299.
- Loppnow H and Libby P (1989b) Adult human vascular endothelial cells express the IL6 gene differentially in response to LPS or IL1. *Cell Immunol* **122**:493–503.
- Loppnow H and Libby P (1990) Proliferating or interleukin 1-activated human

- vascular smooth muscle cells secrete copious interleukin 6. *J Clin Invest* 85:731-738.
- Loppnow H and Libby P (1992) Functional significance of human vascular smooth muscle cell-derived interleukin 1 in paracrine and autocrine regulation pathways. *Exp Cell Res* 198:283-290.
- Losman JA, Chen XP, Hilton D, and Rothman P (1999) Cutting edge: SOCS-1 is a potent inhibitor of IL-4 signal transduction. *J Immunol* 162:3770-3774.
- Louis A, Cleland JG, Crabbe S, Ford S, Thackray S, Houghton T, and Clark A (2001) Clinical Trials Update: CAPRICORN, COPERNICUS, MIRACLE, STAF, RITZ-2, RECOVER and RENAISSANCE and cachexia and cholesterol in heart failure. Highlights of the Scientific Sessions of the American College of Cardiology, 2001. *Eur J Heart Fail* 3:381-387.
- Lubberts E, Joosten LA, Van Den Berselaar L, Helsen MM, Bakker AC, Xing Z, Richards CD, and Van Den Berg WB (2000) Intra-articular IL-10 gene transfer regulates the expression of collagen-induced arthritis (CIA) in the knee and ipsilateral paw. *Clin Exp Immunol* 120:375-383.
- Lutgens E, Cleutjens KB, Heeneman S, Kotliansky VE, Burkly LC, and Daemen MJ (2000) Both early and delayed anti-CD40L antibody treatment induces a stable plaque phenotype. *Proc Natl Acad Sci USA* 97:7464-7469.
- Lynch CM, Hara PS, Leonard JC, Williams JK, Dean RH, and Geary RL (1997) Adeno-associated virus vectors for vascular gene delivery. *Circ Res* 80:497-505.
- Mach F, Schonbeck U, Sukhova GK, Atkinson E, and Libby P (1998) Reduction of atherosclerosis in mice by inhibition of CD40 signalling. *Nature (Lond)* 394:200-203.
- Mach F, Schonbeck U, Sukhova GK, Bourcier T, Bonnefoy JY, Pober JS, and Libby P (1997) Functional CD40 ligand is expressed on human vascular endothelial cells, smooth muscle cells and macrophages: implications for CD40-CD40 ligand signaling in atherosclerosis. *Proc Natl Acad Sci USA* 94:1931-1936.
- Mahboubi K, Biedermann BC, Carroll JM, and Pober JS (2000) IL-11 activates human endothelial cells to resist immune-mediated injury. *J Immunol* 164:3837-3846.
- Mahboubi K, Li FZ, Plescia J, Kirkiles-Smith NC, Mesri M, Du YF, Carroll JM, Elias JA, Altieri DC, and Pober JS (2001) Interleukin-11 up-regulates survivin expression in endothelial cells through a signal transducer and activator of transcription-3 pathway. *Lab Invest* 81:327-334.
- Mahieu M, Deschuyteneer R, Forget D, Vandebussche P, and Content J (1994) Construction of a ribozyme directed against human interleukin-6 mRNA: evaluation of its catalytic activity in vitro and in vivo. *Blood*, 84:3758-3765.
- Maier JA, Voualalas P, Roeder D, and Maciag T (1990) Extension of the life-span of human endothelial cells by an interleukin-1 alpha antisense oligomer. *Science (Wash DC)* 249:1570-1574.
- Maier JA and Ragnotti G (1993) An oligomer targeted against protein kinase C alpha prevents interleukin-1 alpha induction of cyclooxygenase expression in human endothelial cells. *Exp Cell Res* 205:52-58.
- Maini A, Morse PD, Wang CY, Jones RF, and Haas GP (1997) New developments in the use of cytokines for cancer therapy. *Anticancer Res* 17:3803-3808.
- Mallat Z, Besnard S, Duriez M, Deleuze V, Emmanuel F, Bureau MF, Soubrier F, Esposito B, Duez H, Fievet C, Staels B, Duverger N, Scherman D, and Tedgui A (1999b) Protective role of interleukin-10 in atherosclerosis. *Circ Res* 85:e17-24.
- Mallat Z, Corbaz A, Scoazec A, Besnard S, Lesèche G, Chvatchko Y, and Tedgui A (2001a) Expression of Interleukin-18 in Human Atherosclerotic Plaques and Relation to Plaque Instability. *Circulation* 104:1598-1603.
- Mallat Z, Corbaz A, Scoazec A, Graber P, Alouani S, Esposito B, Humbert Y, Chvatchko Y, and Tedgui A (2001b) Interleukin-18/interleukin-18 binding protein signaling modulates atherosclerotic lesion development and stability. *Circ Res* 89:e41-e45.
- Mallat Z, Gobjova A, Marchiol-Fournigault C, Esposito B, Kamate C, Merval R, Fratellizi D, and Tedgui A (2001c) Inhibition of transforming growth factor-beta signaling accelerates atherosclerosis and induces an unstable plaque phenotype in mice. *Circ Res* 89:930-934.
- Mallat Z, Heymes C, Ohan J, Faggin E, Leseche G, and Tedgui A (1999a) Expression of interleukin-10 in advanced human atherosclerotic plaques: relation to inducible nitric oxide synthase expression and cell death. *Arterioscler Thromb Vasc Biol* 19:611-616.
- Mallat Z, Silvestre JS, Le Ricousse-Roussanne S, Lecomte-Raclet L, Corbaz A, Clergue M, Duriez M, Barateau V, Akira S, Tedgui A, Tobelem G, Chvatchko Y, Levy BI (2002) Interleukin-18/interleukin-18 binding protein signaling modulates ischemia-induced neovascularization in mice hindlimb. *Circ Res* 91:441-448.
- Mann MJ, Gibbons GH, Hutchinson H, Poston RS, Hoyt EG, Robbins RC, and Dzau VJ (1999) Pressure-mediated oligonucleotide transfection of rat and human cardiovascular tissues. *Proc Natl Acad Sci USA* 96:6411-6416.
- Mattner F, Ozmen L, Podlaski FJ, Wilkinson VL, Presky DH, Gately MK, and Alber G (1997) Treatment with homodimeric interleukin-12 (IL-12) p40 protects mice from IL-12-dependent shock but not from tumor necrosis factor alpha-dependent shock. *Infect Immun* 65:4734-4737.
- Maus UA, Herold S, Schlingensiepen KH, Schlingensiepen R, Dormayr T, Rosseau S, Maus R, Seeger W, and Lohmeyer J (2000) Antisense oligomers for selective suppression of MCP-1 synthesis in human pulmonary endothelial cells. *Antisense Nucleic Acid Drug Dev* 10:185-193.
- Mazzone A, De Servi S, Vezzoli M, Fossati G, Mazzucchelli I, Gritti D, Ottini E, Mussini A, and Specchia G (1999) Plasma levels of interleukin 2, 6, 10 and phenotypic characterization of circulating T lymphocytes in ischemic heart disease. *Atherosclerosis* 145:369-374.
- McFadden G, Lalani A, Everett H, Nash P, and Xu X (1998) Virus-encoded receptors for cytokines and chemokines. *Semin Cell Dev Biol* 9:359-368.
- McLane MP, Haczk A, van de Rijn M, Weiss C, Ferrante V, MacDonald D, Renaud JC, Nicolaides NC, Holroyd KJ, and Levitt RC (1998) Interleukin-9 promotes allergen-induced eosinophilic inflammation and airway hyperresponsiveness in transgenic mice. *Am J Respir Cell Mol Biol* 19:713-720.
- Meager A (1999) Cytokine regulation of cellular adhesion molecule expression in inflammation. *Cytokine Growth Factor Rev* 10:27-39.
- Mihara M, Kotoh M, Nishimoto N, Oda Y, Kumagai E, Takagi N, Tsunemi K, Ohsugi Y, Kishimoto T, Yoshizaki K, and Takeda Y (2001) Humanized antibody to human interleukin-6 receptor inhibits the development of collagen arthritis in cynomolgus monkeys. *Clin Immunol* 98:319-326.
- Mijatovic T, Krays V, Caput D, Defrance P, and Huez G (1997) Interleukin-4 and -13 inhibit tumor necrosis factor-alpha mRNA translational activation in lipopolysaccharide-induced mouse macrophages. *J Biol Chem* 272:14394-14398.
- Miller BE, Krasney PA, Gauvin DM, Holbrook KB, Koontz DJ, Abruzzese RV, Miller RE, Pagani KA, Dolle RE, Ator MA, and Gilman SC (1995) Inhibition of mature IL-1 beta production in murine macrophages and a murine model of inflammation by WIN 67694, an inhibitor of IL-1 beta converting enzyme. *J Immunol* 154:1331-1338.
- Miller DD, Bach RG, Tio FO, Bailey SR, Waters CA, Woodworth TG, Nichols JC, Paige SB, and Farrar M (1996) Interleukin-2 receptor-specific fusion toxin inhibits barotrauma-induced arterial atherosclerosis. *Atherosclerosis* 126:1-14.
- Minter RM, Ferry MA, Rectenwald JE, Bahjat FR, Oberholzer A, Oberholzer C, La Face D, Tsai V, Ahmed CMI, Hutchings B, et al. (2001) Extended lung expression and increased tissue localization of viral IL-10 with adenoviral gene therapy. *Proc Natl Acad Sci USA* 98:277-282.
- Miyazaki T, Kawahara A, Fujii H, Nakagawa Y, Minami Y, Liu ZJ, Oishi I, Silvenoinen O, Withuhn BA, Ihle JN, and Taniguchi T (1994) Functional activation of Jak1 and Jak3 by selective association with IL-2 receptor subunits. *Science (Wash DC)* 266:1045-1047.
- Mizia-Stec K, Mandecki T, Zahorska-Markiewicz B, Janowska J, Szulc A, Jastrzebska-Maj E, Szymanski L, and Majewski T (2002) Selected cytokines and soluble forms of cytokine receptors in coronary artery disease. *Eur J Intern Med* 13:115-122.
- Modur V, Li Y, Zimmerman GA, Prescott SM, and McIntyre TM (1997) Retrograde inflammatory signaling from neutrophils to endothelial cells by soluble interleukin-6 receptor alpha. *J Clin Invest* 100:2752-2756.
- Molenaar TJM Twisk J, Appeldoorn CAM, de Haas SAM, Michon I, van Berkel ThJC Kuiper J, and Biessen EA L (2001) Peptide antagonists to P-selectin: potential in anti-atherothrombotic therapy. *Circulation* 104:II-38-39.
- Momiyama Y, Hirano R, Taniguchi H, Nakamura H, and Ohsuzu F (2001) Effects of interleukin-1 gene polymorphisms on the development of coronary artery disease associated with Chlamydia pneumoniae infection. *J Am Coll Cardiol* 38:712-717.
- Monahan PE and Samulski RJ (2000) Adeno-associated virus vectors for gene therapy: more pros than cons? *Mol Med Today* 6:433-440.
- Monajemi H, Arkenbout EK, and Pannekoek H (2001) Gene expression in atherogenesis. *Thromb Haemostasis* 86:404-412.
- Moore KW, Malefyt RD, Coffman RL, and O'Garra A (2001) Interleukin-10 and the interleukin-10 receptor. *Annu Rev Immunol* 19:683-765.
- Moreau M, Brocheriou I, Petit L, Ninio E, Chapman MJ, and Rouis M (1999) Interleukin-8 mediates down-regulation of tissue inhibitor of metalloproteinase-1 expression in cholesterol-loaded human macrophages: relevance to stability of atherosclerotic plaque. *Circulation* 99:420-426.
- Moreland L, Gugliotti R, King K, Chase W, Weisman M, Greco T, Fife R, Korn J, Simms R, Tesser J, et al. (2001) Results of a phase-III randomized, masked, placebo-controlled trial of recombinant human interleukin-11 (rhIL-11) in the treatment of subjects with active rheumatoid arthritis. *Arthritis Res* 3:247-252.
- Morishita R, Gibbons GH, Horiuchi M, Kaneda Y, Ogihara T, and Dzau VJ (1998) Role of AP-1 complex in angiotensin II-mediated transforming growth factor-beta expression and growth of smooth muscle cells: using decoy approach against AP-1 binding site. *Biochem Biophys Res Commun* 243:361-367.
- Morishita R, Gibbons GH, Kaneda Y, Ogihara T, and Dzau VJ (1994) Pharmacokinetics of antisense oligodeoxyribonucleotides (cyclin B1 and CDC 2 kinase) in the vessel wall in vivo: enhanced therapeutic utility for restenosis by HIV-liposome delivery. *Gene (Amst)* 149:13-19.
- Moser R, Schleiffenbaum B, Groscurth P, and Fehr J (1989) Interleukin 1 and tumor necrosis factor stimulate human vascular endothelial cells to promote transendothelial neutrophil passage. *J Clin Invest* 83:444-455.
- Moyer CF, Sajuthi D, Tulli H, and Williams JK (1991) Synthesis of IL-1 alpha and IL-1 beta by arterial cells in atherosclerosis. *Am J Pathol* 138:951-960.
- Mtairag E, Chollet-Martin S, Oudghiri M, Laquay N, Jacob MP, Michel JB, and Feldman LJ (2001) Effects of interleukin-10 on monocyte/endothelial cell adhesion and MMP-9/TIMP-1 secretion. *Cardiovasc Res* 49:882-890.
- Muchamuel T, Menon S, Pisacane P, Howard MC, and Cockayne DA (1997) IL-13 protects mice from lipopolysaccharide-induced lethal endotoxemia: correlation with down-modulation of TNF-alpha, IFN-gamma and IL-12 production. *J Immunol* 158:2898-2903.
- Mufson RA, Szabo J, and Eckert D (1992) Human IL-3 induction of c-jun in normal monocytes is independent of tyrosine kinase and involves protein kinase C. *J Immunol* 148:1129-1135.
- Muhl H, Kampfer H, Bosmann M, Frank S, Radeke H, and Pfilschifter J (2000) Interferon-gamma mediates gene expression of IL-18 binding protein in nonleukocytic cells. *Biochem Biophys Res Commun* 267:960-963.
- Mukaida N, Shiroy M, and Matsushima K (1989) Genomic structure of the human monocyte-derived neutrophil chemotactic factor IL-8. *J Immunol* 143:1366-1371.
- Mullberg J, Durie FH, Otten-Evans C, Alderson MR, Rose-John S, Cosman D, Black RA, and Mohler KM (1995) A metalloprotease inhibitor blocks shedding of the IL-6 receptor and the p60 TNF receptor. *J Immunol* 155:5198-5205.
- Mullberg J, Rauch CT, Wolfson MF, Castner B, Fitzner JN, Otten-Evans C, Mohler KM, Cosman D, and Black RA (1997) Further evidence for a common mechanism for shedding of cell surface proteins. *FEBS Lett* 401:235-238.
- Muller-Newen G, Kuster A, Hemmann U, Keul R, Horsten U, Martens A, Graeve L, Wijdenes J, and Heinrich PC (1998) Soluble IL-6 receptor potentiates the antagonistic activity of soluble gp130 on IL-6 responses. *J Immunol* 161:6347-6355.
- Musso T, Calosso L, Zucca M, Millesimo M, Ravarino D, Giovarelli M, Malavasi F, Ponzi AN, Paus R, and Bulfone-Paus S (1999) Human monocytes constitutively express membrane-bound, biologically active and interferon-gamma-up-regulated interleukin-15. *Blood* 93:3531-3539.

- Nabata T, Morimoto S, Koh E, Shiraishi T, and Ogihara T (1990) Interleukin-6 stimulates c-myc expression and proliferation of cultured vascular smooth muscle cells. *Biochem Int* **20**:445–453.
- Nageh MF, Sandberg ET, Marotti KR, Lin AH, Melchior EP, Bullard DC, and Beaudet AL (1997) Deficiency of inflammatory cell adhesion molecules protects against atherosclerosis in mice. *Arterioscler Thromb Vasc Biol* **17**:1517–1520.
- Naka T, Fujimoto M, and Kishimoto T (1999) Negative regulation of cytokine signaling: STAT-induced STAT inhibitor. *Trends Biochem Sci* **24**:394–398.
- Naka T, Narazaki M, Hirata M, Matsumoto T, Minamoto S, Aono A, Nishimoto N, Kajita T, Taga T, Yoshizaki K, Akira S, and Kishimoto T (1997) Structure and function of a new STAT-induced STAT inhibitor. *Nature (Lond)* **387**:924–929.
- Nakashima Y, Raines EW, Plump AS, Breslow JL, and Ross R (1998) Up-regulation of VCAM-1 and ICAM-1 at atherosclerosis-prone sites on the endothelium in the ApoE-deficient mouse. *Arterioscler Thromb Vasc Biol* **18**:842–851.
- Napoli C, D'Armiento FP, Mancini FP, Postiglione A, Witztum JL, Palumbo G, and Palinski W (1997) Fatty streak formation occurs in human fetal aortas and is greatly enhanced by maternal hypercholesterolemia. Intimal accumulation of low density lipoprotein and its oxidation precede monocyte recruitment into early atherosclerotic lesions. *J Clin Invest* **100**:2680–2690.
- Narazaki M, Yasukawa K, Saito T, Ohnogi Y, Fukui H, Koishihara Y, Yancopoulos GD, Taga T, and Kishimoto T (1993) Soluble forms of the interleukin-6 signal-transducing receptor component gp130 in human serum possessing a potential to inhibit signals through membrane-anchored gp130. *Blood* **82**:1120–1126.
- Nassar GM, Morrow JD, Roberts LJ, 2<sup>nd</sup>, Lakakis FG, and Badr KF (1994) Induction of 15-lipoxygenase by interleukin-13 in human blood monocytes. *J Biol Chem* **269**:27631–27634.
- Nicklin MJ, Hughes DE, Barton JL, Ure JM, and Duff GW (2000) Arterial inflammation in mice lacking the interleukin 1 receptor antagonist gene. *J Exp Med* **191**:303–312.
- Nishinakamura R, Nakayama N, Hirabayashi Y, Inoue T, Aud D, McNeil T, Azuma S, Yoshida S, Toyoda Y, Arai K, Miyajima A, and Murray R (1995) Mice deficient for the IL-3/GM-CSF/IL-5 beta c receptor exhibit lung pathology and impaired immune response, while beta IL3 receptor-deficient mice are normal. *Immunity* **2**:211–222.
- Noguchi M, Nakamura Y, Russell SM, Ziegler SF, Tsang M, Cao X, and Leonard WJ (1993a) Interleukin-2 receptor gamma chain: a functional component of the interleukin-7 receptor. *Science (Wash DC)* **262**:1877–1880.
- Noguchi M, Yi H, Rosenblatt HM, Filipovich AH, Adelstein S, Modi WS, McBride OW, and Leonard WJ (1993b) Interleukin-2 receptor gamma chain mutation results in X-linked severe combined immunodeficiency in humans. *Cell* **73**:147–157.
- Novick D, Kim SH, Fantuzzi G, Reznikov LL, Dinarello CA, and Rubinstein M (1999) Interleukin-18 binding protein: a novel modulator of the Th1 cytokine response. *Immunity* **10**:127–136.
- Novick D, Schwartzburd B, Pinkus R, Suissa D, Belzer I, Sthoeger Z, Keane WF, Chvatchko Y, Kim SH, Fantuzzi G, et al. (2001) A novel IL-18BP ELISA shows elevated serum IL-18BP in sepsis and extensive decrease of free IL-18. *Cytokine* **14**:334–342.
- Ohtsubo M, Takayanagi A, Gamou S, and Shimizu N (2000) Interruption of NF-kappaB-STAT1 signaling mediates EGF-induced cell-cycle arrest. *J Cell Physiol* **184**:131–137.
- Okada M, Matsumori A, Ono K, Furukawa Y, Shioi T, Iwasaki A, Matsushima K, and Sasayama S (1998) Cyclic stretch up-regulates production of interleukin-8 and monocyte chemoattractant and activating factor/monocyte chemoattractant protein-1 in human endothelial cells. *Arterioscler Thromb Vasc Biol* **18**:894–901.
- Omrust SV and Wiseman LR (1999) Basiliximab. *Drugs* **57**:207–213.
- Opal SM and DePalo VA (2000) Anti-inflammatory cytokines. *Chest* **117**:1162–1172.
- Oppenheimer-Marks N, Brezinschek RI, Mohamadzeid M, Vita R, and Lipsky PE (1998) Interleukin 15 is produced by endothelial cells and increases the transendothelial migration of T cells in vitro and in the SCID mouse-human rheumatoid arthritis model in vivo. *J Clin Invest* **101**:1261–1272.
- Oppmann B, Lesley R, Blom B, Timans JC, Xu YM, Hunte B, Vega F, Yu N, Wang J, Singh K, et al. (2000) Novel p19 protein engages IL-12p40 to form a cytokine, IL-23, with biological activities similar as well as distinct from IL-12. *Immunity* **13**:715–725.
- Osborn L, Hession C, Tizard R, Vassallo C, Luhowskyj S, Chi-Rosso G, and Lobb R (1989) Direct expression cloning of vascular cell adhesion molecule 1, a cytokine-induced endothelial protein that binds to lymphocytes. *Cell* **59**:1203–1211.
- Owens RJ and Lumb S (2000) Therapeutic regulation of cytokine signalling by inhibitors of p38 mitogen-activated protein kinase, in *Novel Cytokine Inhibitors* (Higgs GA and Henderson B eds) pp 201–215. Birkhäuser Verlag AG, Basel.
- Oyama J, Shimokawa H, Morita S, Yasui H, and Takeshita A (2001) Elevated interleukin-1beta in pericardial fluid of patients with ischemic heart disease. *Coron Artery Dis* **12**:567–571.
- Palmer Crocker RL, Hughes CC, and Pober JS (1996) IL-4 and IL-13 activate the JAK2 tyrosine kinase and Stat6 in cultured human vascular endothelial cells through a common pathway that does not involve the gamma c chain. *J Clin Invest* **98**:604–609.
- Parham C, Chirica M, Timans J, Vaisberg E, Travis M, Cheung J, Pflanz S, Zhang R, Singh KP, Vega F, et al. (2002) A receptor for the heterodimeric cytokine IL-23 is composed of IL-12Rbeta1 and a novel cytokine receptor subunit, IL-23R. *J Immunol* **168**:5699–5708.
- Paul WE (1991) Interleukin-4: a prototypic immunoregulatory lymphokine. *Blood* **77**:1859–1870.
- Paulose M, Bennett BL, Manning AM, and Essani K (1998) Selective inhibition of TNF-alpha induced cell adhesion molecule gene expression by tanapox virus. *Microb Pathog* **25**:33–41.
- Pennica D, Shaw KJ, Swanson TA, Moore MW, Shelton DL, Zioncheck KA, Rosenthal A, Taga T, Paoni NF, and Wood WI (1995) Cardiotrophin-1. Biological activities and binding to the leukemia inhibitory factor receptor/gp130 signaling complex. *J Biol Chem* **270**:10915–10922.
- Perkins AS (2002) Functional genomics in the mouse. *Funct Integr Genomics* **2**:81–91.
- Perlmutter DH (1989) IFN beta 2/IL-6 is one of several cytokines that modulate acute phase gene expression in human hepatocytes and human macrophages. *Ann N Y Acad Sci* **557**:332–557341.
- Pflanz S, Timans JC, Cheung J, Rosales R, Kanzler H, Gilbert J, Hibbert L, Churakov T, Travis M, Vaisberg E, et al. (2002) IL-27, a heterodimeric cytokine composed of EB13 and p28 protein, induces proliferation of naive CD4(+) T cells. *Immunity* **16**:779–790.
- Pierce JW, Schoenleber R, Jesmok G, Best J, Moore SA, Collins T, and Gerritsen ME (1997) Novel inhibitors of cytokine-induced IkappaBalpha phosphorylation and endothelial cell adhesion molecule expression show anti-inflammatory effects in vivo. *J Biol Chem* **272**:21096–21103.
- Pilson RS, Levin W, Desai B, Reik LM, Lin P, Korkmaz-Duffy E, Campbell E, Tso JY, Kerwin JA, and Hakimi J (1997) Bispecific humanized anti-IL-2 receptor alpha beta antibodies inhibitory for both IL-2- and IL-15-mediated proliferation. *J Immunol* **159**:1543–1556.
- Pinderski LJ, Fischbein MP, Subbanagounder G, Fishbein MC, Kubo N, Cheroutre H, Curtiss LK, Berliner JA, and Boisvert WA (2002) Overexpression of interleukin-10 by activated T, lymphocytes inhibits atherosclerosis in LDL receptor-deficient mice by altering lymphocyte and macrophage phenotypes. *Circ Res* **90**:1064–1071.
- Pinderski-Oslund LJ, Hedrick CC, Olvera T, Hagenbaugh A, Territo M, Berliner JA, and Fyfe AI (1999) Interleukin-10 blocks atherosclerotic events in vitro and in vivo. *Arterioscler Thromb Vasc Biol* **19**:2847–2853.
- Podor TJ, Jirik FR, Loskutoff DJ, Carson DA, and Lotz M (1989) Human endothelial cells produce IL-6. Lack of responses to exogenous IL-6. *Ann N Y Acad Sci* **557**:374–557385.
- Pomerantz BJ, Reznikov LL, Harken AH, and Dinarello CA (2001) Inhibition of caspase 1 reduces human myocardial ischemic dysfunction via inhibition of IL-18 and IL-1 beta. *Proc Natl Acad Sci USA* **98**:2871–2876.
- Porreca E, Di Febbo C, Barbacane RC, Panara MR, Cuccurullo F, and Conti P (1993) Effect of interleukin-1 receptor antagonist on vascular smooth muscle cell proliferation. *Atherosclerosis* **99**:71–78.
- Porreca E, Sergi R, Baccante G, Reale M, Orsini L, Febbo CD, Caselli G, Cuccurullo F, and Bertini R (1999) Peripheral blood mononuclear cell production of interleukin-8 and IL-8-dependent neutrophil function in hypercholesterolemic patients. *Atherosclerosis* **146**:345–350.
- Powell PP, Dixon LK, and Parkhouse RM (1996) An IkappaB homolog encoded by African swine fever virus provides a novel mechanism for down-regulation of proinflammatory cytokine responses in host macrophages. *J Virol.* **70**:8527–8533.
- Raizada MK, Francis SC, Wang HW, Gelband CH, Reaves PY, and Katovich MJ (2000) Targeting of the renin-angiotensin system by antisense gene therapy: a possible strategy for the long-term control of hypertension. *J Hypertension* **18**:353–362.
- Ramos CL, Huo YQ, Jung US, Ghosh S, Manka DR, Sarembock LJ, and Ley K (1999) Direct demonstration of P-selectin- and VCAM-1-dependent mononuclear cell rolling in early atherosclerotic lesions of apolipoprotein E-deficient mice. *Circ Res* **84**:1237–1244.
- Rauramaa R, Vaisanen SB, Luong LA, Schmidt-Trucksass A, Penttila IM, Bouchard C, Toyry J, and Humphries SE (2000) Stromelysin-1 and interleukin-6 gene promoter polymorphisms are determinants of asymptomatic carotid artery atherosclerosis. *Arterioscler Thromb Vasc Biol* **20**:2657–2662.
- Ray CA, Black RA, Kronheim SR, Greenstreet TA, Sleath PR, Salvanes GS, and Pickup DJ (1992) Viral inhibition of inflammation: cowpox virus encodes an inhibitor of the interleukin-1 beta converting enzyme. *Cell* **69**:597–604.
- Read MA, Whitley MZ, Williams AJ, and Collins T (1994) NF-kappa B and I kappa B alpha: an inducible regulatory system in endothelial activation. *J Exp Med* **179**:503–512.
- Reape TJ and Groot PH (1999) Chemokines and atherosclerosis. *Atherosclerosis* **147**:213–225.
- Recchia A, Parks RJ, Lamartina S, Toniatti C, Pieroni L, Palombo P, Ciliberto G, Graham FL, Cortese R, La Monica N, and Colloca S (1999) Site-specific integration mediated by a hybrid adenovirus/adeno-associated virus vector. *Proc Natl Acad Sci USA* **96**:2615–2620.
- Renauld JC, Goethals A, Houssiau F, Van Roost E, and Van Snick J (1990) Cloning and expression of a cDNA for the human homolog of mouse T cell and mast cell growth factor P40. *Cytokine* **2**:9–12.
- Renauld JC, van der Lugt N, Vink A, van Room M, Godfraind C, Warnier G, Merz H, Feller A, Berns A, and Van Snick J (1994) Thymic lymphomas in interleukin 9 transgenic mice. *Oncogene* **9**:1327–1332.
- Renz H (1999) Soluble interleukin-4 receptor (sIL-4R) in allergic diseases. *Inflamm Res* **48**:425–431.
- Revoltella RP (1998) Natural and therapeutically-induced antibodies to cytokines. *Biotherapy* **10**:321–331.
- Reznikov LL, Kim SH, Westcott JY, Frishman J, Fantuzzi G, Novick D, Rubinstein M, and Dinarello CA (2000) IL-18 binding protein increases spontaneous and IL-1-induced prostaglandin production via inhibition of IFN-gamma. *Proc Natl Acad Sci USA* **97**:2174–2179.
- Rich BE and Kupper TS (2001) Cytokines: IL-20—a new effector in skin inflammation. *Curr Biol* **11**:R531–R534.
- Richard M, Louahed J, Demoulin JB, and Renauld JC (1999) Interleukin-9 regulates NF-kappaB activity through BCL3 gene induction. *Blood* **93**:4318–4327.
- Ridker PM, Rifai N, Pfeffer M, Sacks F, Lepage S, and Braunwald E (2000a) Elevation of tumor necrosis factor-alpha and increased risk of recurrent coronary events after myocardial infarction. *Circulation* **101**:2149–2153.
- Ridker PM, Rifai N, Pfeffer MA, Sacks FM, Moye LA, Goldman S, Flaker GC, and Braunwald E (1998) Inflammation, pravastatin and the risk of coronary events after myocardial infarction in patients with average cholesterol levels. Cholesterol and Recurrent Events (CARE) Investigators. *Circulation* **98**:839–844.
- Ridker PM, Rifai N, Stampfer MJ, and Hennekens CH (2000b) Plasma concentration

- of interleukin-6 and the risk of future myocardial infarction among apparently healthy men. *Circulation* **101**:1767–1772.
- Romagnani S (2000) T-cell subsets (Th1 versus Th2). *Ann Allergy Asthma Immunol* **85**:9–18.
- Romano M, Sironi M, Toniatti C, Polentarutti N, Fruscella P, Ghezzi P, Faggioni R, Luani W, van Hinsbergh V, Sozzani S, Bussolino F, Poli V, Ciliberto G, and Mantovani A (1997) Role of IL-6 and its soluble receptor in induction of chemokines and leukocyte recruitment. *Immunity* **6**:315–325.
- Rosenfeld ME (1991) Oxidized LDL affects multiple atherogenic cellular responses. *Circulation* **83**:2137–2140.
- Rosenkilde MM, Kleidal TN, Brauner-Osborne H, and Schwartz TW (1999) Agonists and inverse agonists for the herpesvirus 8-encoded constitutively active seven-transmembrane oncogene product, ORF-74. *J Biol Chem* **274**:956–961.
- Ross R (1986) The pathogenesis of atherosclerosis—an update. *N Engl J Med* **314**:488–500.
- Ross R (1993a) The pathogenesis of atherosclerosis: a perspective for the 1990s. *Nature (Lond)* **362**:801–809.
- Ross R (1993b) Rous-Whipple Award Lecture. Atherosclerosis: a defense mechanism gone awry. *Am J Pathol* **143**:987–1002.
- Ross R (1999) Mechanisms of disease—atherosclerosis—an inflammatory disease. *N Engl J Med* **340**:115–126.
- Ross R and Glomset JA (1973) Atherosclerosis and the arterial smooth muscle cell: Proliferation of smooth muscle is a key event in the genesis of the lesions of atherosclerosis. *Science (Wash DC)* **180**:1332–1339.
- Roth M, Nauck M, Tamm M, Perruchoud AP, Ziesche R, and Block LH (1995) Intracellular interleukin 6 mediates platelet-derived growth factor-induced proliferation of nontransformed cells. *Proc Natl Acad Sci USA* **92**:1312–1316.
- Rouvier E, Luciani MF, Mattei MG, Denizot F, and Golstein P (1993) CTLA-8, cloned from an activated T cell, bearing AU-rich messenger RNA instability sequences and homologous to a herpesvirus saimiri gene. *J Immunol* **150**:5445–5456.
- Ruetten H and Thiemermann C (1997) Interleukin-13 is a more potent inducer of the expression of inducible nitric oxide synthase in smooth muscle cells than in macrophages: a comparison with interleukin-4 and interleukin-10. *Shock* **8**:409–414.
- Rundek T, Elkind MS, Pittman J, Boden-Albala B, Martin S, Humphries SE, Juo SH, and Sacco RL (2002) Carotid intima-media thickness is associated with allelic variants of stromelysin-1, interleukin-6 and hepatic lipase genes: the Northern Manhattan Prospective Cohort Study. *Stroke* **33**:1420–1423.
- Rus HG, Vlaicu R, and Niculescu F (1996) Interleukin-6 and interleukin-8 protein and gene expression in human arterial atherosclerotic wall. *Atherosclerosis* **127**:263–271.
- Russell SM, Johnston JA, Noguchi M, Kawamura M, Bacon CM, Friedmann M, Berg M, McVicar DW, Witthuhn BA, et al. (1994) Interaction of IL-2R beta and gamma c chains with Jak1 and Jak3: implications for XSCID and XCID. *Science (Wash DC)* **266**:1042–1045.
- Russell SM, Keegan AD, Harada N, Nakamura Y, Noguchi M, Leland P, Friedmann MC, Miyajima A, Puri RK, and Paul WE (1993) Interleukin-2 receptor gamma chain: a functional component of the interleukin-4 receptor. *Science (Wash DC)* **262**:1880–1883.
- Rygg M, Uhlir CM, Thorn C, Jensen LE, Gaughan DJ, Varley AW, Munford RS, Goke R, Chen Y, and Whitehead AS (2001) In vitro evaluation of an enhanced human serum amyloid A (SAA2) promoter-regulated soluble TNF receptor fusion protein for anti-inflammatory gene therapy. *Scand J Immunol* **53**:588–595.
- Sack M (2002) Tumor necrosis factor-alpha in cardiovascular biology and the potential role for anti-tumor necrosis factor-alpha therapy in heart disease. *Pharmacol Ther* **94**:123.
- Sakai A, Kume N, Nishi E, Tanoue K, Miyasaka M, and Kita T (1997) P-selectin and vascular cell adhesion molecule-1 are focally expressed in aortas of hypercholesterolemic rabbits before intimal accumulation of macrophages and T lymphocytes. *Arterioscler Thromb Vasc Biol* **17**:310–316.
- Sancho D, Yanez-Mo M, Tejedor R, and Sanchez-Madrid F (1999) Activation of peripheral blood T cells by interaction and migration through endothelium: role of lymphocyte function antigen-1/intercellular adhesion molecule-1 and interleukin-15. *Blood* **93**:886–896.
- Sands BE, Bank S, Sninsky CA, Robinson M, Katz S, Singleton JW, Miner PB, Safdi MA, Galandiuk S, Hanauer SB, et al. (1999) Preliminary evaluation of safety and activity of recombinant human interleukin 11 in patients with active Crohn's disease. *Gastroenterology* **117**:58–64.
- Sasaguri T, Arima N, Tanimoto A, Shimajiri S, Hamada T, and Sasaguri Y (1998) A role for interleukin 4 in production of matrix metalloproteinase 1 by human aortic smooth muscle cells. *Atherosclerosis* **138**:247–253.
- Schieffer B, Schieffer E, Hilfiker-Kleiner D, Hilfiker A, Kovanen PT, Kaartinen M, Nussberger J, Harringer W, and Drexler H (2000) Expression of angiotensin II and interleukin 6 in human coronary atherosclerotic plaques: potential implications for inflammation and plaque instability. *Circulation* **101**:1372–1378.
- Schnaper HW, McGuire J, Runyan C, and Hubchak SC (2000) Sex steroids and the endothelium. *Curr Med Chem* **7**:519–531.
- Schonbeck U, Herzberg M, Petersen A, Wohlenberg C, Gerdes J, Flad HD, and Loppnow H (1997) Human vascular smooth muscle cells express interleukin-1beta-converting enzyme (ICE), but inhibit processing of the interleukin-1beta precursor by ICE. *J Exp Med* **185**:1287–1294.
- Schonbeck U, Sukhova GK, Shimizu K, Mach F, and Libby P (2000) Inhibition of CD40 signaling limits evolution of established atherosclerosis in mice. *Proc Natl Acad Sci USA* **97**:7458–7463.
- Schottelius AJ, Mayo MW, Sartor RB, and Baldwin AS (1999) Interleukin-10 signaling blocks inhibitor of kappaB kinase activity and nuclear factor kappaB DNA binding. *J Biol Chem* **274**:31868–31874.
- Schreyer SA, Peschon JJ, and LeBoeuf RC (1996) Accelerated atherosclerosis in mice lacking tumor necrosis factor receptor p55. *J Biol Chem* **271**:26174–26178.
- Schreyer SA, Vick CM, and LeBoeuf RC (2002) Loss of lymphotoxin-alpha but not tumor necrosis factor-alpha reduces atherosclerosis in mice. *J Biol Chem* **277**:12364–12368.
- Schwenger P, Albert D, Skolnik EY, and Vileck J (1998) Activation of p38 mitogen-activated protein kinase by sodium salicylate leads to inhibition of tumor necrosis factor-induced IkappaB alpha phosphorylation and degradation. *Mol Cell Biol* **18**:78–84.
- Schwertschlag US, Trepicchio WL, Dykstra KH, Keith JC, Turner KJ, and Dorner AJ (1999) Hematopoietic, immunomodulatory and epithelial effects of interleukin-11. *Leukemia* **13**:1307–1315.
- Segal GM, Smith TD, Heinrich MC, Ey FS, and Bagby GC (1992) Specific repression of granulocyte-macrophage and granulocyte colony-stimulating factor gene expression in interleukin-1-stimulated endothelial cells with antisense oligodeoxynucleotides. *Blood* **80**:609–616.
- Seino Y, Ikeda U, Ikeda M, Yamamoto K, Misawa Y, Hasegawa T, Kano S, and Shimada K (1994) Interleukin 6 gene transcripts are expressed in human atherosclerotic lesions. *Cytokine* **6**:87–91.
- Selzman CH, Shames BD, Reznikov LL, Miller SA, Meng X, Barton HA, Werman A, Harken AH, Dinarello CA, and Banerjee A (1999) Liposomal delivery of purified inhibitory-kappaBalpha inhibits tumor necrosis factor-alpha-induced human vascular smooth muscle proliferation. *Circ Res* **84**:867–875.
- Senaldi G, Varnum BC, Sarmiento U, Starnes C, Lile J, Scully S, Guo J, Elliott G, McNinch J, Shaklee CL, et al. (1999) Novel neurotrophin-1/B cell-stimulating factor-3: a cytokine of the IL-6 family. *Proc Natl Acad Sci USA* **96**:11458–11463.
- Sheridan BC, Dinarello CA, Meldrum DR, Fullerton DA, Selzman CH, and McIntyre RC (1999) Interleukin-11 attenuates pulmonary inflammation and vasomotor dysfunction in endotoxin-induced lung injury. *Am J Physiol* **277**:L861–L867.
- Shiozawa S, Shimizu K, Tanaka K, and Hino K (1997) Studies on the contribution of c-fos/AP-1 to arthritic joint destruction. *J Clin Invest* **99**:1210–1216.
- Shreeniwas R, Koga S, Karakorum M, Pinsky D, Kaiser E, Brett J, Wolitzky BA, Norton C, Plocinski J, Benjamin W, Burns DK, Goldstein A, and Stern D (1992) Hypoxia-mediated induction of endothelial cell interleukin-1 alpha. An autocrine mechanism promoting expression of leukocyte adhesion molecules on the vessel surface. *J Clin Invest* **90**:2333–2339.
- Sica A, Matsushima K, Van Damme J, Wang JM, Polentarutti N, Dejana E, Colotta F, and Mantovani A (1990a) IL-1 transcriptionally activates the neutrophil chemotactic factor/IL-8 gene in endothelial cells. *Immunology* **69**:548–553.
- Sica A, Wang JM, Colotta F, Dejana E, Mantovani A, Oppenheim JJ, Larsen CG, Zachariae CO, and Matsushima K (1990b) Monocyte chemotactic and activating factor gene expression induced in endothelial cells by IL-1 and tumor necrosis factor. *J Immunol* **144**:3034–3038.
- Silvestre JS, Mallat Z, Duriez M, Tamarat R, Bureau MF, Scherman D, Duverger N, Branellec D, Tedgui A, and Levy BI (2000) Antiangiogenic effect of interleukin-10 in ischemia-induced angiogenesis in mice hindlimb. *Circ Res* **87**:448–452.
- Simon AD, Yazdani S, Wang WZ, Schwartz A, and Rabbani LE (2001) Elevated plasma levels of interleukin-2 and soluble IL-2 receptor in ischemic heart disease. *Clin Cardiol* **24**:253–256.
- Simonini A, Moscucci M, Muller DWM, Bates ER, Pagani FD, Burdick MD, and Strieter RM (2000) IL-8 is an angiogenic factor in human coronary atherectomy tissue. *Circulation* **101**:1519–1526.
- Sioud M, Opstad A, Hendry P, Lockett TJ, Jennings PA, and McCall MJ (1997) A minimised hammerhead ribozyme with activity against interleukin-2 in human cells. *Biochem Biophys Res Commun* **231**:397–402.
- Sironi M, Breviario F, Proserpio P, Biondi A, Vecchi A, Van Damme J, Dejana E, and Mantovani A (1989) IL-1 stimulates IL-6 production in endothelial cells. *J Immunol* **142**:549–553.
- Slifka MK and Whitton JL (2000) Clinical implications of dysregulated cytokine production. *J Mol Med* **78**:74–80.
- Smith MF (2000) Interleukin-1 receptor antagonist, in *Novel Cytokine Inhibitors* (Higgs GA and Henderson B eds) pp 177–199, Birkhäuser Verlag AG, Basel.
- Smith DA, Irving SD, Sheldon J, Cole D, and Kaski JC (2001) Serum levels of the antiinflammatory cytokine interleukin-10 are decreased in patients with unstable angina. *Circulation* **104**:746–749.
- Song L, Leung C, and Schindler C (2001) Lymphocytes are important in early atherosclerosis. *J Clin Invest* **108**:251–259.
- Song S, Ling Hu H, Roebuck KA, Rabbi MF, Donnelly RP, and Finnegan A (1997) Interleukin-10 inhibits interferon-gamma-induced intercellular adhesion molecule-1 gene transcription in human monocytes. *Blood* **89**:4461–4469.
- Spiecker M, Peng HB, and Liao JK (1997) Inhibition of endothelial vascular cell adhesion molecule-1 expression by nitric oxide involves the induction and nuclear translocation of IkappaBalpha. *J Biol Chem* **272**:30969–30974.
- Spriggs MK (1996) One step ahead of the game: viral immunomodulatory molecules. *Annu Rev Immunol* **14**:101–130.
- Spriggs MK, Hruby DE, Maliszewski CR, Pickup DJ, Sims JE, Buller RM, and VanSlyke J (1992) Vaccinia and cowpox viruses encode a novel secreted interleukin-1-binding protein. *Cell* **71**:145–152.
- Stanford SJ, Pepper JR, and Mitchell JA (2000) Cyclooxygenase-2 regulates granulocyte-macrophage colony-stimulating factor, but not interleukin-8, production by human vascular cells—role of cAMP. *Arterioscler Thromb Vasc Biol* **20**:677–682.
- Starnes T, Robertson MJ, Sledge G, Kelich S, Nakshatri H, Broxmeyer HE, and Hromas R (2001) Cutting edge: IL-17F, a novel cytokine selectively expressed in activated T cells and monocytes, regulates angiogenesis and endothelial cell cytokine production. *J Immunol* **167**:4137–4140.
- Starr R, Willson TA, Viney EM, Murray LJ, Rayner JR, Jenkins BJ, Gonda TJ, Alexander WS, Metcalf D, Nicola NA, and Hilton DJ (1997) A family of cytokine-inducible inhibitors of signalling. *Nature (Lond)* **387**:917–921.
- Stary HC, Chandler AB, Dinsmore RE, Fuster V, Glagov S, Insull W, Rosenfeld ME, Schwartz CJ, Wagner WD, and Wissler RW (1995) A definition of advanced types of atherosclerotic lesions and a histological classification of atherosclerosis. A report from the Committee on Vascular Lesions of the Council on Arteriosclerosis, American Heart Association. *Circulation* **92**:1355–1374.
- Stary HC, Chandler AB, Glagov S, Guyton JR, Insull W, Rosenfeld ME, Schaffer SA,



- Schwartz CJ, Wagner WD, and Wissler RW (1994) A definition of initial, fatty streak and intermediate lesions of atherosclerosis. A report from the Committee on Vascular Lesions of the Council on Arteriosclerosis, American Heart Association. *Circulation* **89**:2462–2478.
- Stassen M, Arnold M, Hultner L, Muller C, Neudorfl C, Reineke T, and Schmitt E (2000) Murine bone marrow-derived mast cells as potent producers of IL-9: costimulatory function of IL-10 and kit ligand in the presence of IL-1. *J Immunol* **164**:5549–5555.
- Steidl U, Haas R, and Kronenwett R (2000) Interleukin-10 on monocytes mediates adhesion as well as trans-endothelial migration and can be down-regulated using antisense oligonucleotides. *Ann Hematol* **79**:414–423.
- Stein B and Kung Sutherland MS (1998) IL-6 as a drug discovery target. *Drug Discov Today* **3**:202–213.
- Stenvinkel P, Heimbürger O, and Jøgestrand T (2002) Elevated interleukin-6 predicts progressive carotid artery atherosclerosis in dialysis patients: association with Chlamydia pneumoniae seropositivity. *Am J Kidney Dis* **39**:274–282.
- Stepkowski SM, Tu Y, Condon TP, and Bennett CF (1994) Blocking of heart allograft rejection by intercellular adhesion molecule-1 antisense oligonucleotides alone or in combination with other immunosuppressive modalities. *J Immunol* **153**:5336–5346.
- Su JZ, Fukuda N, Hu WY, and Kanmatsuse K (2000) Ribozyme to human TGF-beta 1 mRNA inhibits the proliferation of human vascular smooth muscle cells. *Biochem Biophys Res Commun* **278**:401–407.
- Sukhova GK, Schonbeck U, Rabkin E, Schoen FJ, Poole AR, Billinghurst RC, and Libby P (1999) Evidence for increased collagenolysis by interstitial collagenases-1 and -3 in vulnerable human atherosclerotic plaques. *Circulation* **99**:2503–2509.
- Sukovich DA, Kausar K, Shirley FD, DelVecchio V, Halks-Miller M, and Rubanyi GM (1998) Expression of interleukin-6 in atherosclerotic lesions of male ApoE-knockout mice: inhibition by 17beta-estradiol. *Arterioscler Thromb Vasc Biol* **18**:1498–1505.
- Sun RX, Gennaro C, Rocco S, Gu ZJ, and Klein B (1997) Interleukin-6 receptor antagonists inhibit interleukin-11 biological activity. *Eur Cytokine Netw* **8**:51–56.
- Suzuki H, Shibano K, Okane M, Kono I, Matsui Y, Yamane K, and Kashiwagi H (1989) Interferon-gamma modulates messenger RNA levels of c-sis (PDGF-B chain), PDGF-A chain and IL-1 beta genes in human vascular endothelial cells. *Am J Pathol* **134**:35–43.
- Svenson M, Hansen MB, Kayser L, Rasmussen AK, Reimert CM, and Bendtzen K (1992) Effects of human anti-IL-1 alpha autoantibodies on receptor binding and biological activities of IL-1. *Cytokine* **4**:125–133.
- Svenson M, Hansen MB, Thomsen AR, Diamant M, Nansen A, Rieneck K, Otterness IG, and Bendtzen K (2000) Cytokine vaccination: neutralising IL-1alpha autoantibodies induced by immunisation with homologous IL-1alpha. *J Immunol Methods* **236**:1–8.
- Takagi N, Mihara M, Moriya Y, Nishimoto N, Yoshizaki K, Kishimoto T, Takeda Y, and Ohsugi Y (1998) Blockage of interleukin-6 receptor ameliorates joint disease in murine collagen-induced arthritis. *Arthritis Rheum* **41**:2117–2121.
- Takeda T, Nakajima K, Kojima H, and Hirano T (1994) E1A repression of IL-6-induced gene activation by blocking the assembly of IL-6 response element binding complexes. *J Immunol* **153**:4573–4582.
- Takeshita S, Isshiki T, Ochiai M, Ishikawa T, Nishiyama Y, Fusano T, Toyozumi H, Kondo K, Ono Y, and Sato T (1997) Systemic inflammatory responses in acute coronary syndrome: increased activity observed in polymorphonuclear leukocytes but not T lymphocytes. *Atherosclerosis* **135**:187–192.
- Takeshita T, Asao H, Suzuki J, and Sugamura K (1990) An associated molecule, p64, with high-affinity interleukin 2 receptor. *Int Immunol* **2**:477–480.
- Taki H, Sakai T, Sugiyama E, Mino T, Kuroda A, Taki K, Hamazaki T, Koizumi H, and Kobayashi M (1999) Monokine stimulation of interleukin-11 production by human vascular smooth muscle cells in vitro. *Atherosclerosis* **144**:375–380.
- Tedgui A and Mallat Z (2001) Anti-inflammatory mechanisms in the vascular wall. *Circ Res* **88**:877–887.
- Tengku-Muhammad TS, Hughes TR, Cryer A, and Ramji DP (1996) Differential regulation of lipoprotein lipase in the macrophage J774.2 cell line by cytokines. *Cytokine* **8**:525–533.
- Terkeltaub RA (1999) IL-10: An “immunologic scalpel” for atherosclerosis? *Arterioscler Thromb Vasc Biol* **19**:2823–2825.
- Terkeltaub R, Banka CL, Solan J, Santoro D, Brand K, and Curtiss LK (1994) Oxidized LDL induces monocyte cell expression of interleukin-8, a chemokine with T-lymphocyte chemotactic activity. *Arterioscler Thromb* **14**:47–53.
- Tipping PG and Hancock WW (1993) Production of tumor necrosis factor and interleukin-1 by macrophages from human atherosclerotic plaques. *Am J Pathol* **142**:1721–1728.
- Toda K, Kayano K, Karimova A, Naka Y, Fujita T, Minamoto K, Wang CY, and Pinsky DJ (2000) Antisense intercellular adhesion molecule-1 (ICAM-1) oligodeoxynucleotide delivered during organ preservation inhibits posttransplant ICAM-1 expression and reduces primary lung isograft failure. *Circ Res* **86**:166–174.
- Tominaga A, Takaki S, Koyama N, Katoh S, Matsumoto R, Migita M, Hitoshi Y, Hosoya Y, Yamauchi S, Kanai Y, et al. (1991) Transgenic mice expressing a B cell growth and differentiation factor gene (interleukin 5) develop eosinophilia and autoantibody production. *J Exp Med* **173**:429–437.
- Tone M, Thompson SA, Tone Y, Fairchild PJ, and Waldmann H (1997) Regulation of IL-18 (IFN-gamma-inducing factor) gene expression. *J Immunol* **159**:6156–6163.
- Torigoe K, Ushio S, Okura T, Kobayashi S, Taniai M, Kunikata T, Murakami T, Sanou O, Kojima H, Fujii M, et al. (1997) Purification and characterization of the human interleukin-18 receptor. *J Biol Chem* **272**:25737–25742.
- Touw IP, De Koning JP, Ward AC, and Hermans MH (2000) Signaling mechanisms of cytokine receptors and their perturbances in disease. *Mol Cell Endocrinol* **160**:1–9.
- Trepicchio WL, Bozza M, Pedneault G, and Dorner AJ (1996) Recombinant human IL-11 attenuates the inflammatory response through down-regulation of proinflammatory cytokine release and nitric oxide production. *J Immunol* **157**:3627–3634.
- Trepicchio WL, Ozawa M, Walters IB, Kikuchi T, Gilleaudeau P, Bliss JL, Schwertschlag U, Dorner AJ, and Krueger JG (1999) Interleukin-11 therapy selectively down-regulates type I cytokine proinflammatory pathways in psoriasis lesions. *J Clin Invest* **104**:1527–1537.
- Trepicchio WL, Wang L, Bozza M, and Dorner AJ (1997) IL-11 regulates macrophage effector function through the inhibition of nuclear factor-kappaB. *J Immunol* **159**:5661–5670.
- Ueda K, Takahashi M, Ozawa K, and Kinoshita M (1999) Decreased soluble interleukin-6 receptor in patients with acute myocardial infarction. *Am Heart J* **138**:908–915.
- Upton C, Mossman K, and McFadden G (1992) Encoding of a homolog of the IFN-gamma receptor by myxoma virus. *Science (Wash DC)* **258**:1369–1372.
- Ushio S, Namba M, Okura T, Hattori K, Nukada Y, Akita K, Tanabe F, Konishi K, Micallef M, Fujii M, et al. (1996) Cloning of the cDNA for human IFN-gamma-inducing factor, expression in *Escherichia coli* and studies on the biologic activities of the protein. *J Immunol* **156**:4274–4279.
- Uyemura K, Demer LL, Castle SC, Jullien D, Berliner JA, Gately MK, Warriar RR, Pham N, Fogelman AM, and Modlin RL (1996) Cross-regulatory roles of interleukin (IL)-12 and IL-10 in atherosclerosis. *J Clin Invest* **97**:2130–2138.
- Uyttenhove C, Simpson RJ, and Van Snick J (1988) Functional and structural characterization of P40, a mouse glycoprotein with T-cell growth factor activity. *Proc Natl Acad Sci USA* **85**:6934–6938.
- Vadiveloo PK, Stanton HR, Cochran FW, and Hamilton JA (1994) Interleukin-4 inhibits human smooth muscle cell proliferation. *Artery* **21**:161–181.
- van der Meer IM, de Maat MP, Bots ML, Breteler MM, Meijer J, Kiliaan AJ, Hofman A, and Witteman JC (2002) Inflammatory mediators and cell adhesion molecules as indicators of severity of atherosclerosis: the Rotterdam Study. *Arterioscler Thromb Vasc Biol* **22**:838–842.
- van Deventer SJ, Elson CO, and Fedorak RN (1997) Multiple doses of intravenous interleukin 10 in steroid-refractory Crohn's disease. Crohn's Disease Study Group. *Gastroenterology* **113**:383–389.
- van Leeuwen BH, Martinson ME, Webb GC, and Young IG (1989) Molecular organization of the cytokine gene cluster, involving the human IL-3, IL-4, IL-5 and GM-CSF genes, on human chromosome 5. *Blood* **73**:1142–1148.
- van Lenten BJ and Fogelman AM (1992) Lipopolysaccharide-induced inhibition of scavenger receptor expression in human monocyte-macrophages is mediated through tumor necrosis factor-alpha. *J Immunol* **148**:112–116.
- Verma S, Li SH, Badiwala MV, Weisel RD, Fedak PW, Li RK, Dhillion B, and Mickle DA (2002) Endothelin antagonism and interleukin-6 inhibition attenuate the proatherogenic effects of C-reactive protein. *Circulation* **105**:1890–1896.
- Vidal-Vanaclocha F, Fantuzzi G, Mendoza L, Fuentes AM, Anasagasti MJ, Martin J, Carrascal T, Walsh P, Reznikov LL, Kim SH, et al. (2000) IL-18 regulates IL-1 beta-dependent hepatic melanoma metastasis via vascular cell adhesion molecule-1. *Proc Natl Acad Sci USA* **97**:734–739.
- Vincenti F, Kirkan M, Light S, Bumgardner G, Pescovitz M, Halloran P, Neylan J, Willkinson A, Ekberg H, Gaston R, et al. (1998) Interleukin-2-receptor blockade with daclizumab to prevent acute rejection in renal transplantation Daclizumab Triple Therapy Study Group. *N Engl J Med* **338**:161–165.
- Volk HD, Asadullah K, Gallagher G, Sabat R, and Grütz G (2001) IL-10 and its homologs: important immune mediators and emerging immunotherapeutic agents. *Trends Immunol* **22**:414–417.
- Volpato S, Guralnik JM, Ferrucci L, Balfour J, Chaves P, Fried LP, and Harris TB (2001) Cardiovascular disease, interleukin-6 and risk of mortality in older women—the women's health and aging study. *Circulation* **103**:947–953.
- von der Thüsen JH, Kuiper J, Pekkes ML, de Vos P, van Berkel ThJC, and Biessen EAD (2001a) Attenuation of atherogenesis by systemic and local adenovirus-mediated gene transfer of interleukin-10 in LDLr<sup>-/-</sup> Mice. *FASEB J* **15**:2730–2732.
- von der Thüsen JH, van Berkel ThJC, and Biessen EAD (2001b) Induction of Rapid Atherogenesis by Perivascular Carotid Collar Placement in ApoE<sup>-/-</sup> and LDLr<sup>-/-</sup> Mice. *Circulation* **103**:1164–1170.
- Vosshenrich CAJ and Di Santo JP (2001) Cytokines: IL-21 joins the gamma(c)-dependent network? *Curr Biol* **11**:R175–R177.
- Waldmann TA and Tagaya Y (1999) The multifaceted regulation of interleukin-15 expression and the role of this cytokine in NK cell differentiation and host response to intracellular pathogens. *Annu Rev Immunol* **17**:1719–1749.
- Walmsley M, Butler DM, Marinova-Mutafchieva L, and Feldmann M (1998) An anti-inflammatory role for interleukin-11 in established murine collagen-induced arthritis. *Immunology* **95**:31–37.
- Wang CY, Mazer SP, Minamoto K, Takuma S, Homma S, Yellin M, Chess L, Fard A, Kalled SL, Oz MC, and Pinsky DJ (2002) Suppression of murine cardiac allograft arteriopathy by long-term blockade of CD40-CD154 interactions. *Circulation* **105**:1609–1614.
- Wang JM, Sica A, Peri G, Walter S, Padura IM, Libby P, Ceska M, Lindley I, Colotta F, and Mantovani A (1991) Expression of monocyte chemotactic protein and interleukin-8 by cytokine-activated human vascular smooth muscle cells. *Arterioscler Thromb* **11**:1166–1174.
- Wang LH, Yang XY, Kirken RA, Resau JH, and Farrar WL (2000) Targeted disruption of stat6 DNA binding activity by an oligonucleotide decoy blocks IL-4-driven TH2 cell response. *Blood* **95**:1249–1257.
- Wang N, Tabas I, Winchester R, Ravalli S, Rabbani LE, and Tall A (1996) Interleukin 8 is induced by cholesterol loading of macrophages and expressed by macrophage foam cells in human atheroma. *J Biol Chem* **271**:8837–8842.
- Wang X, Feuerstein GZ, Gu JL, Lysko PG, and Yue (1995) TL Interleukin-1 beta induces expression of adhesion molecules in human vascular smooth muscle cells and enhances adhesion of leukocytes to smooth muscle cells. *Atherosclerosis* **115**:89–98.
- Weber KS, Draude G, Erl W, de Martin R, and Weber C (1999) Monocyte arrest and

- transmigration on inflamed endothelium in shear flow is inhibited by adenovirus-mediated gene transfer of IkappaB-alpha. *Blood* **93**:3685-3693.
- Wendling D, Racadot E, and Wijdenes J (1993) Treatment of severe rheumatoid arthritis by anti-interleukin 6 monoclonal antibody. *J Rheumatol* **20**:259-262.
- Wesemann DR, Dong Y, O'Keefe GM, Nguyen VT, and Benveniste EN (2002) Suppressor of cytokine signaling 1 inhibits cytokine induction of CD40 expression in macrophages. *J Immunol* **169**:2354-2360.
- Whitman SC, Ravisankar P, and Daugherty A (2002) Interleukin-18 enhances atherosclerosis in apolipoprotein E-/- mice through release of interferon-gamma. *Circ Res* **90**:e34-e38.
- Wildbaum G, Westermann J, Maor G, and Karin N (2000) A targeted DNA vaccine encoding fas ligand defines its dual role in the regulation of experimental autoimmune encephalomyelitis. *J Clin Invest* **106**:671-679.
- Williams JK, Sukhova GK, Herrington DM, and Libby P (1998) Pravastatin has cholesterol-lowering independent effects on the artery wall of atherosclerotic monkeys. *J Am Coll Cardiol* **31**:684-691.
- Wilson KP, Black JA, Thomson JA, Kim EE, Griffith JP, Navia MA, Murcko MA, Chambers SP, Aldape RA, Raybuck SA, and Livingston DJ (1994) Structure and mechanism of interleukin-1 beta converting enzyme. *Nature (Lond)* **370**:270-275.
- Wilson SH, Best PJ, Edwards WD, Holmes DR Jr, Carlson PJ, Celermajer DS, and Lerman A (2002) Nuclear factor-kappaB immunoreactivity is present in human coronary plaque and enhanced in patients with unstable angina pectoris. *Atherosclerosis* **160**:147-153.
- Wolk K, Kunz S, Asadullah K, and Sabat R (2002) Cutting edge: immune cells as sources and targets of the IL-10 family members? *J Immunol* **168**:5397-5402.
- Wrighton CJ, Hofer-Warbinek R, Moll T, Eytner R, Bach FH, and de Martin R (1996) Inhibition of endothelial cell activation by adenovirus-mediated expression of I kappa B alpha, an inhibitor of the transcription factor NF-kappa B. *J Exp Med* **183**:1013-1022.
- Wuttge DM, Eriksson P, Sirsjo A, Hansson GK, and Stemme S (2001) Expression of interleukin-15 in mouse and human atherosclerotic lesions. *Am J Pathol* **159**:417-423.
- Xing Z, Gauldie J, Cox G, Baumann H, Jordana M, Lei XF, and Achong MK (1998) IL-6 is an antiinflammatory cytokine required for controlling local or systemic acute inflammatory responses. *J Clin Invest* **101**:311-320.
- Xu D, Chan WL, Leung BP, Hunter D, Schulz K, Carter RW, McInnes IB, Robinson JH, and Liew FY (1998a) Selective expression and functions of interleukin 18 receptor on T helper (Th) type 1 but not Th2 cells. *J Exp Med* **188**:1485-1492.
- Xu WF, Andersen H, Whitmore TE, Presnell SR, Yee DP, Ching A, Gilbert T, Davie EW, and Foster DC (1998b) Cloning and characterization of human protease-activated receptor 4. *Proc Natl Acad Sci USA* **95**:6642-6646.
- Yacyshyn BR, Bowen-Yacyshyn MB, Jewell L, Tami JA, Bennett CF, Kisner DL, and Shanahan WR (1998) A placebo-controlled trial of ICAM-1 antisense oligonucleotide in the treatment of Crohn's disease. *Gastroenterology* **114**:1133-1142.
- Yahata N, Kawai S, Higaki M, and Mizushima Y (1996) Antisense phosphorothioate oligonucleotide inhibits interleukin 1 beta production in the human macrophage-like cell line, U937. *Antisense Nucleic Acid Drug Dev* **6**:55-61.
- Yamamoto K, Morishita R, Tomita N, Shimozato T, Nakagami H, Kikuchi A, Aoki M, Higaki J, Kaneda Y, and Ogihara T (2000) Ribozyme oligonucleotides against transforming growth factor-beta inhibited neointimal formation after vascular injury in rat model—potential application of ribozyme strategy to treat cardiovascular disease. *Circulation* **102**:1308-1314.
- Yang GY, Zhao YJ, Davidson BL, and Betz AL (1997) Overexpression of interleukin-1 receptor antagonist in the mouse brain reduces ischemic brain injury. *Brain Res* **751**:181-188.
- Yao Z, Fanslow WC, Seldin MF, Rousseau AM, Painter SL, Comeau MR, Cohen JI, and Spriggs MK (1995a) Herpesvirus Saimiri encodes a new cytokine, IL-17, which binds to a novel cytokine receptor. *Immunity* **3**:811-821.
- Yao Z, Painter SL, Fanslow WC, Ulrich D, Macduff BM, Spriggs MK, and Armitage RJ (1995b) Human IL-17: a novel cytokine derived from T cells. *J Immunol* **155**:5483-5486.
- Yin T, Taga T, Tsang ML, Yasukawa K, Kishimoto T, and Yang YC (1993) Involvement of IL-6 signal transducer gp130 in IL-11-mediated signal transduction. *J Immunol* **151**:2555-2561.
- Ying S, Meng Q, Barata LT, Robinson DS, Durham SR, and Kay AB (1997) Associations between IL-13 and IL-4 (mRNA and protein), vascular cell adhesion molecule-1 expression, and the infiltration of eosinophils, macrophages and T cells in allergen-induced late-phase cutaneous reactions in atopic subjects. *J Immunol* **158**:5050-5057.
- Yoshida K, Taga T, Saito M, Suematsu S, Kumanogoh A, Tanaka T, Fujiwara H, Hirata M, Yamagami T, Nakahata T, Hirabayashi T, Yoneda Y, Tanaka K, Wang WZ, Mori C, Shiota K, Yoshida N, and Kishimoto T (1996) Targeted disruption of gp130, a common signal transducer for the interleukin 6 family of cytokines, leads to myocardial and hematological disorders. *Proc Natl Acad Sci USA* **93**:407-411.
- Young JL, Sukhova GK, Foster D, Kiesel W, Libby P, and Schonbeck U (2000) The serpin proteinase inhibitor 9 is an endogenous inhibitor of interleukin 1 beta-converting enzyme (caspase-1) activity in human vascular smooth muscle cells. *J Exp Med* **191**:1535-1544.
- Young PR (1998) Pharmacological modulation of cytokine action and production through signaling pathways. *Cytokine Growth Factor Rev* **9**:239-257.
- Youssef S, Wildbaum G, Maor G, Lanir N, Gour-Lavie A, Grabie N, and Karin N (1998) Long-lasting protective immunity to experimental autoimmune encephalomyelitis following vaccination with naked DNA encoding C-C chemokines. *J Immunol* **161**:3870-3879.
- Yudkin JS, Kumari M, Humphries SE, and Mohamed-Ali V (2000) Inflammation, obesity, stress and coronary heart disease: is interleukin-6 the link? *Atherosclerosis* **148**:209-214.
- Yue TL, Wang X, Sung CP, Olson B, McKenna PJ, Gu JL, and Feuerstein GZ (1994) Interleukin-8. A mitogen and chemoattractant for vascular smooth muscle cells. *Circ Res* **75**:1-7.
- Zan H, Cerutti A, Dramitinos P, Schaffer A, and Casali P (1998) CD40 engagement triggers switching to IgA1 and IgA2 in human B cells through induction of endogenous TGF-beta: evidence for TGF-beta but not IL-10-dependent direct S mu->S alpha and sequential S mu->S gamma, S gamma->S alpha DNA recombination. *J Immunol* **161**:5217-5225.
- Zaug AJ, Been MD, and Cech TR (1986) The Tetrahymena ribozyme acts like an RNA restriction endonuclease. *Nature (Lond)* **324**:429-433.
- Zhang JG, Hilton DJ, Willson TA, McFarlane C, Roberts BA, Moritz RL, Simpson RJ, Alexander WS, Metcalf D, and Nicola NA (1997) Identification, purification and characterization of a soluble interleukin (IL)-13-binding protein. Evidence that it is distinct from the cloned IL-13 receptor and IL-4 receptor alpha-chains. *J Biol Chem* **272**:9474-9480.
- Zhao L, Cuff CA, Moss E, Wille U, Cyrus T, Klein EA, Pratico D, Rader DJ, Hunter CA, Pure E, and Funk CD (2002) Selective interleukin-12 synthesis defect in 12/15-lipoxygenase-deficient macrophages associated with reduced atherosclerosis in a mouse model of familial hypercholesterolemia. *J Biol Chem* **277**:35350-35356.
- Zhou RH, Shi Q, Gao HQ, and Shen BJ (2001a) Changes in serum interleukin-8 and interleukin-12 levels in patients with ischemic heart disease in a Chinese population. *J Atheroscler Thromb* **8**:30-32.
- Zhou YH, McLane M, and Levitt RC (2001b) Th2 cytokines and asthma—interleukin-9 as a therapeutic target for asthma. *Resp Res* **2**:80-84.
- Zimmerman MA, Selzman CH, Reznikov LL, Raeburn CD, Barsness K, McIntyre RC Jr, Hamiel CR, and Harken AH (2002) Interleukin-11 attenuates human vascular smooth muscle cell proliferation. *Am J Physiol Heart Circ Physiol* **283**:H175-H180.
- Zorn U, Dallmann I, Grosse J, Kirchner H, Poliwoda H, and Atzpodien J (1994) Soluble interleukin 2 receptors abrogate IL-2 induced activation of peripheral mononuclear cells. *Cytokine* **6**:358-364.
- Zurawski G and de Vries JE (1994) Interleukin 13, an interleukin 4-like cytokine that acts on monocytes and B cells, but not on T cells. *Immunol Today* **15**:19-26.
- Zurawski SM, Vega F, Huyghe B, and Zurawski G (1993) Receptors for interleukin-13 and interleukin-4 are complex and share a novel component that functions in signal transduction. *EMBO J* **12**:2663-2670.